

Taxonomic notes about some Indo-Australian Lamiini, especially for the genus *Acalolepta* Pascoe, 1858 (Coleoptera, Cerambycidae)

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Abstract

Orsidis cariosus Pascoe, 1866 and *Trachystola puncticollis* Fisher, 1935 (currently, both in *Acalolepta*) are transferred to *Granulorsidis* Breuning, 1980, resulting in *G. cariosus* (Pascoe, 1866) n. comb. and *G. puncticollis* (Fisher, 1935) n. comb. *Sternohammus femoralis* (Aurivillius, 1927) is considered to be a senior synonym of *S. femoraloides* Breuning, 1980 n. syn. *Dihammus submaculatus* Gilmour, 1947 is considered to be a senior synonym of *Sternohammus sanghiricus* Breuning, 1956 n. syn. and *S. celebensis submaculatus* (Gilmour, 1947) n. comb., n. status is proposed. *Acalolepta (Pilohammus)* n. subgen. (type species: *Monohammus mixtus* Hope, 1841) is proposed for *A. mixta* (Hope, 1841) and *A. ampliata* (Gahan, 1888). *A. (Dihammus) producta* (Pascoe, 1866) is recognised as a senior synonym of *A. buruana* Breuning, 1970 n. syn. The holotype of *A. holotephra* (Boisduval, 1835) is considered invalid due to its labels and the original picture is proposed as lectotype. *A. strandiella websteri* Breuning, 1970 and *A. densemarmorata* Breuning, 1970 are considered to be possible junior synonyms of this species. *A. (Dihammus) ceramensis* n. sp. from Ceram (Indonesia: Moluccas) is described.

Résumé

Orsidis cariosus Pascoe, 1866 et *Trachystola puncticollis* Fisher, 1935 (deux espèces appartenant au genre *Acalolepta*) sont transférés dans le genre *Granulorsidis* Breuning, 1980 : *G. cariosus* (Pascoe, 1866) n. comb. et *G. puncticollis* (Fisher, 1935) n. comb. *Sternohammus femoralis* (Aurivillius, 1927) est reconnu comme senior synonyme de *S. femoraloides* Breuning, 1980 n. syn. *Dihammus submaculatus* Gilmour, 1947 est considéré comme senior synonyme de *Sternohammus sanghiricus* Breuning, 1956 n. syn. et *S. celebensis submaculatus* (Gilmour, 1947) n. comb., n. status est proposé. *Acalolepta (Pilohammus)* n. subgen. (génotype: *Monohammus mixtus* Hope, 1841) est proposé pour *A. mixta* (Hope, 1841) et *A. ampliata* (Gahan, 1888). *A. (Dihammus) producta* (Pascoe, 1866) est reconnu comme senior synonyme de *A. buruana* Breuning, 1970 n. syn. L'holotype d'*A. holotephra* (Boisduval, 1835) est reconnu invalide à cause de ses étiquettes et le dessin original est proposé comme lectotype. *A. strandiella websteri* Breuning, 1970 and *A. densemarmorata* Breuning, 1970 sont considérées comme possibles junior synonymes de cette espèce. *A. (Dihammus) ceramensis* n. sp. du Céram (Indonésie: Moluques) est décrit.

Key-words

Coleoptera, Lamiinae, Philippines, Moluccas, taxonomy.

Introduction

The genus *Acalolepta* Pascoe, 1858 currently includes almost 300 valid taxa, many with taxonomic positions that still need to be verified. In fact, nineteenth century pioneers of entomology furnished descriptions characterised by restrictive diagnostic characters, and subsequently, Breuning produced an enormous number of descriptions using subjective, relative, or even equivocal characters. In both cases, the lack of illustrations and considerations of biogeography and paleogeography were other factors of confusion. The result has been that the same taxon was sometimes described several times or some subspecies were attributed to species having a completely uncorrelated distribution.

In this paper, some taxonomic problems related to the genus *Acalolepta* have been addressed, and solutions proposed, that involve the related genera *Granulorsidis* Breuning, 1980 and *Sternohammus* Breuning, 1935.

Materials and methods

Materials coming from the following collections were examined:

BMNH: British Museum of Natural History, London (United Kingdom).

CAS: Collection Andre Skale, Hof / Saale (Germany).

CDH: Collection Daniel Heffern, Houston, Texas (USA).

CDT: Collection Dimitry Telnov, Riga (Latvia).

CFV: Collection Francesco Vitali, Luxembourg (Luxembourg).

CGC: Collection Gérard Chemin, Champigny, (France).

CJPR: Collection Jean-Philippe Roguet, Nogent-sur-Marne (France).

CRV: Robert Vigneault, Maddington (Canada).

CVN: Collection Vitali Nagirnyi, Tartu (Estonia).

MNHNP: National Museum of Natural History, Paris (France).

USNM: National Museum of Natural History, Washington, D. C. (USA).

Photographs of the specimens preserved in the CFV were taken by the author with a Canon Power Shot SX10IS camera in natural light (*Acalolepta ceramensis* n. sp. holotype) or with a CMOS Camera mounted on a Keyence VHX 6000 digital microscope equipped with a VHX-S660E free-angle observation system, a VH-ZST 20-2000x double zoom objective, 2D/3D image stitching system and stacking system taking 200 images at 2 million pixels of resolution, owned by the National Museum of Natural History of Luxembourg. Those of the specimens preserved in the MNHNP and in CGC were taken by Gérard Chemin with an Olympus E-300 camera with an objective Zuiko Digital 35 mm lens. All photographs in this work were enhanced with the Ulead PhotoImpact SE software.

Results

Granulorsidis flavidosignatus (Aurivillius, 1927) (Fig. 1a-b)

Orsidis flavidosignatus Aurivillius, 1927: 554 (Philippines: Mindanao) or. comb.

Dihammus flavidosignatus BREUNING, 1944: 464.

Acalolepta flavidosignata Breuning, 1961: 372; Lingafelter et al., 2014: 138, Fig. 67 w-x.

Granulorsidis granulipennis Breuning, 1980: 172 (Philippines: Mindanao); CHEMIN & VIVES, 2017: 14, Fig. 38.

Granulorsidis flavidosignatus VIVES, 2012: 80.

Examined material. – SYNTYPE, 1 ex., Butuan / Mindanao / [C. F.] Baker (printed on a white label) // 36 (printed on a white label) // *Orsidis / flavidosignatus / Auriv.[illis] [19]27 Auriv.[illius]* (handwritten on a white label) // Typus (printed on a red label) // BLNO / 000750 (printed on a sky-blue label), in USNM; HOLOTYPE, 1♀, *Granulorsidis / granulipennis* / Breuning det. Typ. (handwritten by Breuning and printed on a yellowish label) // 193 (handwritten on a yellowish label) // Type (printed on a red label) // *Granulorsidis / granulipennis* nov. (typed on a yellowish label) // Holotype (printed on a red label) // Muséum / Paris (printed on a white label), in MNHNP; 1♂, Philippines, Mindanao, Lanao del Sur, Wao, X-2017, I. Lumawig leg., in CFV; 1♀, ditto, Kabangasan Bukidnon, IX-2014, I. Lumawig leg., in CFV.

Granulorsidis cariosus (Pascoe, 1866) n. comb. (Fig. 2a-b)

Orsidis cariosus Pascoe, 1866: 309 (Singapore), AURIVILLIUS, 1921: 99. or. comb.

Cypriola cariosa GEMMINGER & HAROLD, 1873: 3034.

Dihammus cariosus BREUNING, 1944: 465.

Acalolepta cariosa BREUNING, 1961: 372.

Examined material. – *Orsidis cariosus* Pascoe, 1866: HOLOTYPE, 1♂, Singapore, in BMNH; 1♀, Malaysia, Borneo, Sabah, Kuamut, 500 m, 4-IV-2014, loc. coll., donum D. Heffern, in CFV; 1♀, Indonesia, Borneo, Kalimantan Mt. Bawang, in CGC.

Granulorsidis puncticollis (Fisher, 1935) n. comb. (Fig. 3)

Trachystola puncticollis Fisher, 1935: 596 (Malaysia, Borneo: Mt. Kinabalu); BREUNING, 1943b: 187 or. comb.

Dihammus pustulatus Breuning, 1940: 412 (Malaysia, Borneo: Mt. Tibang); BREUNING, 1944: 465.

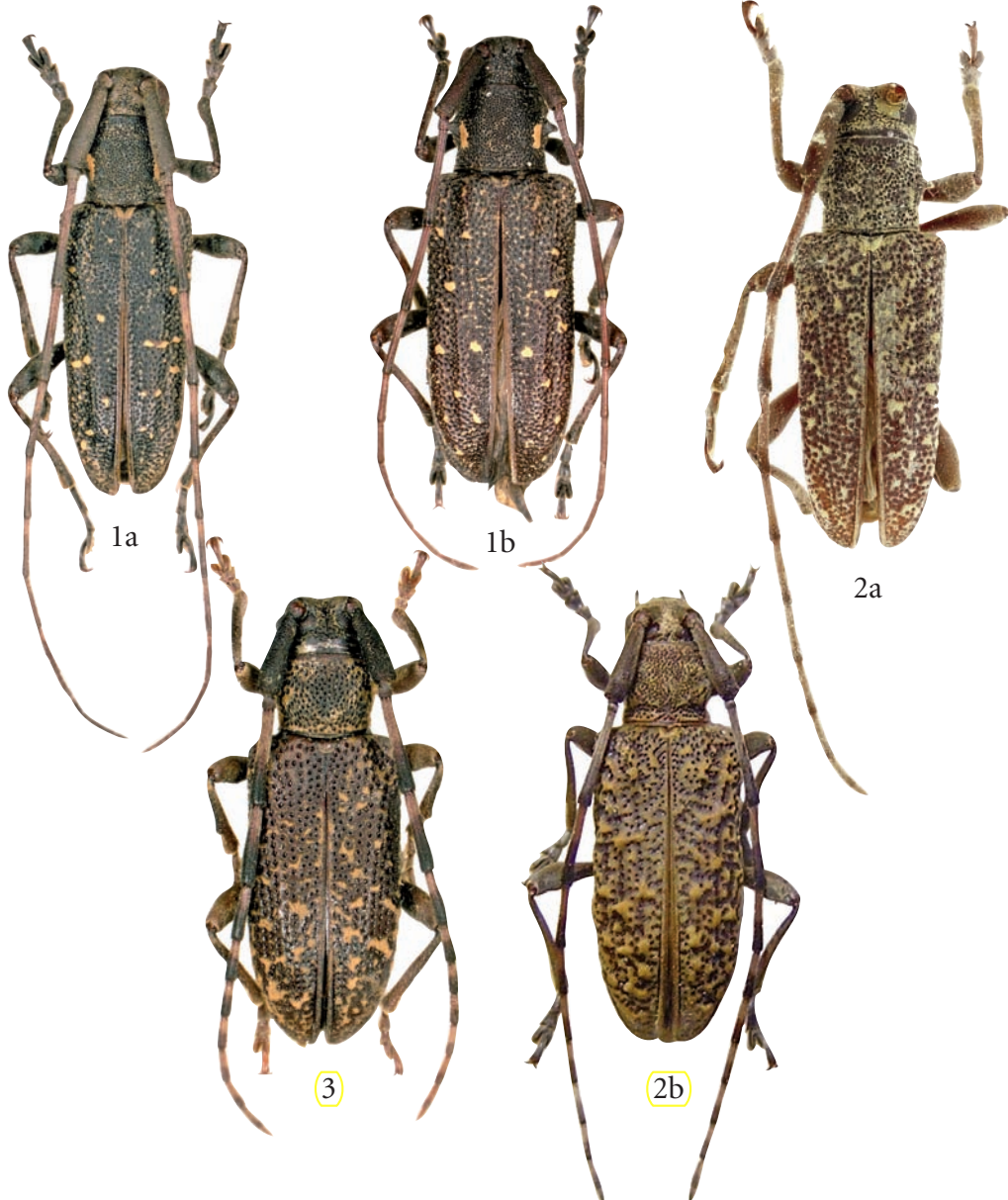
Cypriola puncticollis BREUNING, 1960: 35.

Acalolepta puncticollis BREUNING, 1961: 372; LINGAFELTER et al., 2014: 42, Fig. 139w-x.

Examined material. – *Trachystola puncticollis*: HOLOTYPE, 1♀, B.N. Borneo / Mt. Kinabalu, / Lumu Lumu, / 5.500 ft. / April 7 1929 (handwritten by Fisher and printed on a pink label) // H. M. Pendlebury / coll. / F.M.S. Museums (verso of the same label), in USNM; ♂ Borneo (CRV).

Remarks. – Both *Orsidis cariosus* and *Trachystola puncticollis* show many characters not corresponding to any other *Acalolepta* species: dorsal side entirely covered by a very dense punctation; interantennal space restricted; pronotum elongated, cylindrical, with acute pronotal spines abruptly emerging at the sides; elytra with pubescent bulges or granules.

Analogue structures can only be found in the Philippine genus *Granulorsidis* Breuning, 1980 (type-species: *Orsidis flavidosignatus* Aurivillius, 1927), where both species should be transferred. Consequently, the genus *Granulorsidis* should no longer be considered a Philippine endemic, but a Bornean genus with expansion into neighbouring regions.



1. *Granulorsidis flavidosignatus* (Aurivillius, 1927): 1a, ♂ (in CFV); 1b, ♀ (in CFV); 2. *Granulorsidis cariosus* (Pascoe, 1866): 2a, ♂, HOLOTYPE (in BMNH); 2b, ♀ (in CFV); 3. *Granulorsidis puncticollis* (Fisher, 1935): ♂ (in CRV).

***Sternohammus celebensis* Breuning, 1935** (Fig. 4a-b)

Examined material. – 1♂, Sulawesi Tengah, Palolo, III-1997 in CVN; 1♂, 1♀, Sulawesi Utara, Pangia, XII-2018, loc. coll., in CDH.

***Sternohammus celebensis submaculatus* (Gilmour, 1947) n. comb., n. status** (Fig 5a-b)

Dihammus submaculatus Gilmour, 1947: 148 (Indonesia, Sangir Besar: Tahuna) or. comb.

Sternohammus sanghiricus Breuning, 1956: 670 (Indonesia, Sangir); BREUNING, 1961: 378; BREUNING, 1962: 1, **n. syn.**

Acalolepta submaculata BREUNING, 1961: 378.

Examined material. – 15♂♂, 10♀♀, Indonesia, Sangir, Talawid, III-2014, S. Alam *leg.*, in CFV; 2♂♂, ditto, in CJPR.

Remarks. – GILMOUR (1947) described *Dihammus submaculatus* from “Taroena, Groot-Sanghir”, currently Tahuna, the chief town of Sangir Besar, the main island of the archipelago. Nine years later, BREUNING (1956) described *Sternohammus sanghiricus* and then (BREUNING, 1962) synonymised this species with *D. submaculatus*, incomprehensibly considering valid his own species even if the description was subsequent. The anteriorly truncate prosternum supports Breuning’s attribution at genus level, but Gressitt’s species should have the priority.

S. submaculatus constitutes the natural bridge between *S. celebensis* (Fig. 4) and the Philippine *S. femoralis* (Aurivillius, 1927) (Fig. 6). The elytral apex is so variable that this character alone does not allow distinguishing it from either *S. celebensis* (which shows a minute, nearly absent, spine) or from *S. femoralis* (which shows a long spine). However, its inflated antennomere VIII in males makes it apparently closely related to *S. celebensis*, of which it might be more correctly considered as a subspecies.

***Sternohammus femoralis* (Aurivillius, 1927)** (Fig. 6a-b)

Dihammus femoralis Aurivillius, 1927: 553 (Philippines: Luzon, Samar and Mindanao).

Sternohammus femoralis BREUNING, 1944: 498.

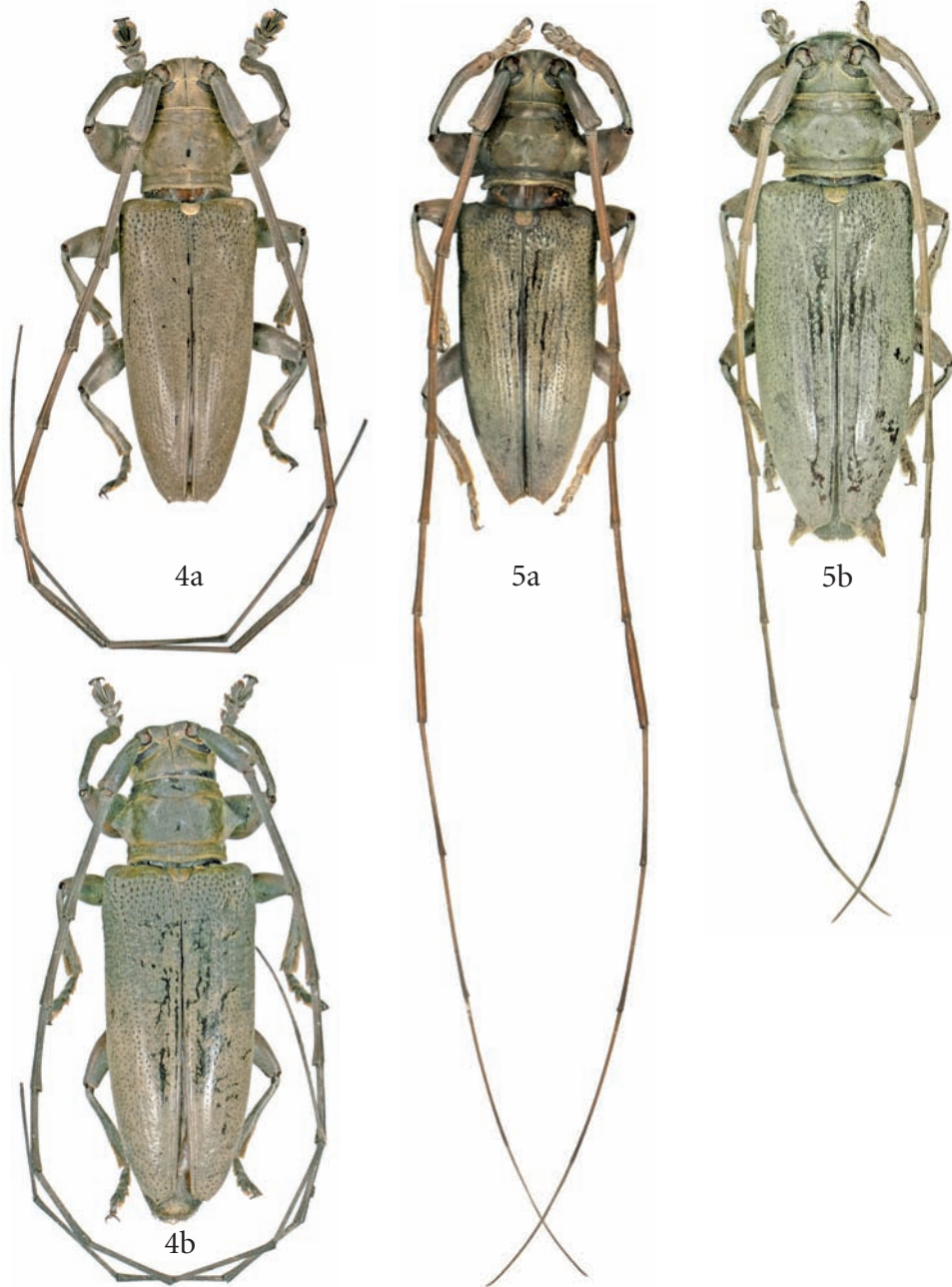
Sternohammus femoraloides Breuning, 1980: 173 (Philippines: Mindanao), **n. syn.**

Agniohammus philippinensis VIVES, 2017: 55, Fig. 12 misidentification.

Examined material. – 1♂, Philippines, VI-2014, loc. coll., in CDH; 1♂, Central Luzon, Aurora, Dingalan, Sierra Madre, X.2017, I. Lumawig *leg.*, in CFV; 1♀, Leyte, III-2002, D. Layron, in CDH.

Remarks. – This species is known from the entire Philippine Archipelago (AURIVILLIUS, 1927; BREUNING, 1944). According to the original description, *S. femoraloides* differed in vague relative characters, i.e. antennal proportions, smooth pronotal disc (vaguely punctured in femoratus) and brown pubescence (yellowish grey in femoratus). All types have disappeared (HÜDEPOHL, 1983), but the original description does not show characters supporting the validity of this taxon, which was in all likelihood a synonym of the cohabitant *S. femoralis*. In fact, the examined material, including a long series of the closely related *submaculatus*, shows the great variability of these characters (also depending on the age of the specimen).

The male specimen from Mindanao that VIVES (2017) misidentified as “*Agniohammus philippinensis* Breuning, 1938” actually belongs to *S. femoralis*. In fact, the pronotum evidently misses the internal apical furrow (typical of *Agniohammus*) and shows two straight basal furrows (typical of *Sternohammus* but absent in *Agniohammus*). Moreover, *Agniohammus philippinensis* is a species endemic to Basilan (BREUNING, 1961).



4. *Sternohammus celebensis* Breuning, 1935: 4a, ♂ (in CDH); 4b, ♀ (in CDH); 5. *Sternohammus celebensis submaculatus* (Gilmour, 1947): 5a, ♂ (in CFV); 5b, ♀ (in CFV).

Acalolepta (Pilohammus) n. subgen. (Fig. 7-8)

In order to rationalize the study of the genus *Acalolepta*, I propose the subgenus *Pilohammus* (type species: *Monohammus mixtus* Hope, 1841) for the species showing the following combination of characters: male protibia with a simple furrow; scape provided with scattered recumbent setae of contrasting colour (Fig. 7-8). Additionally, the pronotal disc is depressed and provided with three elevated bulges, the elytra are almost parallel-sided in both sexes and apically truncated.

Pilohammus n. subgen. includes *Acalolepta mixta* (Hope, 1841) (Fig. 7) and *A. ampliata* (Gahan, 1888) (Fig. 8) and is widespread in Australia, Solomon Islands and Vanuatu. Possibly, further species living in Papua New Guinea might be attributed to this subgenus.

Acalolepta (Dihammus) producta (Pascoe, 1866) (Fig. 9)

Monochamus productus Pascoe, 1866: 294 (Indonesia, Moluccas: Buru).

Monohammus productus GEMMINGER & HAROLD, 1873: 3016.

Haplohammus productus AURIVILLIUS, 1914: 692.

Dihammus productus AURIVILLIUS, 1921: 98; BREUNING, 1944: 487, *partim*.

Dihammus holotephrus BREUNING, 1944: 487–488, *partim*.

Acalolepta producta BREUNING, 1961: 377; VITALI, 2011: 293, 296, *partim*.

Acalolepta buruana Breuning, 1970: 476 (Indonesia, Moluccas: Buru); VITALI, 2011: 294, 296, **n. syn.** (Fig. 10a-b).

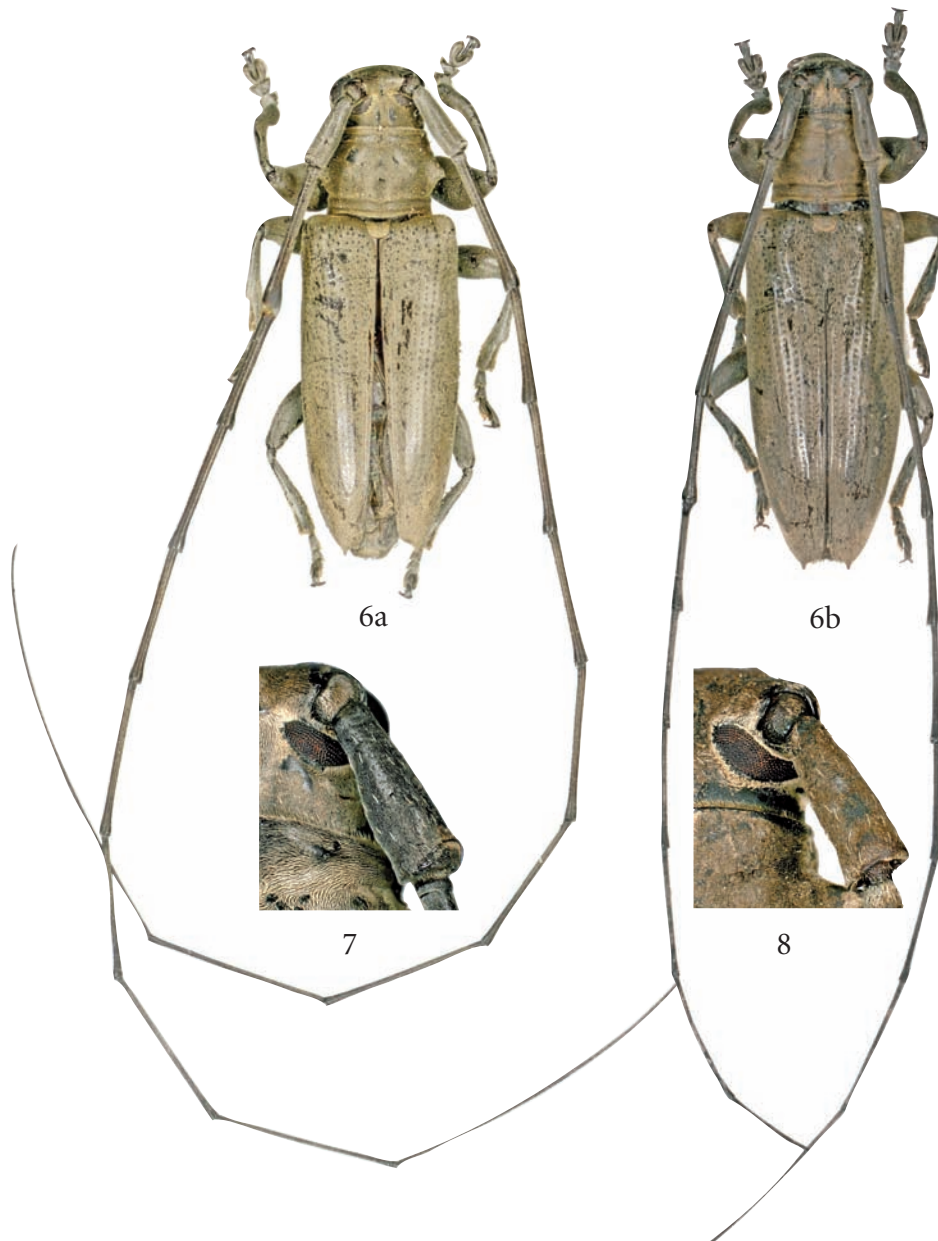
Examined material. – *Monochamus productus* Pascoe, 1866, 1♂, SYNTYPE, Bouru, [1861, A. Wallace leg.], in BMNH; *Acalolepta buruana* Breuning, 1970, 1♂, HOLOTYPE, Mt. Mada / Buru 3000' / Sept. [18]98. / (Dumas) (handwritten on a red label), *Acalolepta buruana mihi* Typ. Breuning dét. (handwritten by Breuning and printed on a white label); Muséum Paris / 1952 / coll. A. Oberthür (printed on a white label), TYPE (printed on a red label), in MNHNP. 1♀, Paratype, Kayeli / Buru' Oct. [18]98. / (Dumas) (handwritten on a yellow label), *Acalolepta buruana mihi* Paratyp. Breuning dét. (handwritten by Breuning and printed on a white label); Muséum Paris / 1952 / coll. A. Oberthür (printed on a white label), PARATYPES (printed on a red label), in MNHNP; 4♂♂, 7♀♀, Ambon, Kalauly village, XII-2017, V. Voitshovskii leg., in CFV; 1♂, Bacan, IV-2009, B. Cavelius leg., in CFV.

Remarks. – PASCOE (1866) described *Monochamus productus* on the basis of at least one pair of specimens that Alfred Wallace had collected in Buru in May and June 1861 (WALLACE, 1890: 293). The description (“*piceus, supra pube grisea interrupta tectus; prothorace postice vage punctato, disco sub-bituberculato; elytris maculis glabris dispersis, apicibus oblique truncatis ...* 13 lines”) does not leave any doubt that it is about the large species of Buru.

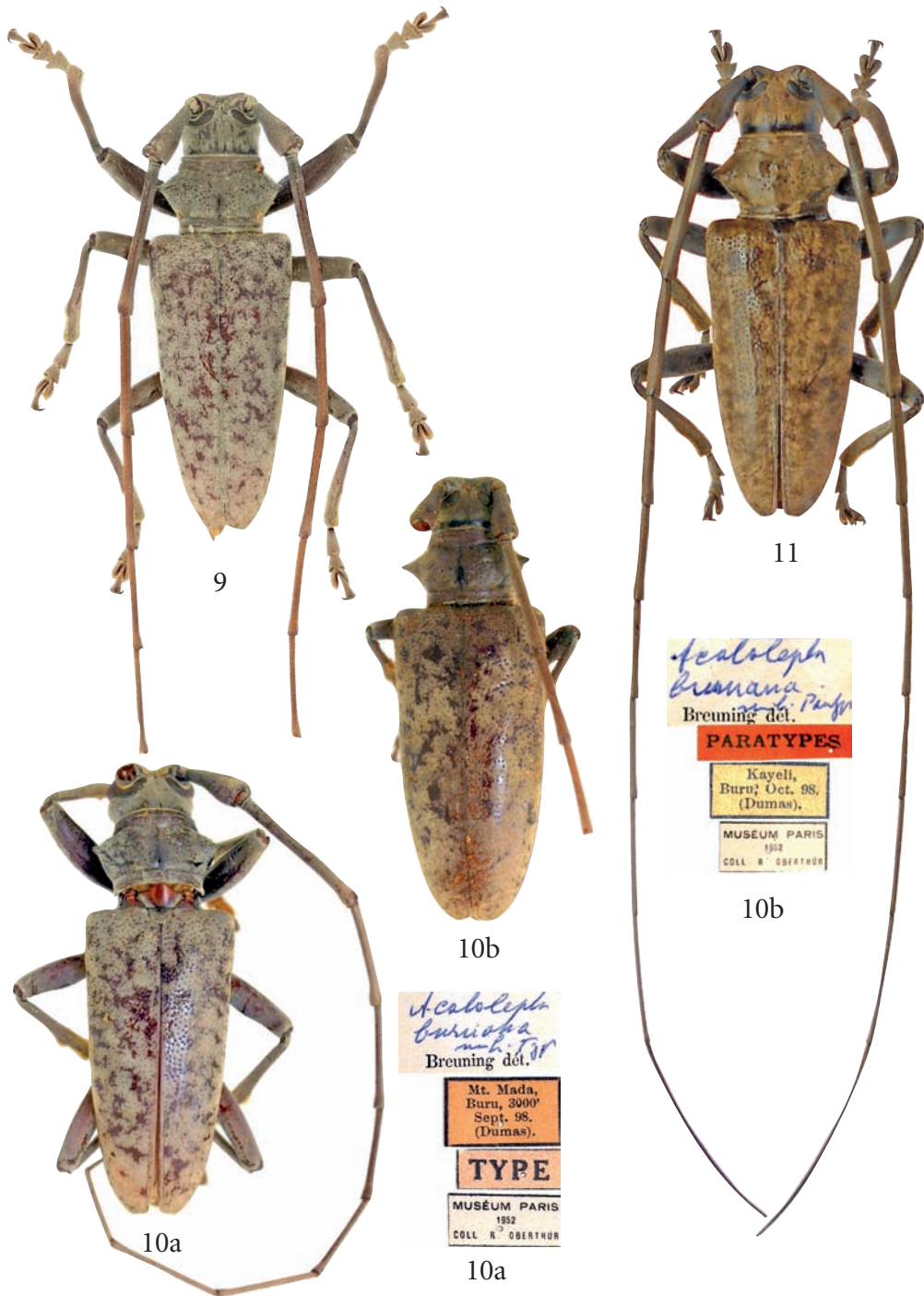
Ignoring both original description and the true aspect of the type (Fig. 9), BREUNING (1943a; 1944) attributed to *producta* wrong characters (body reddish brown with yellowish brown pubescence rather than body pitch-brown with yellowish grey pubescence) that perpetuated further incorrect identifications. Consequently, BREUNING (1970) described *buruana* (Fig. 10a-b) with characters perfectly corresponding to *producta*. The author also stated that the holotype was female and the paratype of undetermined sex; actually, the holotype is male and the paratype is female. As previously noted (VITALI, 2011), *buruana* is indistinguishable from *producta* and should be considered as a junior synonym.

Following BREUNING (1943a), VITALI (2011) mistakenly identified as *producta* specimens coming from Ceram, which (BREUNING, 1944) had incorrectly recorded as *A. celebensis* (Breuning, 1935). Actually, they belong to a new taxon that will be described subsequently.

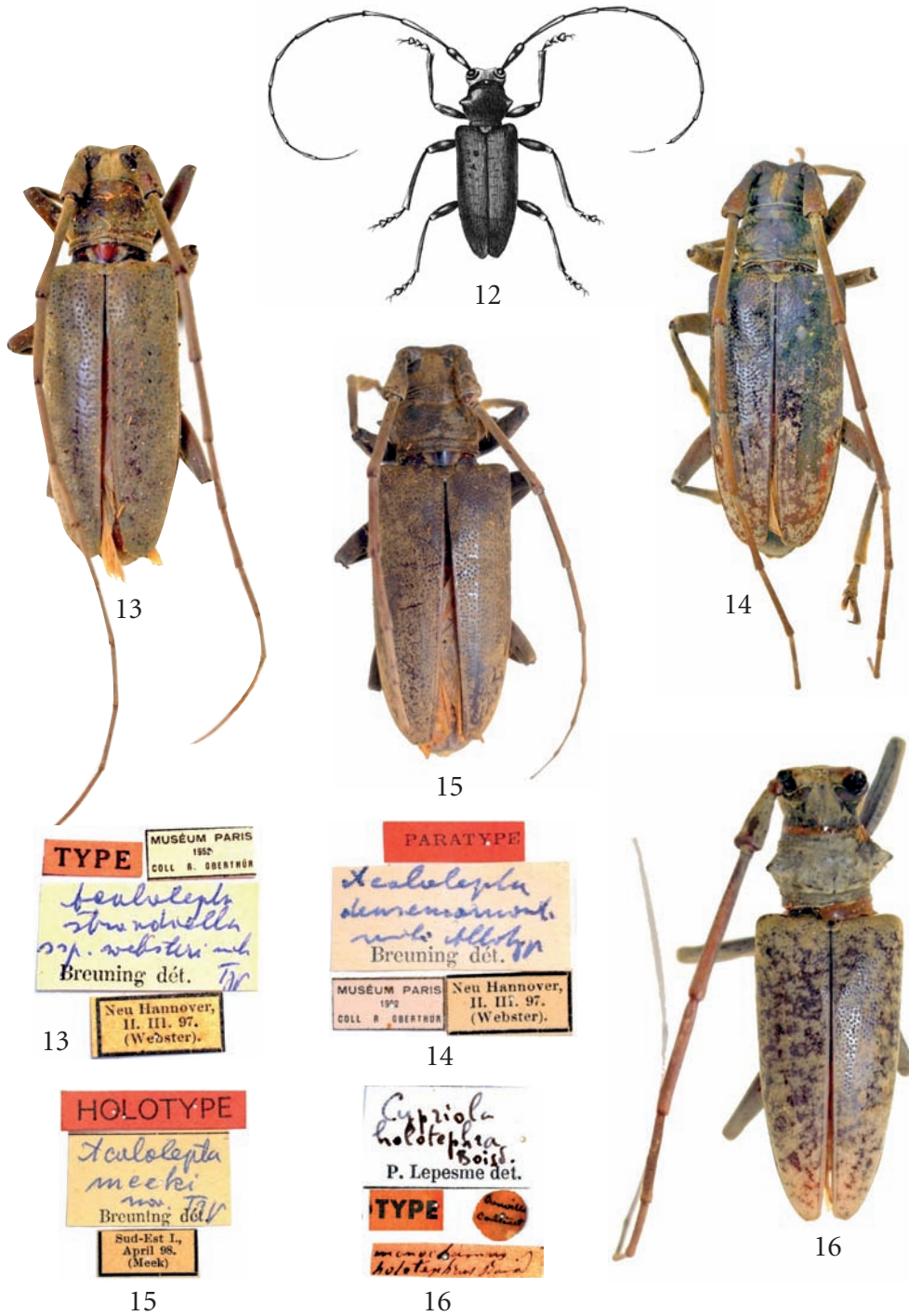
Distribution. – The species is present in Buru, Ambon, Watoebela and possibly, Bacan. Records from Ceram (VITALI, 2011) must be referred to the following taxon.



6. *Sternohammus femoralis* (Aurivillius, 1927): 6a, ♂ (in CDH); 6b, ♀ (in CFV); 7. *Acalolepta (Pilohammus) mixta* (Hope, 1841): scape (in CFV); 8 *Acalolepta (Pilohammus) ampliata* (Gahan, 1888): scape (in CFV).



9. *Monochamus productus* Pascoe, 1866: ♂, HOLOTYPE (in BMNH). 10. *Acalolepta buruana* Breuning, 1970: 10a, ♂, HOLOTYPE (in MNHNP); 10b, ♀, PARATYPE (in MNHNP). 11. *Acalolepta (Dihammus) ceramensis* n. sp.: ♂ HOLOTYPE (in CFV).



12. *Lamia holotephra* Boisduval, 1835: LECTOTYPE. Fig. 13. *Acalolepta strandiella websteri* Breuning, 1970: ♂, HOLOTYPE (in MNHNP). 14. *Acalolepta densemarmorata* Breuning, 1970: ♂, HOLOTYPE (in MNHNP). 15. *Acalolepta meeki* Breuning, 1982: ♀, Holotype (in MNHNP). 16. *Lamia holotephra* Boisduval, 1835: ♂, false Holotype (in MNHNP) = *Acalolepta (Dihammus) producta* (Pascoe, 1866).

***Acalolepta (Dihammus) ceramensis* n. sp.** (Fig. 11)

Dihammus celebensis BREUNING, 1944: 478 *partim*.

Acalolepta producta VITALI, 2011: 293, 296, Pl. 40, figs. 5–6 misidentification.

Examined material. – HOLOTYPE: 1♂, Indonesia, Moluccas, Seram, V-2017, S. Jakush *leg.*, in CFV. PARATYPES: 1♂, Indonesia E, Maluku Tengah prov., Seram N, Seram Utara distr., trans-Seram road between Masohi and Sawai, between Makariki and Horale (former Saka), 13 km SW from Horale vill., river valley, shrubs, gardens and secondary lowland forest, UV light, 5/6-IV-2009, *leg.* D. Telnov & K. Greke, in CDT; 1♀, ditto, beaten, 5/6-IV-2009, *leg.* D. Telnov & K. Greke, in CDT; 2♂♂, ditto, edge of secondary lowland forest, white light, 7-IV-2009, *leg.* D. Telnov & K. Greke, in CDT; 2♂♂, ditto, secondary lowland forest, on branches, 8-IV-2009, *leg.* D. Telnov & K. Greke, in CDT; 1♂, ditto, in CFV; 1♂, Indonesia E, Maluku Tengah prov., Lease Is., Saparua, ~2.5 km E from kota Saparua, young shrubs and clearing in secondary forest, beaten from thin branches, 12-IV-2009, *leg.* D. Telnov & K. Greke, in CDT.

Description. – Body length 19–33 mm. Habitus slightly elongated; integument pitch-brown, covered with a fine, more or less dense, brownish yellowish pubescence, which forms irregular patches, where the ground colour appears more or less visible. Head, scutellum, limbs and ventral side covered with uniform pubescence.

Frons covered with coarse strong punctures; eyes about twice as long as genae; vertex relatively wide, marked with some fine punctures.

Antennae almost thick, long, 2.5 times as long as body in male; antennomere V does not reach the elytral apex; scape strongly thickened apically, with an open apical cicatrix; antennomere III twice as long as scape; antennomere XI twice as long as X.

Pronotum feebly transverse, armed with an obtuse large perpendicular tubercle at each side, furrowed by two grooves at both apical and basal margin; external apical margin and both basal margins straight; internal apical margin posteriorly convex; disc feebly uneven, more or less densely covered with fine punctures, as fine as those of vertex. Scutellum semicircular.

Elytra relatively short (each elytron 4 times as long as wide at base), regularly tapered to the apex; sides slightly convex; apices obliquely subtruncate; disk covered with a fine punctation, denser at base, sparse in the middle and extremely fine or lacking on the apical third.

Legs and tarsi relatively short, protibiae arcuate at apex, with a pre-apical furrow and a tooth in male.

Last visible urosternite of male posteriorly truncate, with semi-recumbent dense yellow pubescence at apex.

Female: Similar to the male but with antennae twice as long as body, protibiae without apical tooth and elytra parallel-sided.

Differential diagnosis. – This species is closely related to *A. rusticatrix* (Fabricius, 1801) and *A. itzingeri* (Breuning, 1935), with which it was probably confused in the past (PASCOE, 1866). It differs from *A. rusticatrix* in the punctured vertex, the uniform colour of the head (in *A. rusticatrix*, the frons is always lighter as the vertex, a fact easily observable from the upper side), the scutellum having the same colour of the elytra (in *A. rusticatrix*, the scutellum is always lighter than elytra), the elytra with much finer, sparse punctures on the disc and almost without punctures on the apical third (in *A. rusticatrix*, the elytra show distinct punctures to the apex, nearly as dense as those of the middle).

It differs from *A. itzingeri* in the darker mottled pubescence (uniformly grey-yellowish in *A. itzingeri*), the pronotum with a well marked internal apical margin (internal apical margin absent in *A. itzingeri*) and with the discal punctures anteriorly reaching the internal apical margin (reaching the external apical margin in *A. itzingeri*) and finally, the slightly convex elytral sides of male (nearly rightly tapered from base to apex in *A. itzingeri*). Moreover, the scutellum is posteriorly rounded in *A. ceramensis* n. sp., while it is posteriorly truncate in both observed males of *A. itzingeri* (Kei, Ohoidertawun 17/20-II-2011, *A. Skale leg., in CAS*), but I ignore if this character is constant.

Considering other *Dihammus* species present in the Moluccas, *A. ceramensis* n. sp. differs from *A. producta* (Fig. 9) in the elytra mottled with more or less pubescent brown spots (mottled with hairless large spots in *producta*) and from *A. celebensis* (Breuning, 1935) in the head and pronotum covered with strong punctures (last abdominal sternite of male without erect pubescence; head and pronotum covered with some fine punctures in *celebensis*) and in the last abdominal sternite of male covered with erect pubescence.

Distribution. – Endemic to Ceram. I examined the photos of a female, 29 mm long coming from Halmahera (*in CGC*), which nevertheless, I do not add to the list of types since the origin seems to be doubtful.

***Acalolepta holotephra* (Boisduval, 1835)** (Fig. 12)

Lamia holotephra Boisduval, 1835: 498, Pl. VIII, Fig. 3 (Papua New Guinea, Bismarck Is., New Ireland: Port Carteret); BREUNING, 1944: 487–488 *partim*.

=? *Acalolepta strandiella websteri* Breuning, 1970: (Papua New Guinea, Bismarck Is., New Hanover) (Fig. 13).

=? *Acalolepta densemarmorata* Breuning, 1970: (Papua New Guinea, Bismarck Is., New Hanover) (Fig. 14).

=? *Acalolepta meeki* Breuning, 1982: (Papua New Guinea, Louisiade Is.: Vanatinai) (Fig. 15).

Examined material. – 1♂, Durville [Port] Carteret (handwritten on a round yellowish label), *Monochamus / holotephrus* Boisd. (handwritten on a pink label), *Cypriola holotephra* Boisd. / P. Lepasme det. (handwritten by Lepasme and printed on a white label), TYPE (printed on a red label), *in* MNHNP.

Remarks. – *Lamia holotephra* was one of the first described species of *Acalolepta*. According to BOISDUVAL (1835), it was characterised by a dense, nearly fasciculate, silky greyish pubescence, which almost obscured the dorsal puncturing and the pronotum striated transversely.

It was described from the surroundings of Port Carteret, an anchorage located in southern New Ireland, north of Lambom, 4°39'S, 152°54'E (KRUZENSTERN, 1824). But, due to its lacks of salient distinguishing features (Fig. 12), several authors (BLANCHARD, 1853; MONTROUZIER, 1855, BREUNING, 1944) misidentified it with other species collected in different parts of the Australian region. According to BREUNING (1944), *holotephra* was similar to the Moluccan *Acalolepta producta*, a very odd observation, since *holotephra* should be similar to the numerous grey species described from the same biogeographical region (Bismarck, Solomon and Vanuatu Islands).

Unfortunately, Breuning's observation derived from the assumed type conserved in the MNHNP (Fig. 16). Actually, this specimen is not the true type of *Lamia holotephra*.

In fact, the labels have wrong names. First, “Durville” must be referred to Jules Dumont d’Urville, captain of the French ship *Astrolabe*, but the spelling is different. Moreover, BOISDUVAL (1835) nowhere expressly mentioned the fact that d’Urville collected this species, as for other ones collected by the captain. Second, the identification label refers to “*Monochamus holotephrus* Boisd.,” rather than “*Lamia holotephra*,” as this species was originally described. The combination “*Monochamus holotephrus*” was originally used only 18 years later, when BLANCHARD (1853) misidentified with the name *Acalolepta antenor magnetica* (Pascoe, 1866) (cf. VITALI, 2017). Third, the specimen does not correspond to *holotephra* since it shows a yellowish opaque pubescence and large hairless black spots missing in original description and drawing. In addition, BOISDUVAL (1835: 80) noted that “il y a eu, relativement aux insectes de l’Astrolabe, confusion au Muséum national d’histoire naturelle, et on a noté comme de Vanikoro une foule d’espèces qui ont été prises soit à la Nouvelle-Hollande, soit à la Nouvelle-Guinée” (concerning the insects of *Astrolabe*, there has been confusion at the Museum of Natural History of Paris and a lot of species collected in New Holland and New Guinea were recorded from Vanikoro). But this confusion also concerned many other specimens.

Thus, the presumed type (Fig. 16) was misidentified, as it also occurred with the “type” of *Cypriola acanthocinoides* Thomson, 1864 (cf. VITALI, 2017). In reality, this is the first known specimen of *Acalolepta producta*, which PASCOE (1866) described only 40 years later from other specimens collected by Wallace in 1861 in Buru (Fig. 9). The *Astrolabe* as well had visited this island during its first trip (1826-1829).

The true type of *Lamia holotephra* seems to be lost, as also occurs with the type of *Lamia (Monohammus) fasciata* Montrouzier, 1855 (cf. VITALI, 2017). Waiting for further researches in the MNHNP, I temporarily designate as LECTOTYPE the specimen figured by BOISDUVAL (1835, Pl. 8. Fig. 3), accordingly to the ICZN, 1999, Art. 74.4. (designation by means of an illustration or description).

This species was also recorded from the Solomon Islands (BREUNING, 1944; BIGGER & SCHOFIELD, 1983; VITALI & CASADIO, 2007) and I examined some specimens from Guadalcanal that could correspond to HOLOTEPHRA.

Acalolepta strandiella websteri Breuning, 1970 (Fig. 13) and *A. densemarmorata* Breuning, 1970 (Fig. 14), this latter described from a rubbed specimen, as well coming from the Bismarck Islands and corresponding at all to the description of *Lamia holotephra*, are in all likelihood junior synonyms of this species. *Acalolepta meeki* Breuning, 1982 (Fig. 15) from the Louisiade Archipelago is possibly another synonym or a closely related taxon. However, the taxonomic status of all these species will be clarified through the examination of further specimens coming from New Ireland.

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