# Advances in Carabidology

Papers Dedicated to the Memory of Prof. Dr. Oleg L. Kryzhanovskij

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# Morphological study of *Perileptus* larvae (Coleoptera Carabidae Trechitae)

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Abstract. Larvae of the genus Perileptus Schaum, 1860 are described or redescribed: P. areolatus (Creutzer, 1799) (all instars) and P. mesasiaticus Uéno, 1976 (first and second instars). Based on the larval morphology, the genus Perileptus is closely related to the genus Thalassophilus Wollaston, 1854. This relationship is supported with three synapomorphic character states: (1): Anterior claw bears a hyaline organ on its dorsal surface which extends to its apex; (2): Presence of a flat, ventrally curved, long seta attached between the clawes; (3): Seta UR, on urogomphi located near seta UR, not near UR,. The presence of two claws is presumably a plesiomorphic character. Taxonomic position of the genera Perileptus and Thalassophilus within the supertribe Trechitac is unclear. Neither of these two genera posses any of the significant autapomorphic larval characters thought to be definitive for Trechitae. Some bionomics data of Perileptus larvae are shortly discussed.

Резюме. Описаны или переописаны личинки рода Perileptus Schaum, 1860: P. areolatus (Creutzer, 1799) (все возрасты) и P. mesasiaticus Uéno, 1976 (первый и второй возрасты). На основании морфологии личинок, род Perileptus наиболее близок роду Thalassophilus Wollaston, 1854. Их родственные связи подтверждаются тремя синапоморфными состояниями признаков: 1) Передний коготок несет на дорсальной поверхности гиалиновый орган, направленный вперед; 2) Присутствует плоская, изогнутая вентрально длинная хета, прикрепленная между коготками; 3) Хета UR<sub>3</sub> на урогомфах расположена рядом с хетой UR<sub>4</sub>, а не UR<sub>2</sub>. Присутствие двух коготков является, возможно, плезиоморфным признаком. Таксономическое положение родов Perileptus и Thalassophilus в надтрибе Trechitae остается неясным. Ни один из этих двух родов не обладает каким-либо важным аутапоморфным личиночным признаком, который считается определяющим для Trechitae. Кратко обсуждены некоторые данные по биологии личинок Perileptus.

Key words: larvae, morphology, taxonomy, phylogeny, Coleoptera, Carabidae, Trechini, Perileptus.

#### Introduction

The genus Perileptus Schaum, 1860, along with the Asiatic Neoblemus Jeannel, 1923 and Apoplotrechus Alluaud, 1915 from Madagascar, belongs to the subtribe Perileptina of the tribe Trechini. The subtribe is distributed in the Palaearctical, Afrotropic, Oriental, and Australian Regions with one species in Cuba (Casale & Laneyrie, 1982). According to these authors, the genus includes 42 species. Since then, Baehr (1987) described 3 new species from Australia.

Very little is known about the larval morphology of *Perileptus*. Boldori (1936) described the larva of the European *Perileptus areolatus* (Creutzer, 1799). Almost a half of century later, Luff (1985) redescribed this species based on Boldori's material. This genus was included several times in identification keys of carabid larvae (Van Emden, 1942; Sharova, 1958, 1964; Larsson, 1968; Arndt, 1991; Luff, 1993; Makarov, 1994). So, our knowledge about the larval morphology of *Perileptus* is based on the three specimens, collected by Leonida Boldori in July 21, 1935.

In 1996 the senior author reared ex ovo larvae of two species of the genus Perileptus: P. areolatus from Caucasus and P. mesasiaticus Uéno, 1976 from Kugitang Range (East Turkmenistan). This new material allows us to conduct a new study of the morphology, including larval chaetotaxy, of this genus. Additionally, larvae from five tribes and a number of genera of the supertribe Trechitae were part of this compartive study.

#### MATERIAL AND METHODS

All studied larvae were reared ex ovo from the field collected females. Geographic data are provided below, under each studied species. The material is deposited in the collections of the authors, the British Museum of Natural History, London (BMNH) and the Canadian National Collection, Ottawa (CNC).

Some larvae of each species were mounted on permanent slides. A light stereo microscope MBS-1 was used at magnification 80 to 900 x. All drawings have been done with the aid of a camera lucida. Notation of the setae and pores follows Bousquet & Goulet (1984) and Bousquet (1985).

All measurements were made using a micrometer. The following abbreviations are used in the descriptions of species: HW – maximum width of head; HL – length of head along middle line. Symbols  $L_1$ ,  $L_2$ , and  $L_3$  mean the first, second, and third instar larvae respectively.

For comparison larvae of 25 genera, representing 5 tribes of the supertribe Trechitae (sensu Kryzhanovski), 1976, 1983; Kryzhanovski) et all, 1995) have been studied: the tribe Trechini (genera Aepopsis Jeannel, 1992, Thalassophilus Wollaston, 1854, Temnostega Enderlein, 1905, Amblystogenium Enderlein, 1905, Trechus Clairville, 1806, Epaphius Stephens, 1827 and Trechimorphus Jeannel, 1927); the tribe Bembidiini (genera Bembidion Latreille, 1802, Asaphidion Des Gozis, 1886, Ocys Stephens, 1829 and Phrypeus Casey, 1924); the tribe Tachyini (genera Tachys Stephens, 1829, Paratachys Casey, 1918, Sphaerotachys J. Müller, 1926, Elaphropus Motschulsky, 1839, Porotachys Netolitzky, 1914, Tachyta Kirby, 1837, Mioptachys Bates, 1882 and Polyderis Motschulsky, 1862); the tribe Pogonini (genera Pogonus Nicolai, 1822, Pogonistes Chaudoir, 1870, Cardioderus Dejean, 1829 and Thalassotrechus Van Dyke, 1918); the tribe Zolini (genera Oopterus Guérin-Méneville, 1841 and Idacarabus Lea, 1910).

# Genus Perileptus Schaum, 1860

#### DIAGNOSIS

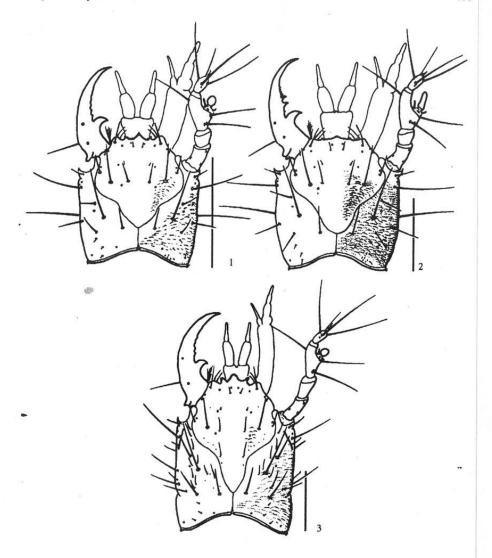
Within the supertribe Trechitae, larvae of the genus *Perileptus* (and *Thalassophilus*; see Grebennikov, 1996) can be easily recognised by the presence of a flat, curved claw seta as long as the claws themselves (Figs 13, 14). Differences between the genera *Perileptus* and *Thalassophilus* are noted below:

Perileptus (Figs 1, 3, 4, 6, 7, 9, 11 - 17):  $L_1$ - $L_3$ : pore PA<sub>b</sub> on parietale present (Figs 1, 3);  $L_1$ - $L_3$ : antennomere 3 with rounded sensorium (Figs 1, 3, 7);  $L_1$ - $L_3$ : mandibles with small teeth on terebra (Figs 1, 3);  $L_1$ : nasale as Fig. 4;  $L_1$ : setae EM<sub>1</sub> on meso- and as metanotum modified as pore-like sensilla (Fig. 12);  $L_1$ : central ventrite of abdomen without additional setae;  $L_1$ : seta EP<sub>1</sub> on ninth abdominal segment modified as 2 or 3 pore-like sensilla (Fig. 17).

Thalassophilus (Figs 2, 5, 8, 10):  $L_1$ - $L_3$ : pore PA<sub>b</sub> on parietale absent (Fig. 2);  $L_1$ - $L_3$ : antennomere 3 with elongated sensorium (Figs 2, 8);  $L_1$ - $L_3$ : mandibles with large teeth on terebra (Fig. 2);  $L_1$ : nasale as fig. 5;  $L_1$ : setae EM<sub>1</sub> on meso- and metanotum absent;  $L_1$ : central ventrite of abdomen with one long additional seta on each side;  $L_1$ : seta EP<sub>1</sub> on ninth abdominal segment absent.

#### DESCRIPTION

First instar larvae. Body almost colourless, head light brown; larva less sclerotised than usual for Trechitae members, sclerotisation of lateral margins of tergites



Figs 1-3. Larvae of the genera *Perileptus* and *Thalassophilus*: cephalic capsule, right antenna, left mandible, right maxilla, and labium, dorsal view: 1-2, first instar; 3, third instar. 1-3, *Perileptus areolatus*; 2, *Thalassophilus longicornis*. Scale bars: 0.2 mm.

unclear; all primary sensilla present (except as noted below), their positions as in the generalized type (see Bousquet & Goulet, 1984); additional sensilla absent (except as noted below).

Cephalic capsule (Fig. 1) parallel sided or slightly convergent posteriorly; ocelli absent; ocellar tubercles little developed or absent; postocellar and cervical grooves absent; frontal sutures more or less curved; epicranial suture present, rather long; nasale (Fig. 4) with numerous teeth; central projection of nasale composed of several rows of teeth one under another. Microsculpture (Fig. 1): frontale with slight transverse microsculpture at base; parietale with transverse microsculpture

throughout or on dorso-lateral surfaces; parietale near seta  $PA_1$  with 6 - 8 microspones; egg-bursters absent. Chaetotaxy (Fig. 1): distance between setae  $FR_1$ - $FR_2$  =  $FR_2$ - $FR_3$ ;  $FR_3$ - $FR_4$  =  $FR_4$ - $FR_5$ ; anterior angles of hypopharynx with single seta on each side.

Antennae (Fig. 7). Distal part of antennomere 3 very long and fully sclerotised (including lateral surface between sensorium and base of antennomere 4); all conical and companiform sensilla on antennomeres 3 and 4 well developed; sensorium on antennomere 3 relatively large, round; seta AN<sub>6</sub> about 1.5 x length of antennomere 4.

Mandibles (Fig. 1) relatively straight, little curved; terebra with some slightly developed teeth; dorsal keel at apex of mandible not developed; microsculpture on dorsal surface absent; retinaculum perpendicular; penicillus not extending to apex of retinaculum.

Maxillae (Fig. 9) with comparatively narrowed and elongated articles, especially stipes and galeomere 2; dorsal surface fully sclerotised; group gMX comprising 6 - 11 setae, one of them placed apically at level of seta MX<sub>6</sub>, the rest of the setae on basal half of stipes; seta MX<sub>6</sub> short, its length about 0.4 x that of MX<sub>5</sub>; second galeomere 2 x longer than galeomere 1; seta MX<sub>7</sub> relatively short, reach only to basal 1/3 of galeomere 2; seta MX<sub>8</sub> at middle of galeomere 2; setae MX<sub>11</sub>, MX<sub>12</sub> small, not longer than 1/4 width of palpomere 3; palpomere 4 undivided.

Labium (Fig. 9). Setae LA<sub>3</sub> and LA<sub>4</sub> comparatively short and thin; ligula not developed and not sclerotised; seta LA<sub>6</sub> flat; second article complete, undivided.

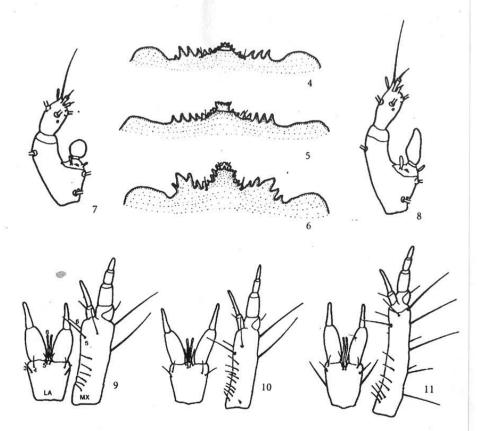
Thorax (Fig. 12) without primary pores PR<sub>c</sub>, PR<sub>e</sub>, PR<sub>i</sub>, PR<sub>j</sub> on pronotum, ME<sub>d</sub>, ME<sub>e</sub> on meso- and metanotum, and seta ES<sub>1</sub> on meso- and metathorax; setae EM<sub>1</sub> on meso- and metathorax absent; one pore-like sensillum presents at position of EM<sub>1</sub> on meso- and metathorax; all other sensilla present (including PS<sub>2</sub> and MS<sub>4</sub> on ventral surface); additional pore-like sensillum presents near seta MS<sub>1</sub>; setae PR<sub>7</sub> and ME<sub>10</sub> comparatively long, as long as 7 - 10 basal diameters of the longest setae on tergite; length of setae PR<sub>13</sub> and ME<sub>14</sub> about 0.7-0.9 x that of setae PR<sub>12</sub> and ME<sub>13</sub> respectively; surface of tergites smooth; fine microsculpture present only at middle of pretergites of meso- and metanotum.

Legs (Fig. 13) with two claws (Fig. 14), anterior one about 1/10 longer than posterior; middle of anterior claw with hyaline organ on dorsal surface, extending to its apex; a long, flat, ventrally curved single claw seta attached between base of the two claws, its length about as long as claw; setae TA<sub>3</sub>, TA<sub>4</sub>, TA<sub>5</sub>, TA<sub>6</sub> absent; seta TA<sub>1</sub> at basal 1/5 of tarsus; setae TI<sub>1</sub> and TI<sub>2</sub> much thinner and longer than setae TI<sub>3</sub>, TI<sub>4</sub>, TI<sub>5</sub>, TI<sub>6</sub>, TI<sub>7</sub>.

Abdomen without primary pore  $TE_b$  on tergites 1-8; seta  $TE_8$  as long as 5-7 basal diameters of seta  $TE_9$ ; length of seta  $TE_{11}$  about 0.7-0.9 x that of  $TE_{10}$ ; tergites with fine pointed microsculpture on all surface.

Urogomphi and pygidium (Fig. 17). Urogomphi relatively straight; seta EP<sub>1</sub> on ninth abdominal segment absent; 2 or 3 pore-like sensilla present at position of EP<sub>1</sub>; seta UR<sub>3</sub> near seta UR<sub>4</sub>; seta UR<sub>9</sub> as long as 7-10 basal diameters of setae UR<sub>8</sub>.

Second and third instar larvae. Cephalic capsule (Fig. 3) more elongated and sclerotised; with developed latero-ventral keel; postocellar grooves present as short lines medially to seta PA<sub>9</sub>; cervical grooves absent; nasale: Fig. 6. Microsculpture



Figs 4–11. Larvae of the genera Perileptus and Thalassophilus, dorsal view: 4-6, nasale; 7-8, right antennomeres 3 and 4; 9-11, labium and right maxilla. 4, 5, 7-10, first instar larvae; 6, 11, third instar larvae. 4, 6, 7, 9, 11, Perileptus areolatus; 5, 8, 10: Thalassophilus longicornis.

(Fig. 3): parietale with developed transverse microsculpture; without microspines medially of seta PA<sub>1</sub>; frontale smooth, without microsculpture. Chaetotaxy (Fig. 3): parietale with 45-55 secondary setae on each side; frontale with a pair of secondary setae at base and 18-22 microsetae on basal half.

Antennae without secondary setae.

Mandibles (Fig. 3) without microsculpture and secondary setae; teeth on terebra larger than in first instar.

Maxillae (Fig. 11). Group gMX with 7 (in  $L_2$ ), 7-11 (in  $L_3$ ) setae; stipes laterally with 5 ( $L_3$ ) or 7 ( $L_3$ ) setae.

Labium (Fig. 11) with one thin and long secondary seta on each side.

Thorax (Fig. 15). Each side of tergite with 110-130 (pronotum) or 100-120 (meso- and metanotum) secondary setae; membranous zone at posterior angles of pronotum with three setae on each side near seta  $PR_{11}$ ; ventrite of pronotum with 18-22 setae on each side; pleurites of meso- and metanotum with one secondary seta; each small sclerite around seta  $MS_1$  with  $1 \, (L_2)$  or  $3-5 \, (L_3)$  pore-like sensilla; epimerite without sensilla; metathorax with one secondary seta at place of primary seta  $ES_1$ .

Legs (Fig. 13) without any secondary sensilla (except 2-3 setae on coxa).

ABDOMEN. Tergites with 50-70 secondary setae on each side; epipleurites with 2 secondary setae; hypopleurites with 15, outer postventrites with 15, inner postventrites with 8-10, central ventrite with 20 setae; sclerites of seta ST1 with 1  $(L_2)$  or 2-6  $(L_3)$  pore-like sensilla.

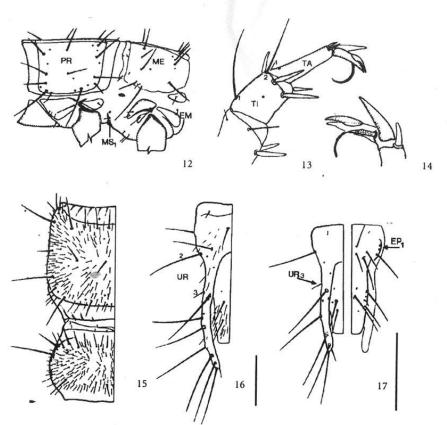
Urogomphi and pygidium (Fig. 16). Seta UR<sub>alpha</sub> present; urogomphi with 7 long setae (5 of them primary); lateral margins of ninth tergite with secondary seta at middle; urogomphi with 18-33 small secondary setae on each side; sternal sclerite with 15 small secondary setae on each side; epipleurites without pore-like sensilla, with 2 setae (the longest seta placed more anteriorly than the shorter one); pygidium with 70-90 small secondary setae on apical half dorsally, ventrally and laterally.

#### PHYLOGENETIC POSITION

- 1. Why does Perileptus belong to the supertribe Trechitae? Larvae of the genus Perileptus share all presumably apomorphic characters of the supertribe Trechitae: lack of primary pores PR<sub>c</sub>, PR<sub>e</sub>, PR<sub>i</sub>, PR<sub>j</sub> on pronotum, pores ME<sub>d</sub>, ME<sub>e</sub> on meso-and metanotum, seta ES<sub>1</sub> on metathorax, pore TE<sub>b</sub> on abdominal tergites 1-8, and setae TA<sub>3</sub>, TA<sub>4</sub>, TA<sub>5</sub>, TA<sub>6</sub> on tarsus. Presence of these sensilla is considered as plesiomorphic condition.
- 2. Sister group of Perileptus. Within all Trechitae larvae known to us, the genus Perileptus is most closely related to the genus Thalassophilus based on the presence of these three synapomorphies of all instars larvae:
- 1. The anterior claw bears a hyaline organ on its dorsal surface, extending to its apex (Figs 13, 14) (presumed plesiomorphic character state: claw (or claws) is without any hyaline organ on dorsal surface). This is a unique feature within the supertribe, but it should be noted that a somewhat similar structure is found by BOUSQUET (1996) on the claws of *Schizogenius lineolatus* Say, 1823, a member of the tribe Clivinini.
- 2. Claws bear a long, flat, ventrally curved single claw seta, which is about as long as the claws themselves (Figs 13, 14) (presumed plesiomorphic character state: claw seta short, at least 5 x shorter than claw (or claws), not flat, not curved).
- 3. Seta UR<sub>3</sub> on urogomphi located near seta UR<sub>4</sub>, not near UR<sub>2</sub> (Figs 16, 17) (presumed plesiomorphic character state: seta UR<sub>3</sub> on urogomphi located near UR<sub>2</sub>).

As for the presence of two claws, we believe this is a plesiomorphic character, shared by larvae in question with those of the genera *Temnostega*, *Amblystogenium* and *Trechimorphus* (unpublished data, partly). We do not use this assumed ancestral similarity to support a relationship between these genera. All other known members of the supertribe Trechitae have a single claw (presumably the apomorphic character state).

3. Relationship with Trechini is not supported. Taxonomic position of the genera Perileptus and Thalassophilus within the Trechitae is unclear. According to the imaginal classification, the genera Perileptus and Thalassophilus belong to the tribe Trechini. However, their larvae share no important apomorphic characteristics of the tribe Trechini. The synapomorphies of Trechini (which are absent in Perileptus and Thalassophilus) are as following:



Figs 12-17. Larvae of *Perileptus areolatus*: 12-14, lateral view; 15, 16, dorsal view; 17, dorsal and ventral views. 13, 14, all instar larvae; 12, first instar larvae; 15-17, third instar larvae. 12, prothorax and mesothorax; 13, claws, tarsus, tibia and apex of femur; 14, claws and apex of tarsus; 15, pronotum and mesonotum, left side; 16, urogomphi and pygidium, left side; 17, urogomphi and pygidium, left side dorsally and ventrally. Scale bars: 0.2 mm.

- 1.  $L_2$ - $L_3$ : Antennomere 2 bears relatively long secondary seta at apex (presumed plesiomorphic character state of *Perileptus* and *Thalassophilus*: antennomere 2 bears no secondary seta (Fig. 3)).
- 2.  $L_2$ - $L_3$ : Tarsus and tibia bear secondary setae (presumed plesiomorphic character state of *Perileptus* and *Thalassophilus*: tarsus and tibia bear no secondary seta (Fig. 13)).
- 3. The third important difference between larvae of *Perileptus* + *Thalassophilus* and those of other Trechini is the number of setae at anterior angles of hypopharynx in all larval instars. The phylogenetic direction of this transformation series is unclear. Larvae of *Perileptus* and *Thalassophilus* (as well as all known larvae of the tribes Bembidiini, Tachyini, Pogonini and Zolini) have a single seta at anterior angles of hypopharynx in all instar larvae. On the other hand, known Trechini larvae have two setae at anterior angles of hypopharynx (except for the larvae of the strange genus *Amblystogenium*).
- 4. Homologue or homoplasy with different Trechitae larvae. Larvae of the genera Perileptus (and Thalassophilus) share some presumably apomorphic characters with

other members of the different branches of the supertribe Trechitae. We believe, however, that almost all of these similarities are the result of convergention and do not reflect the true relationships between the taxa. These taxa and presumably convergent characters are as following:

- 1. L<sub>1</sub>-L<sub>3</sub>: Seta LA<sub>6</sub> on ligula is flat (Figs 9, 10, 11) (presumed plesiomorphic character state: seta LA<sub>6</sub> on ligula normal, more or less conical, not flat). This character is shared by larvae of the genera *Perileptus + Thalassophilus* and the subgenera *Synechostictus* Motschulsky, 1864 and *Pseudolimnaeum* Kraatz, 1888 of the genus *Bembidion* of the tribe Bembidiini (Grebennikov, 1997).
- 2. L<sub>1</sub>-L<sub>3</sub>: Distal part of antennomere 3 is relatively long; lateral surface between sensorium and base of antennomere 4 is fully sclerotised (Figs 7, 8) (presumed plesiomorphic character state: distal part of antennomere 3 is relatively short; lateral surface between sensorium and base of antennomere 4 is membranous, not sclerotised). This character state is shared between larvae of the genera *Perileptus* + *Thalassophilus* and those of the tribe Trechini.
- 3. L<sub>1</sub>-L<sub>3</sub>: Terebra on mandible bears some teeth (Figs 1-3) (presumed plesiomorphic character state: terebra on mandible smooth, without teeth). This character state is shared between larvae of the genera *Perileptus + Thalassophilus* and those of the genera *Sphaerotachys*, *Paratachys* and *Porotachys* of the tribe Tachyini (unpublished data).
- 4.  $L_1$ - $L_3$ : Seta MX<sub>8</sub> is located at middle of galeomere 2 (Figs 9-11) (presumed plesiomorphic character state: seta MX<sub>8</sub> is located at base of galeomere 2). This character state is shared between larvae of the genera *Perileptus* + *Thalassophilus* and those of the tribes Tachyini and Trechini (unpublished data).
- 5. L<sub>1</sub>: Setae EM<sub>1</sub> on both meso- and metathorax and seta EP<sub>1</sub> on ninth abdominal segment are strongly modified. They are absent (*Thalassophilus*) or replaced by pore-like sensilla (*Perileptus*) (presumed plesiomorphic character state: all these sensilla are short trichoid setae, as in generalized type; see BOUSQUET & GOULET, 1984). This character (modified sensilla EM<sub>1</sub> and EP<sub>1</sub>) is shared by larvae of the genera *Perileptus* + *Thalassophilus* and all members of the tribes Pogonini (spindle-like setae), Tachyini (pore-like sensilla) and some genera of the tribe Trechini (pore-like sensilla) (unpublished data).
- 6. L1: Setae  $TI_1$  and  $TI_2$  on tibia are much thinner and longer than setae  $TI_3$ ,  $TI_4$ ,  $TI_5$ ,  $TI_6$ ,  $TI_7$  (Fig. 13) (presumed plesiomorphic character state: setae  $TI_1$  and  $TI_2$  on tibia are similar to setae  $TI_3$ ,  $TI_4$ ,  $TI_5$ ,  $TI_6$ ,  $TI_7$ ). This character state is shared by larvae of the genera *Perileptus* + *Thalassophilus* and those of the genus *Aepopsis* (unpublished data).

#### BIONOMICS

Little is known about the bionomics of *Perileptus* larvae. The majority of larval material in our hands was reared in Petri-dishes - neither of us collected larvae in nature. During the two years that we kept adults on a soil substrate (granules of soil taken under broad-leaved forest with diameter ranging from 2 to 5 mm) we had no success in obtaining larvae. We reared many larvae as soon as we changed the soil substrate for the moist sand and fine alluvium from the river's beach where the adults were captured.

## Perileptus areolatus (Creutzer, 1799)

MATERIAL: nearly 30 specimens reared ex ovo (6L<sub>1</sub>, 2L<sub>2</sub>, 2L<sub>3</sub> were mounted on permanent microscope slides), from adults collected 28. V. 1996, Goriachiy Klyuch, West Caucasus, Russia, V. Grebennikov. 1L<sub>1</sub> (BMNH), labelled «Coll. V.E.», «b.VIII-35», «Perileptus 21.VII-35», «F.I. van Emden Collection B.M. 1977-248». 1L<sub>3</sub> (BMNH), labelled «Larvae (1?) Perileptus ?areolatus Cr. MPL 92», «F.I. van Emden Collection B.M. 1977-248».

DIAGNOSIS. First instar larvae hardly differ from *P. mesasiaticus* with less developed microsculpture on parietale; older instar larvae can not be separate from those of the second known species.

DESCRIPTION. With features of the genus.  $L_1$ : HW - 0.22 mm, HL - 0.23 mm (n=1);  $L_2$ : HW -0.28 mm, HL - 0.30 mm (n=1);  $L_3$ : HW - 0.38 mm, HL - 0.47 - 0.49 mm (n=2). First instar larvae with less developed microsculpture on parietale dorsally; ventral surface of parietale without microsculpture.

# Perileptus mesasiaticus Uéno, 1976

MATERIAL: nearly 15 specimens reared ex ovo (2L<sub>1</sub>, 2L<sub>2</sub> were mounted on permanent microscope slides), from adults collected 9. V. 1996, Dere-Dere stream, west slope of Kugitang Mountain Range, West Turkmenia, V. Grebennikov.

DIAGNOSIS. First instar larvae differ from those of *P. areolatus* by having the parietale almost fully covered with microsculpture.

DESCRIPTION. With features of the genus.  $L_1$ : HW - 0.23 mm, HL - 0.23 mm, (n=1);  $L_2$ : HW - 0.30 mm, HL - 0.31 - 0.32 mm (n=2). First instar larvae with well developed microsculpture on parietale dorsally; ventral surface of parietale with microsculpture.

#### Conclusion

We want to reemphasize the following. First, the genera *Perileptus* and *Thalassophilus* are likely highly derived members of the supertribe Trechitae since they share a number of apomorphic character states (at least three of them are autapomorphic). Second, they exhibit no evident relationships with the tribe Trechini (the group, to which the genera *Perileptus* and *Thalassophilus* belong according to imaginal classification), or with any other members of the supertribe. Probably, *Perileptus* and *Thalassophilus* could be more appropriately considered as a distinct evolutionary line because their larvae are so derived and isolated. However, because this study is based on a limited number of Trechitae taxa, we refrain from proposing any taxonomic changes to the current classification.

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#### REFERENCES

- ARNDT E. 1991. Familie Carabidae. In Klausnitzer B. (Ed). Die Larven der Käfer Mitteleuropas. Band 1: Adephaga. Goecke & Evers, Krefeld.: 45-141.
- BAEHR M. 1987. The Australian species of the carabid genus *Perileptus* (Coleoptera: Carabidae: Trechinae). Invertebrate Taxonomy, 1: 1-16.
- BOLDORI L. 1936. Larve dei Trechini 6. Studi Trentini di Sci. Natur., 17: 65-71.
- BOUSQUET Y. 1985. Morphologie comparée des larves de Pterostichini (Coleptera: Carabidae):

  Description et tables de détermination des espèces du Nord-Est de l'Amérique du Nord.

  Le Naturaliste Canadien, 122: 191-251.
- BOUSQUET Y. 1996. Description of the larva of Schizogenius lineolatus (Coleoptera: Carabidae: Clivinini). Acta Societatis Zoologicae Bohemicae, 60 (4): 347-353.
- BOUSQUET Y., GOULET H. 1984. Notation of primary setae and pores on larvae of Carabidae (Coleoptera: Adephaga). Canadian Journal of Zoology, 62: 573-588.
- CASALE A., LANEYRIE R. 1982. Trechodinae et Trechinae du monde: tableau des sous-familles, tribus, séries phylétiques, genres, et catalogue général des espèces. Memoires de biospéléologie, 9: 1-226.
- EMDEN F.I. van. 1942. A key to the genera of larval Carabidae (Coleoptera). Transactions of the Royal Entomological Society of London, 92: 1-99.
- GREBENNIKOV V.V. 1996. Description of the first instar larva of Thalassophilus longicornis (Coleoptera: Carabidae: Trechodina). Acta Societatis Zoologicae Bohemicae, 60 (4): 373-379.
- GREBENNIKOV V.V. 1997. Larvae of Bembidiini: subgenera Synechostictus and Pseudolimnaeum of the genus Bembidion and their taxonimic position (Coleoptera: Carabidae). Zoosystematica Rossica, 5 (2): 263-272.
- KRYZHANOVSKIJ O.L. 1976. An attempt of a revised classification of the family Carabidae (Coleoptera). Entomol. obozr., 55 (1): 80-91 [In Russian].
- KRYZHANOVSKIJ O.L. 1983. The beetles of the suborder Adephage: families Rhysodidae, Trachypachidae, Carabidae (introduction and a review of the USSR fauna). Fauna USSR, 1 (2). Leningrad, Nauka. 341 p. [in Russian].
- KRYZHANOVSKIJ O.L., BELOUSOV. I. A., KABAK I.I., KATAEV B.M., MAKAROV K.V., SHILENKOV V.G. 1995. A Checklist of the ground-beetles of Russia and adjacent lands (Insecta: Coleoptera: Carabidae). Pensoft Publishers, Sofia-Moscow. 271 p.
- LARSSON S.G. 1968. Lobebillernes larver. In Hansen V (ed).: Biller XXIV. Sandspringere og lobebiller (Cicindelidae og Carabidae). Danmarks Fauna, 76. Gads Forlag, Kobenhavn.: 282-433.
- LUFF M.L. 1985. The larvae of the British Carabidae (Coleoptera) VII Trechini and Pogonini. Entomologist's Gazette, 36: 177-194.
- LUFF M.L. 1993. The Carabidae (Coleoptera) larvae of Fennoscandia and Denmark. Fauna Entomologica Scandinavica, 27: 1-187.
- MAKAROV K.V. 1994. A key to the genera of the ground-beetle larvae (Coleoptera, Carabidae) of the Palaearctic Region. Bollettino del Museo Regionale di Scienze Naturali Torino, 12 (1): 221-254.
- SHAROVA I.Kh. 1958. The larvae of Carabidae beneficial and noxious to agriculture. Uchenye zapiski Moskovskogo Gosudarstvennogo Pedagogicheskogo Instituta Imeni Linina, 124: 1-165 [in Russian].

SHAROVA I.Kh. 1964. Family Carabidae. In Gylarov M.S. (ed): Opredelitel' obitaushikh v pochve lichinok nasekomykh. Muscow, Nauka Press: 112-195 [in Russian].

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