

***Renefouqueosis peruviansis*, a new genus and species
of Stenosini (Coleoptera: Tenebrionidae) from Peru
with a key to the Stenosini of the World and notes
on the genera *Anchomma* and *Fitzsimonsium***

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Abstract. *Renefouqueosis peruviansis* gen. et sp. nov., a new tenebrionid genus and species of the tribe Stenosini (subtribe Stenosina) is described from the arid mountains of Northern Peru. Including the new genus *Renefouqueosis* gen. nov., the tribe Stenosini now includes 40 valid genera of which nine are from the New World. The genera are placed in six subtribes (two worldwide, two New World and two Old World). Type species and subtribal assignment for each genus is presented. Notes on the placement of the genera *Anchomma* LeConte, 1858 and *Fitzsimonsium* Koch, 1962 are given. The proper placement of these genera is uncertain. Because of numerous morphological similarities to the Stenosini, we have decided to place these in a key to the world genera of Stenosini, which we provide.

Key words. Coleoptera, Tenebrionidae, Stenosini, new genus, new species, key to genera of world, Peru, Neotropical Region

Introduction

When René Fouquè, expert on the tribe Stenosini, visited the first author's collection, along with other tenebrionid colleagues, before the Third International Tenebrionoidea Symposium in Tempe, Arizona, USA in 2013, he confirmed that a small series, identified by the first author in 1995 as a new genus of Stenosini: *Stenosina*, was indeed

undescribed. Tentative plans were made to describe this new genus and species with René in the near future. This was not to be. This new genus and species is described here and dedicated to his memory.

Two major keys have been published for the Stenosini. The first by REITTER (1916) for the Palearctic fauna. Included were three subtribes and 12 genera which composed the then known genera of Stenosini of Europe, Middle East and Central Asia. He did not include tropical Asian, African or New World fauna known at that time. MEDVEDEV (1994) was the first to provide a key to the world Stenosini genera and subgenera composed of 30 genera and 129 subgenera. He did not include five, at that time existing genera of Stenosini, two of which (*Typhlusechus* Linell, 1897 and *Hexagonochilus* Solier, 1851) he excluded based on the position of apices of setae on the upper surface of the head which are directed posteriorly, not anteriorly as all other known Stenosini, a character he believed was important in distinguishing Stenosini. Subtribes were not included. Since 1994, five new genera and numerous species have been described. Additional keys to groups of genera were provided by FOUQUÉ (2008, 2013, 2015). Three then existing genera have also been re-diagnosed, and three new subgenera were added, by FOUQUÉ (2015). In the hopes that future researchers will continue his work on the Stenosini, a generic key to the tribe is also provided in his honor.

Materials and methods

Images of specimens or characters were taken using a Passport Imaging system (R. Larimer, www.visionarydigital.com). Montaged images were assembled using Zerene Stacker (zerenesystems.com/stacker/) and backgrounds were cleaned up in Adobe Photoshop CS. The spelling and authorship of family-group names follows data presented in BOUCHARD et al. (2011).

Material for this study was borrowed from the following individuals and institutions. These persons (in parentheses) are gratefully acknowledged for loan of their materials:

CASC California Academy of Sciences, San Francisco, California, USA (Dave Kavanaugh).

RLAC Rolf L. Aalbu collection, El Dorado Hills, California, USA.

Description of a new genus and species

Renfouqueosis gen. nov.

Type species. *Renfouqueosis peruviansis* sp. nov. by present designation.

Composition. The genus includes only the type species, *Renfouqueosis peruviansis* sp. nov.

Description. Body relatively small (around 3 mm). Head without dorsal keels, with antennomeres completely separated; eyes positioned above and anterior to an antero-lateral extended genal keel, composed of an elongate series of facets. Pronotum widest at anterior third, 1.125 times wider than long, cordate, anterior margin concave, posterior margin truncate, with two keels. Scutellum small, triangular. Elytra with keels on intervals 3, 5 and 7.

Comparative diagnosis. *Renefouqueosis* gen. nov. (Stenosini: Stenosina) can be distinguished from all other genera of New World Stenosini (subtribe Stenosina) by the following characters: (1) the antennomeres are completely separated (*Ecnomoderes* Gebien, 1928 as well as *Grammicus* Waterhouse, 1845, which also occurs in Southern Peru has the antennomere XI embedded in the X apically); (2) pronotum with coarsely serrate lateral margins and (3) the presence of two keels on the central aspect of the

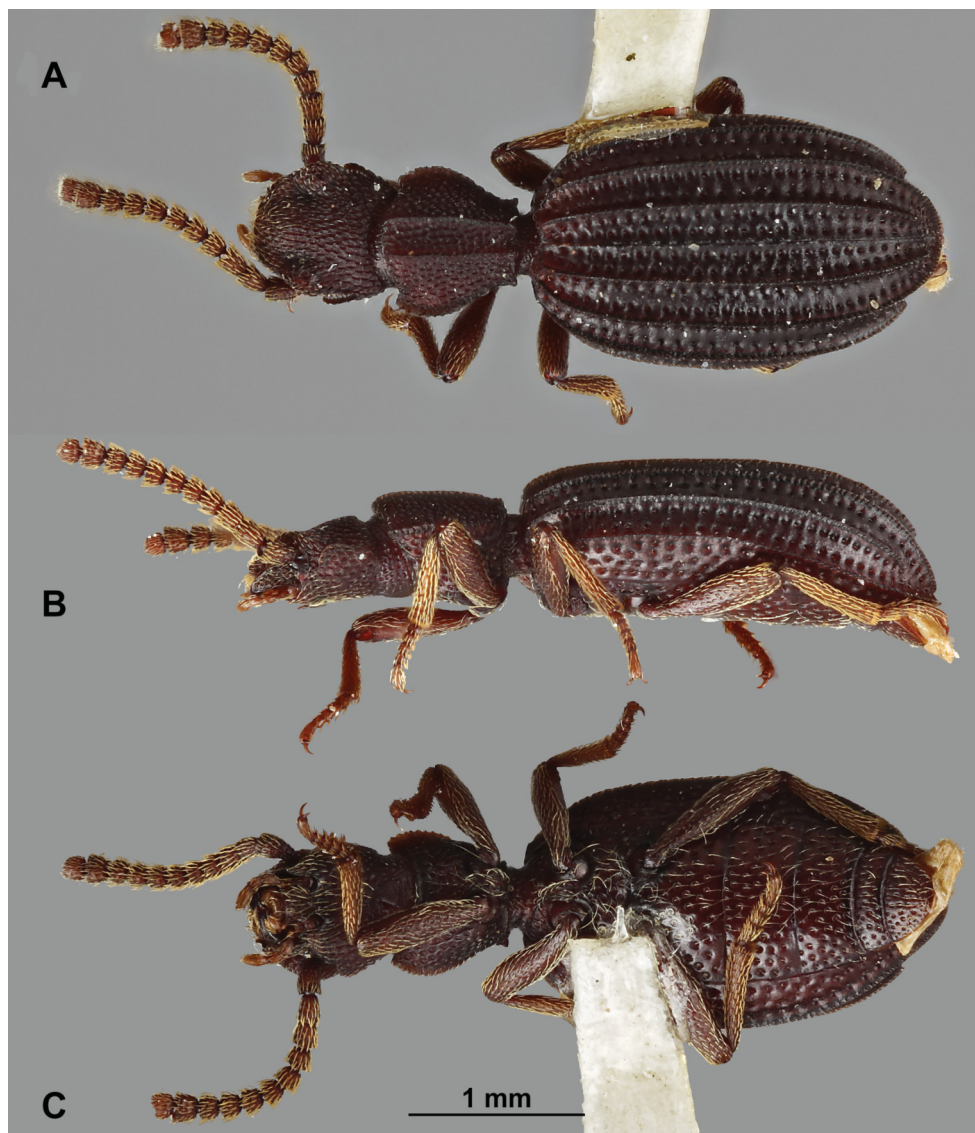
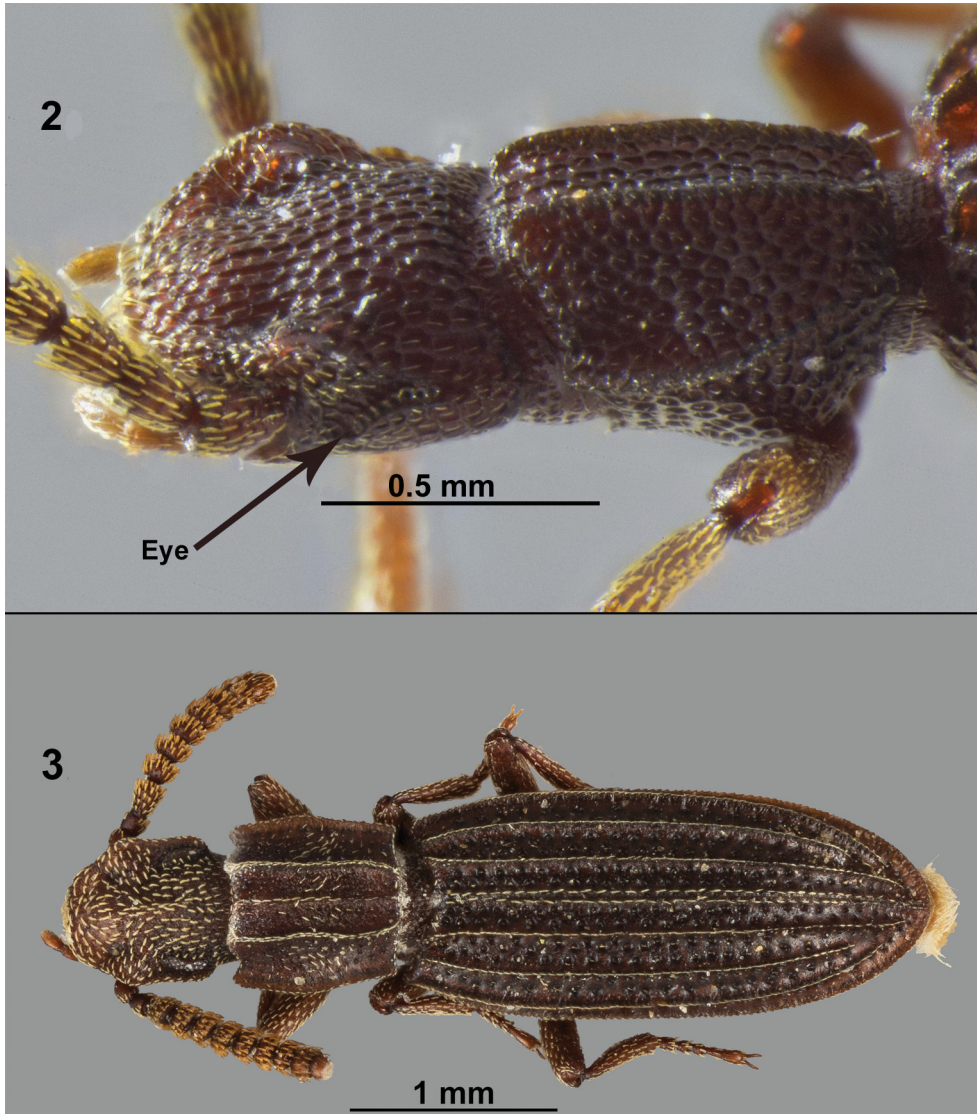


Fig. 1. *Renefouqueosis peruvienensis* gen. et sp. nov., habitus female. Dorsal (A) lateral (B) and ventral (C) views.

pronotum (*Caribanosis* Nabozhenko et al., 2016 has a single keel centrally positioned on the pronotum while, *Schizaraeus* Kulzer, 1955, lacks keels). In Old World Stenosini (subtribe Stenosina), of the genera which have both pronotal and elytral keels, but not head keels, which includes *Anethas* Jakobson, 1924, *Gebieniella* Koch, 1940, *Sten-*



Figs 2–3. 2 – *Renefouqueosis peruviansis* gen. et sp. nov. (female), dorso-lateral view of head showing eye. 3 – *Anchomma costatum* LeConte, 1858, dorsal habitus.

oethas Kaszab, 1975, and *Tetranosis* Medvedev, 1995, all have the pronotum clearly longer than wide, sometimes much longer than wide. In *Caribanosis* the pronotum is slightly wider than long (1.05 : 1.00). *Ethas* Pascoe, 1862 and *Perdicus* Fairmaire, 1899 have pronotal elevations but in these the elevations are rounded, not keeled. In *Renefouqueosis* gen. nov., the pronotum is clearly wider than long (ratio 1.125 : 1.000). The wide pronotum with serrate margins and of the eye which is composed of an elongate series of facets positioned above an antero-lateral extended genal keel, rounded anteriorly, visible from above only at posterior, rest of eye facets, not separated by a furrow, facing anteriorly, not visible from above due to genal extension above antennal insertion and not visible from below due to lateral genal extension is also unique.

***Renefouqueosis peruviansis* sp. nov.**

(Figs 1a–c, 2)

Type locality. Peru, Lambayeque Region, 98 mi E of Olmos.

Type material. HOLOTYPE: ♂, 'PERU: 98 mi. E. Olmos Lambayeque, I-19-1955, E. I. Schlinger & E. S. Ross collectors' (CASC). PARATYPE: ♀, 'PERU: 98 mi. E. Olmos Lambayeque, I-19-1955, E. I. Schlinger & E. S. Ross collectors. Tenebrionid Base, Aaron D. Smith, Catalog # 16893' (RLAC).

Description. Holotype, male: length 3.0 mm, width 1.6 mm. Integument matt, glabrous, heavily punctate, dark reddish brown.

Head broadened from base to genae lateral to eyes, widest at level lateral to eyes. Surface covered with broad, shallow punctures, each bearing a short seta, setae becoming slightly longer at apex. Eyes composed of an elongate series of facets positioned above an antero-lateral extended genal keel, rounded anteriorly, visible from above only at posterior, rest of eye facets facing anteriorly, not visible from above due to genal extension above antennal insertion. Antennae 11-segmented, antennomeres I–III longer than wide, IV–X wider than long, XI longer than wide. Antennomere (refer to Fig. 1) X larger than IX, antennomere XI subequal in width to antennomere IX but longer in length, width. Antennomere XI smaller than X, completely separated from antennomere X. Mentum trapezoidal, weakly transverse, apex wider than base, anteriorly concave, with sparse elongate setae; ligula visible, small. Maxillary palps with stipes, palpifer visible, apical segments elongate oval. Labial palps small. Gular region strongly produced anteriorly around mentum, anterior angles acute, apex between angles arcuate, convex. Gular sides (temples) with antero-lateral plates which covers eyes from below. Lower surface of head covered with yellow setae, longer medially.

Pronotum 1.125 times wider than long, about equally as long as head, cordate, sharply reflexed at posterior lateral angles, forming a triangular shaped spike, with longitudinal keels at about 1/5 distance to lateral margin at broadest point. Antero-lateral margins expanded, with anterior angles protruding slightly. Anterior margin rounded, posterior margin nearly truncate. Lateral margins serrate. Surface covered with broad, shallow punctures, each bearing a short seta. Lateral sides of disc clearly flattened. Lower surface strongly punctate with few short setae laterally, longer medially.

Scutellum very small, triangular.

Elytra broadly rounded, with weakly expressed rounded humeral angles, base concave before scutellum, surface deeply serially punctate, punctures with small short seta. Intervals 3, 5, and 7 forming a sharp keel; keels 3 and 7 terminate before apex; keel 5 reaching and meeting at apex of abdomen. Elytral suture also slightly elevated.

Ventral surface (Fig. 1C) with prosternal process narrowing behind coxae, strongly punctate, each puncture bearing moderately long yellow setae. Abdomen as in Fig. 1C, with intercoxal process of abdomen nearly parallel, truncate at apex. Surface strongly punctate bearing short setae laterally, longer medially.

Legs relatively short. Tibiae and femora straight, setose. Femora broader than tibiae. Tibia expanded gradually to apex. Abdomen punctured as elytra.

Male genitalia not dissected but genitalia partly exposed externally. Aedeagus inverted with parameres narrowing at apex, split apically.

Female genitalia not dissected but genitalia partly exposed externally (see Fig. 1) with coxites total length approximately equal to length of last visible abdominal segment. Gonostyli small, sub-apical with setae at apex.

Distribution. Peru: Lambayeque.

Remarks. The area mentioned on the label puts this collecting area in the dry, rain shadow areas in the mountains of Northern Peru, probably at a mid-elevation.

Discussion on the genera *Anchomma* and *Fitzsimonsium*

Anchomma costatum LeConte, 1858 (Fig. 3) was described by LECONTE (1858) and placed in various tribes of Colydiinae (Zopheridae) based on its 4 : 4 : 4 tarsal count and lack of visible membranes between abdominal ventrites. DOYEN & LAWRENCE (1979) transferred this genus to Stenosini based on the following characters: ‘antennae thick, moniliform; eyes divided by epistomal canthus; mandibles with mola transversely narrow, lunate; labrum rectangular, with medial tormal processes directed posterad; tentorium with sides long, subparallel or gradually converging anteriorly, with simple transverse bridge located anterad of middle; mesendosternite with long, anterior arms terminating in muscle discs and long, slender dorsal arms; and metendosternite with short, thick stalk and long, slender, tapering arms’. They suggested a close relationship to *Grammicus*, noting that in both genera ‘the mesocoxal cavities are almost closed by the sterna, which are separated by a very narrow groove. In *Stenosis* and *Araeoschizus* the mesocoxal closure by the sterna is complete’. Later DOYEN (1993) moved *Anchomma* to the tribe Anepsiini without suggesting close relationship with other members of that tribe. Doyen analyzed pimeliine beetles using three different outgroups (*Belopus* Gebien, 1911 (a lagriine); *Zolodinus* Blanchard, 1853 (a zolodinine) and what he considered to be all ‘primitive’ character states for the family. Characters used to separate these tribes were mainly those of internal and external reproductive structures. In light of the molecular analyses in KANDA (2017), *Anchomma* should be excluded from Anepsiini, but its taxonomic affinities are still uncertain. Because of its morphological similarities to Stenosini (see above), *Anchomma* (Fig. 3) is included in the key to the world genera

of Stenosini, but at present should be considered as *incertae sedis* within Pimeliinae.

ENDRÖDY-YOUNGA (1996) transferred the monotypic genus *Fitzsimonsium* Koch, 1962, from Caenocrypticini to Stenosini based on (1) moniliform antenna; (2) apically split parameres with fine setae; (3) very broad scutellum (three times broader than long); and (4) simply truncate apex of protibia. Both the first and fourth of these are generally considered plesiomorphic. Apically split parameres are found in some Stenosini but a very broad scutellum is not. However, besides a very broad scutellum (not mentioned in the redescription by ENDRÖDY-YOUNGA (1996: 45), *Fitzsimonsium* has the following character states not found in any Stenosini: (1) a stridulatory gula; and (2) a dilated anterior tibia, both common in Coenocrypticini. In ENDRÖDY-YOUNGA'S (1996: 11) cladogram, *Fitzsimonsium* is placed apically to *Boromorpha* Wollaston, 1854, currently in its own subtribe. It does have a similar body shape and antenna to members of the Stenosini subtribe Platamodina so it is here included in the key. However, the authors would like to emphasize that the tribal placement of *Fitzsimonsium* is in need of further study.

Including the new genus *Renefouqueosis* gen. nov., the tribe Stenosini now includes 40 valid genera of which nine are from the New World. A key to the known subtribes and genera is presented, and includes the unplaced genera *Anchomma* and *Fitzsimonsium*. The genera are placed in six subtribes (two worldwide, two New World and two Old World; see Table 1). Type species and subtribal assignments for each genus are also included (Table 2). A few of the genera, due to their varied morphology, key out in more than one couplet. Including images or photos of each of these genera would be difficult, as many are very rare in collections. Instead readers are referred to a footnote in the key for images, photos, and/or additional diagnostic information. Also refer to LÖBL et al. (2008) for Palearctic species. Subgenera have not been included here unless the specific subgenus distinguishes this group from other subgenera which may be grouped together in the key, separated in another couplet. For further subgeneric separations, see MEDVEDEV (1994) and FOUQUÉ (2008, 2013, 2015).

Table 1. Subtribes of the Stenosini.

Subtribe	Author, year	Comment
Stenosina	Schaum, 1859: 66 (1834)	proposed as a subtribe by REITTER (1916: 137)
Dichillina	Reitter, 1916: 137	
Araeoschizina	Casey, 1907: 484	proposed as a subtribe by DOYEN & LAWRENCE (1979: 351)
Typhlusechina	Casey, 1907: 494	proposed as a subtribe by AALBU & ANDREWS (1985: 1)
Platamodina	Reitter, 1900: 137	
Harvengiina	Ferrer, 2004: 370	

Key to subtribes and genera of world Stenosini (plus *Anchomma*)

- 1 Antennae short, slender, almost compact, the antennomeres scarcely separated; legs short, all femora not or barely exceeding lateral margin of body. **2 (Platamodina)**
- Antennae with normal articulation; legs longer, all femora as long as or exceeding lateral margin of body. 4
- 2 Body longer; pronotum with basal angles rounded, convex, not closely joined to elytra. ***Microblemma* Semenow, 1890**
Six species in Afghanistan, Turkmenistan.
- Body wider; pronotum not or barely smaller than elytra, not exceeding margin of body. 3
- 3 Pronotum with basal angles angulate, concave, closely joined to elytra; femora untoothed. ***Fitzsimonsium* Koch, 1962**
Monotypic: *F. cymbium* (Koch, 1962) in Namibia.
- Pronotum with basal angles rounded not closely joined to elytra; all femora with a tooth. ***Platamodes* Ménétriés, 1849**
One species and two subspecies in Central Asia, Transcaucasia.
- 4 Eyes completely absent, size very small, less than 1.4 mm. ***Harvengina: Harvengia* Ferrer, 2004**
Monotypic: *H. vietnamita* Ferrer, 2004 from Vietnam.
- Eyes with at least four facets present, size usually much larger but at least larger than 1.6 mm. 5
- 5 Eyes entirely divided by lateral margin of head into upper and lower sections, sections not connected by a connecting furrow. 6
- Eyes whole or not entirely divided, reduced medially, sometimes upper and lower regions appearing divided but connected by a connecting furrow. **22 (Stenosina)**
- 6 Upper and lower eyes further divided but connected by a connecting furrow. ***Typhlusechina: Typhlusechus* Linell, 1897**
Five species in U.S. and Mexico.
- Upper and lower eyes not further divided. 7
- 7 Body covered with numerous squamiform setae. ***Araeoschizina: Araeoschizus* LeConte, 1861**
Fifty-two species in U.S. and Mexico.
- Body bare or with simple setae only. **8 (*Dichillina* and *Anchomma*)**
- 8 Pronotum and elytra sharply costate (keeled). 9
- Pronotum and elytra not sharply costate, at most with rounded elevations. 14
- 9 Head sharply costate. 10
- Head not costate. 11
- 10 Body small, less than 3 mm long. ***Discopleurus* Lacordaire, 1859**
Six species in the Neotropics.
- Body larger, length greater than 3 mm. ***Microtelus* Solier, 1838**
Seventeen species and subspecies, in the Mediterranean Region, the Middle East and Central Asia.
- 11 Both pronotum and elytra costate (Fig. 3). ***Anchomma* LeConte 1858**
Monotypic: *A. costatum* LeConte, 1858 in U.S.A.: California, Nevada.
- Pronotum not costate. 12

- 12 Elytral base equal in width to pronotal base. ***Pseudochillus*** Fouquè, 2015
Seven species in India, Andaman Islands, Myanmar, Thailand, Vietnam, Laos, and Philippines.
– Elytral base wider than pronotal base. 13
- 13 Tempora concave; occiput with sharply bordered triangular impression with mid-longitudinal keel. ***Indochillus*** Koch, 1941
Monotypic: *Indochillus cristatus* Koch, 1941 in India.
– Tempora convex; occiput without sharply bordered triangular impression and mid-longitudinal keel. ***Pseudethas*** Fairmaire, 1896
Twenty-one species in Afghanistan, Pakistan, India, Nepal, Tibet, and Thailand.
- 14 Sides of the pronotum and elytra distinctly crenate. 15
– Sides of the pronotum and elytra smooth. 16
- 15 Lateral margins of pronotum distinctly serrate; elytra broadest near midpoint, tapering arc-wise to apex. ***Oogaster*** Falderman, 1837
Two species in the Central Asia and Transcaucasia.
– Lateral margins of pronotum not serrate; elytra broadest at basal 1/3, tapering as a straight line to apex. ***Afghanillus*** Kaszab, 1960
Monotypic: *A. klapperichi* Kaszab, 1960 in Afghanistan.
- 16 Body broad, pronotum quadrate, sides reflexed, elytral intervals with serially rounded tubercles. ***Hexagonochilus*** Solier, 1851
Two species in Chile.
– Pronotum without lateral margins reflexed; elytra without serially rounded tubercles. 17
- 17 Abdomen broadest before 1/3 from base, evenly tapering toward apex; body smooth and shiny, tear shaped head, pronotum and abdomen, setae, punctures not apparent, very small. ***Reitterella*** Semenov, 1891
Three species in Turkmenistan and Uzbekistan.
– Abdomen broadest at or around midpoint, elytra often strongly punctate and setose. 18
- 18 Suborbital keels prominent. 19
– Suborbital keels reduced or absent. 20
- 19 Pronotum quadrate, sides rounded, pronotum wider than head.
..... ***Aspidocephalus*** Motschulsky, 1839
Monotypic: *A. desertus* Motschulsky, 1839 from Central Asia, Transcaucasia and south of European part of Russia.
– Pronotum elongate, head wider than pronotum. ***Dichillus*** Jacquelin du Val, 1861
Eighty two species in eight subgenera in the western and central Palearctic Region.
- 20 Suborbital keels reduced. ***Herbertfranzia*** Kaszab, 1973
Monotypic: *H. nepalica* Kaszab, 1973 from Nepal.
– Suborbital keels completely absent. 21
- 21 Base of elytra wider than base of pronotum (sometimes only slightly); elytral interval 9 forming humeral corner. ***Herbertfranziella*** Kaszab, 1973
Eight species in Nepal, Tajikistan, Pakistan, and India.
– Base of elytra as wide as base of pronotum; elytral interval 9 not forming humeral corner. ***Nepalofranziella*** Fouquè, 2013
Two species in Nepal.

- 22 Head with keels (costae) on top of head. 23
 – Head without keels on top. 25
- 23 Head with a central and lateral keels. **Tetranillus** Wasmann, 1899
 Four species in Iran, China and Oriental Region.
 – Head with lateral keels only. 24
- 24 Pronotum with keels; eyes large, positioned near front of keel.
 **Stenoethas** Kaszab, 1975
 Monotypic: *S. carinipennis* Kaszab, 1975 from India.
 – Pronotum without keels; eyes very small, positioned near back of keel.
 **Timosmithus** Ardoin, 1974
 Monotypic: *T. basilewskyi* Ardoin, 1974 from South Africa.
- 25 Pronotum with keels. 26
 – Pronotum without keels. 34
- 26 Pronotum with single central keel. **Caribanosis** Nabozhenko et al., 2016
 Monotypic: *C. quisqueyanus* (Garrido & Varela, 2011) from Hispaniola.
 – Pronotum with two keels. 27
- 27 Antennomere I as long as II and III combined. **Anethas** Jakobson, 1924 (partim)
 Eleven species in Madagascar and South Africa.
 – Antennomere I not longer than II and III. 28
- 28 Antennomere X larger than IX, XI very small positioned inside apex of X. 29
 – Antennomere XI equal in size to IX, not positioned inside apex of X. 30
- 29 Base of pronotum and apex of elytra with dense pads of yellow setae.
 **Ecnomoderes** Gebien, 1928
 Two species in Argentina.
 – Base of pronotum and apex of elytra without dense pads of yellow setae.
 **Grammicus** Waterhouse, 1845
 Six species and subspecies in Chile and Peru.
- 30 Antennomere III equal to next four segments combined. **Schizaraeus** Kulzer, 1955
 Monotypic: *S. acuticosta* Kulzer, 1955 from Argentina.
 – Antennomere III not equal to next four segments combined. 31
- 31 Pronotum wider than long. **Renefouqueosis** gen. nov.
 Monotypic: *R. peruviensis* sp. nov. from Peru.
 – Pronotum longer than wide. 32
- 32 Elytra with three keels. **Gebieniella** Koch, 1940
 Four species and one subspecies in Myanmar, Thailand, Philippines,
 China (Hainan Is.), Vietnam, and Indonesia (Java).
 – Elytra with at least four partial or whole keels. 33
- 33 Head with apex inflated and strongly grooved in middle. ... **Ethas** Pascoe, 1862 (partim)
 Eleven species in India, Indochina, and Sri Lanka.
 – Head with apex simple or slightly inflated, not strongly grooved in middle.
 **Tetranosis** Medvedev, 1995
 Ten species in five subgenera in North India, Pakistan, Nepal, and China (Xizang).
- 34 Elytra with keels, head, pronotum rugose, with rounded elevations.
 **Perdicus** Fairmaire, 1899
 Monotypic: *P. anthrophilus* Fairmaire, 1899 from Madagascar.

- Elytra usually without keels; head and pronotum not rugose, or with rounded elevation. 35
- 35 Pronotum and elytra with rounded elevations. 36
- Pronotum and elytra without rounded elevations. 38
- 36 Head with apex inflated and grooved in middle. *Ethas* Pascoe, 1862 (partim)
Eleven species in India, Indochina, and Sri Lanka.
- Head with apex simple or slightly inflated, not strongly grooved in middle. 37
- 37 Head with anterior margin distinctly asymmetric, with left half slightly concave, right half slightly convex, with groove behind eyes; lower aspect of eyes truncated by lateral angulate expansion of genae behind eyes, reduced to few ocelli arranged in strip.
..... *Stenosis* subgen. *Burmanosis* Medvedev, 1995
Four species in Myanmar.
- Head with anterior margin not asymmetric, without groove behind eyes, rounded; lower aspect of eyes not truncated by lateral angulate expansion of genae.
..... *Anethas* Jakobson, 1924 (partim)
Eleven species in Madagascar and South Africa.
- 38 Antennomere I as long as II and III combined; pronotum U-shaped with base round and apex truncate. *Itampolis* Koch, 1962
Monotypic: *I. oceanica* Koch, 1962 from Madagascar.
- Antennomere I shorter than II and III combined; pronotum not U-shaped. 40
- 39 Elytra at humeral base abruptly expanded forming angle at humerus; eyes not visible from above. *Tagenostola* Reitter, 1916
Four species in Transcaucasia, south of European part of Russia, Central Asia, North Africa, the Middle East, Laos, and Myanmar.
- Elytra at humeral base gradually expanded, not forming angle at humerus; eyes visible from above. 40
- 40 Lower aspect of eyes truncated by lateral angulate expansion of genae, reduced to few ocelli arranged in strip. 41
- Genae not expanded laterally behind eyes, not truncating lower aspect of eyes to a few ocelli arranged in a strip. 42
- 41 Apical antennomere (XI) almost always smaller than X. *Stenosis* Herbst, 1799
Hundred-and-twenty-one species and subspecies in four other subgenera in Africa, Central Asia, Mediterranean.
- Apical antennomere (XI) as wide, longer than X. *Eutagenia* Reitter, 1886
Eleven species and subspecies in the Mediterranean area and Central Asia.
- 42 Head large, as broad as elytra, wider than pronotum. *Mitotagenia* Reitter, 1916
Ten species and subspecies in Africa (Afrotropical Region), the Middle East and Afghanistan.
- Head not as wide as elytra. 43
- 43 Pronotum broad with lateral apical angles produced, rounded.
..... *Schusteriella* Koch, 1940
Monotypic: *S. ruficornis* (Reitter 1886) from Ethiopia.
- Pronotum narrow, with lateral apical angles not produced, angulate.
..... *Indostola* Medvedev, 1991
Two species in India and Thailand.

Table 2. Genera, type species and subtribal assignments of Stenosini.

Genus	Type Species	Fixed by	Subtribe	References
<i>Afghanillus</i> Kaszab, 1960: 1	<i>Afghanillus klapperichi</i> Kaszab, 1960	original designation	Dichillina	see KASZAB (1960: Fig. 18; 1969b: pl. 6, Fig. 7)
<i>Anethas</i> Jakobson, 1924: 242	<i>Pseudethas longiceps</i> Fairmaire, 1898	monotypy	Stenosina	see FERREIRA (1950, 1952); KOCH (1962)
<i>Araeoschizus</i> LeConte, 1861: 138	<i>Araeoschizus costipennis</i> LeConte, 1851	monotypy	Araeoschizina	see PAPP (1981)
<i>Aspidocephalus</i> Motschulsky, 1839: 63	<i>Aspidocephalus desertus</i> Motschulsky, 1839	monotypy	Dichillina	see REITTER (1916); ABDURAKHIMANOV & NABOZHENKO (2011)
<i>Caribanosis</i> Nabozhenko et al., 2016: 568	<i>Rhyasma quisqueyanus</i> Garrido & Varela, 2011	original designation	Stenosina	see NABOZHENKO et al. (2016)
<i>Dichillus</i> Jacquelin du Val, 1861: 253	<i>Tagenia minuta</i> Solier, 1838	original designation	Dichillina	see REITTER (1916); KASZAB (1960, 1969b, 1975, 1981); FOUQUÉ (2015: 240); MEDVEDEV (1975, 1977, 2008); MEDVEDEV & NEPEŠOVA (1985); ABDURAKHIMANOV & NABOZHENKO (2011)
<i>Discopleurus</i> Lacordaire, 1859: 105	<i>Pleurophorus quadricollis</i> Solier, 1851	monotypy	Dichillina	see AALBU & ANDREWS (1996)
<i>Ecnomoderes</i> Gebien, 1928: 109	<i>Ecnomoderes barbatus</i> Gebien, 1928	subsequent designation	Stenosina	see GEBIEN (1928: pl. II, Figs 2 & 3)
<i>Eithas</i> Pascoe, 1862: 324	<i>Eithas carbonarius</i> Pascoe, 1862	subsequent designation	Stenosina	see KOCH (1940: pl. XX, Fig. 1 & 2); KASZAB (1981)
<i>Eutagenia</i> Reitter, 1886: 125	<i>Stenosia smymensis</i> Reitter, 1889 (= <i>Eutagenia cribricollis</i> Reitter, 1916)	subsequent designation	Stenosina	see REITTER (1916); see internet for photo
<i>Fitzsimonsium</i> Koch, 1962: 152	<i>Fitzsimonsia cymbium</i> Koch, 1955	original designation	Platamodina	see ENDRÖDY-YOUNG (1996: Fig. 3D)
<i>Gebieniella</i> Koch, 1940: 736	<i>Eithas stenoides</i> Pascoe, 1862	original designation	Stenosina	see KOCH (1940: pl. XIX, Fig. 5); MEDVEDEV (2009)
<i>Grammicus</i> Waterhouse, 1845: 323	<i>Grammicus chilensis</i> Waterhouse, 1845	monotypy	Stenosina	see KASZAB (1969a: p. 307, Figs 4 & 5)
<i>Harvengia</i> Ferrer, 2004: 367	<i>Harvengia vietnamita</i> Ferrer, 2004	original designation	Harvengiina	see FERRER (2004: Figs 1 & 2)

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Genus	Type Species	Fixed by	Subtribe	References
<i>Herbertfranzia</i>	<i>Herbertfranzia nepalica</i> Kaszab, 1973	original designation	Dichillina	see FOUQUÉ (2013; Fig. 11)
<i>Herbertfranzietta</i>	<i>Herbertfranzietta eutagenoides</i> Kaszab, 1973	original designation	Dichillina	see FOUQUÉ (2013)
<i>Hexagonochilus</i>	<i>Hexagonochilus dilatocollis</i> Solier, 1851	original designation	Dichillina	see internet for photo
<i>Indochillius</i>	<i>Indochillius cristatus</i> Koch, 1941	monotypy	Dichillina	see KOCH (1941; Fig. 3); KASZAB (1981; Figs 19–20); FOUQUÉ (2015; Fig. 5)
<i>Indostola</i>	<i>Indostola puchella</i> Medvedev, 1991	original designation	Stenosina	see FOUQUÉ (2015)
<i>Itampolis</i>	<i>Itampolis oceanica</i> Koch, 1962	original designation	Stenosina	see KOCH (1962; Fig. 5).
<i>Microblemma</i>	<i>Microblemma simplex</i> Semenov, 1889	monotypy	Platamodina	see REITTER (1916); KASZAB (1960: pl. 2, Fig. 19; 1969b: pl. 7); MEDVEDEV (2008)
<i>Microtelus</i>	<i>Microtelus asiaticus</i> Solier, 1838	original designation	Dichillina	see CARL (1992); MEDVEDEV (2008)
<i>Mitotaqenia</i>	<i>Stenosis arabs</i> Baudi di Selve, 1881	monotypy	Stenosina	see FERRIERA (1950; Fig. 3; 1952)
<i>Nepalofranziella</i>	<i>Nepalofranziella kaszabi</i> Fouqué, 2013	original designation	Dichillina	see FOUQUÉ (2013)
<i>Oogaster</i>	<i>Oogaster menetriesii</i> Faldermann, 1837	monotypy	Dichillina	see REITTER (1916); KASZAB (1969b: pl. 6, Fig. 11); ABDURAKHMANOV & NABOZHENKO (2011)
<i>Pardicus</i>	<i>Pardicus anthrophilus</i> Fairmaire, 1899	monotypy	Stenosina	see KOCH (1943)
<i>Platamodes</i>	<i>Platamodes dentipes</i> Ménétriés, 1849	monotypy	Platamodina	see REITTER (1916); KASZAB (1959: pl. 2, Fig. 20; 1969b: pl. 8)
<i>Pseudethas</i>	<i>Pseudethas quadraticeps</i> Fairmaire, 1896	monotypy	Dichillina	see KASZAB (1969b: pl. 5, Fig. 12); FOUQUÉ (2008); BA & REN (2014; Figs 7–12); ABDURAKHMANOV & NABOZHENKO (2011)
<i>Pseudochillius</i>	<i>Indochillius bangaloreanus</i> Kaszab, 1981	original designation	Dichillina	see FOUQUÉ (2015; Figs 6–16)

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Genus	Type Species	Fixed by	Subtribe	References
<i>Reitterella</i> Semenow, 1891: 362	<i>Reitterella fusiformis</i> Semenow, 1891	monotypy	Dichillina	see REITTER (1916); see internet for photo
<i>Renefouqueosis</i> gen. nov.	<i>Renefouqueosis peruvienis</i> sp. nov.	present designation	Stenosina	present paper
<i>Schizaraeus</i> Kulzer, 1955: 479	<i>Schizaraeus acuticosta</i> Kulzer, 1955	original designation	Stenosina	see KULZER (1955: Fig. 3)
<i>Schusteriella</i> Koch, 1940: 746	<i>Stenosis ruficornis</i> Reitter 1886	monotypy	Stenosina	see KOCH (1940: pl. XX, Fig. 8; 1956: Fig. 4)
<i>Stenoethas</i> Kaszab, 1975: 11	<i>Stenoethas carinipennis</i> Kaszab, 1975	original designation	Stenosina	see KASZAB (1975: Fig. 3)
<i>Stenosis</i> Herbst, 1799: 160	<i>Stenosis angustata</i> Herbst, 1799 (= <i>Stenosis intermedia</i> Kuster, 1848)	subsequent designation	Stenosina	see REITTER (1916); ANTOINE (1948, 1949); KOCH (1940, 1941, 1956: Fig. 8); KASZAB (1979, 1980, 1981); MEDVEDEV (1994, 2009)
<i>Tagenostola</i> Reitter, 1916: 151	<i>Stenosis turkestanica</i> Reitter, 1886	subsequent designation	Stenosina	see KOCH (1956: Fig. 2); FOUQUÉ (2015)
<i>Tetranillus</i> Wasmann, 1899: 167	<i>Tetranillus costatus</i> Wasmann, 1899	monotypy	Stenosina	see KOCH (1940: pl. 19, Fig. 4); BA & REN (2013: Fig. 1–6); REN & SHI (2006)
<i>Tetranosis</i> Medvedev, 1994: 858*	<i>Tetranosis clypeolobus</i> Koch, 1940	original designation	Stenosina	see KOCH (1940: pl. XX, Figs 3 & 4); KASZAB (1973: pl. 1, Figs 2 & 3; 1975; 1981)
<i>Timosmithus</i> Ardoïn, 1974: 457	<i>Timosmithus basilewskyi</i> Ardoïn, 1974	monotypy	Stenosina	see ARDOÏN (1974: Fig. 1)
<i>Typhlusechus</i> Linell, 1897: 154	<i>Typhlusechus singularis</i> Linell, 1897	original designation	Typhlusechina	see AALBU & ANDREWS (1985)

* The genus name *Tetranosis* was first introduced by KOCH (1940: 740). As pointed out by LOBL & MERKL (2003: 251) however Koch failed to designate a type species when he described his new genus. MEDVEDEV (1994: 858) was the first author to provide a description of *Tetranosis* as well as fixing a type species for the genus, thereby making the name nomenclaturally available for the first time.

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