# Viedebanttia egorovi sp. n., a new acarid mite from South Korea, with notes on the genus (Acariformes: Acaridae)

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Viedebanttia egorovi sp. n., a new species of acarid mite phoretic on the beetle Rhombonyx testaceipes from South Korea is described. The following species are transferred to the genus Viedebanttia Oudms., 1929: Tyroglyphus fuscipes Vitz., 1924; Robinisca macrocnemis Zachv., 1941; R. longipes Volgin, 1951; R. coniferae Sevastianov & Marroch, 1993 and Caloglyphus vitzthumi Mah., 1979.

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#### Introduction

Up to now the genus Viedebanttia Oudemans, 1929 was known from two species, V. schmitzi Oudemans, 1929 and V. diamanus Fain & Schwan, 1984. Examination of Dr. A. Egorov's entomological collection from South Korea (1997) has yielded a new species of this genus. In addition, five other species described in the genera Robinisca Zachvatkin, 1941, Tyroglyphus Latreille, 1796 and Caloglyphus Berlese, 1923 should be placed in Viedebanttia on the basis of Fain & Schwan's (1984) definition of the genus.

The terms of body parts and idiosomal chaetotaxy follow Griffiths et al. (1990); terms of leg chaetotaxy and solenidiotaxy follow Griffiths (1970) (except  $\omega_3$  which is replaced by *ba*). All measurements are given in micrometers ( $\mu$ m).

The holotype and the coleopteran host of the new species are kept in the Institute of Biology and Pedology, Vladivostok (IBPV).

## Genus Viedebanttia Oudemans, 1929

Viedebanttia: Zachvatkin, 1941: 218; Fain, 1985: 276.

- Robinisca: Zachvatkin, 1941: 215 (part.); Volgin, 1951: 36; Sevastianov & Marroch, 1993: 149; Bugrov, 1997: 152 (part.). Misidentifications.
- Acotyledon (non Oudemans, 1903; part.): Turk & Turk, 1957: 93; Černý & Samšińák, 1971: 508.
- Caloglyphus (non Berlese, 1923): Mahunka, 1979: 326 (part.).

Type species Viedebanttia schmitzi Oudemans, 1929, by original designation.

The genus Viedebanttia had been proposed for a single species, V. schmitzi (hypopus), by Oudemans (1929). Owing to insufficient original description, Zachvatkin (1941) considered this genus as incertae sedis. He had believed that Viedebanttia is closely related to Robinisca Zachvatkin, 1941. Turk & Turk (1957) described adults of V. schmitzi and placed this species in Acotyledon Oudemans, 1903, while Mahunka (1979) has synonymized Viedebanttia with Caloglyphus Berlese, 1923. Later, Fain & Schwan (1984) and Fain (1985) redescribed the holotype specimen of V. schmitzi and resurrected Viedebanttia as a valid genus.

The main features of the genus Viedebanttia are the enlarged tibiae of legs I (II), very short propodosoma, and long setae scx as Fain & Schwan (1984) defined. Based on this definiton, the following species should be transferred to the genus Viedebanttia: Tyroglyphus fuscipes Vitzthum, 1924; Robinisca macrocnemis Zachvatkin, 1941; Robinisca longipes Volgin, 1951; Robinisca coniferae Sevastianov & Marroch, 1993 and Caloglyphus vitzthumi Mahunka, 1979.

Turk & Turk (1957) synonymized Robinisca macrocnemis with Viedebanttia schmitzi. In the present paper V. macrocnemis is considered as a separate species, because it differs from V. schmitzi in the arrangement of setae la, ra and wa on tarsus I (la is located near seta wa in V. schmitzi and near seta ra in R. macrocnemis) and in the length of seta of trochanters I (longer than width of gnathosoma in V. schmitzi and shorter in V. macrocnemis). The genus *Viedebanttia* comprises 8 species distributed in Palaearctic, Nearctic and Neotropic regions. Species of *Viedebanttia* are associated with scarabaeid beetles (Coleoptera: Scarabaeidae), fleas (Siphonaptera), ants (Hymenoptera: Formicidae) and mammals (Insectivora and Rodentia). Only *V. schmitzi*, the type species of the genus, is known from both adults and hypopi, other species are described from hypopi only.

*Included species* (with data on distribution, hosts and habitats). V. schmitzi Oudemans, 1929 - Netherlands, Belgium, Germany, Czech and Slovak Republics, Russia (Moscow Prov.), nest of Talpa europea Linnaeus, 1758 (Insectivora: Talpidae), forest litter; V. fuscipes (Vitzthum, 1924), comb. n. = Tyroglyphus fuscipes Vitzthum, 1924 - Brasilia; V. macrocnemis (Zachvatkin, 1941), comb. n. = Robinisca macrocnemis Zachvatkin, 1941 – Russia (Ivanovo and Moscow Provinces, Primorsk Terr.), soil mixed with waste products of rye threshing; V. longipes (Volgin, 1951), comb. n. = Robinisca longipes Volgin, 1951 - Ukraine (Poltava), ex Lethrus (Lethrus) apterus Laxmann, 1770 (Coleoptera: Scarabaeidae); V. vitzthumi (Mahunka, 1979), comb. n. = Caloglyphus vitzthumi Mahunka, 1979 – Costa Rica (Puerto Viejo), ex Neivamyrmex rugulosus Borgmeier, 1953 (Hymenoptera: Formicidae); V. diamanus Fain & Schwan, 1984 – USA (California), ex Oropsylla (Diamanus) montana (Baker, 1895) (Siphonaptera: Ceratophyllidae) found in nest of Citellus (= Spermophilus) beecheyi (Richardson, 1829) (Rodentia: Sciuridae); V. coniferae (Sevastianov & Marroch, 1993), comb. n. = Robinisca coniferae Sevastianov & Marroch, 1993 – Lithuania, ex Geotrupes (Anoplotrupes) stercorosus (Scriba, 1791) = G. sylvaticus Panzer (Coleoptera: Scarabaeidae); V. egorovi sp. n. – South Korea (Suwon), ex Rhombonyx testaceipes Motshulsky, 1860 (after Zachvatkin, 1941; Volgin, 1951; Turk & Turk, 1957; Tareev, 1971; Černý & Samšińák, 1971; Mahunka, 1979; Fain & Schwan, 1984; Fain, 1985; Sevastianov & Marroch, 1993; Bugrov, 1997, and personal observations).

# Viedebanttia egorovi sp. n.

(Figs 1-8)

Holotype. Hypopus, South Korea, Suwon-city, Seocho River, Yogisan Mt., ex Rhombonyx testaceipes Motshulsky, 1860 (Coleoptera: Scarabaeidae) (No. 8.1), 3.VII.1997 (A.B. Egorov).

*Description.* Body brown, smooth, ovoid, anterior edge straight. Idiosoma 365.1 long, 255.2 wide.

Gnathosoma (Fig. 7) elongate, protruding from anterior idiosomal edge, 53.3 long, 8.5-11.9 wide at base and tip, respectively. Length of free palpomeres 11.3. Palpal solenidia (75.1) longer than gnathosoma. Anterior gnathosomal setae 17.0, posterior ones 14.5 long.

Dorsum (Fig. 2). Propodosoma strongly reduced and comparatively weakly sclerotized, almost completely hidden under hysterosoma. Only short (8.0) tubercle-like rostrum and narrow lateral edges of propodosoma not covered by hysterosomal shield. vi short (7.3), placed under transparent rostrum; ve situated laterally of vi at base of rostrum. se and si both covered by hysterosoma, well visible. scx 55.7 long, placed ventrally but most part of the setae visible dorsally. Hysterosoma 357.1 long. Anterior edge of hysterosoma straight, emarginated by wide (39.5) band bearing characteristic transverse lineal pattern. Transverse lines of the pattern gradually interrupted toward posterior edge of the band. All idiosomal setae developed, represented by microsetae (only vi and  $h_3$  longer than other setae).  $h_3$ 32.7 long. Setae  $c_1$  placed on anterior hysterosomal band. Distance between several idiosomal setae: si-si 42.9, se-se 121.1, d1-d1 79.9,  $c_1$ - $c_1$  136.4,  $e_1$ - $e_1$  95.7,  $h_1$ - $h_1$  89.6,  $h_2$ - $h_2$ 41.2, h<sub>3</sub>-h<sub>3</sub> 58.1. Opening of opisthogastric glands displaced ventrally, located near anterior hysterosomal angle just posterior to *cp*.

Venter (Fig. 1). Sternum 50.4 long, not reaching posterior edge of sternal shield (distance between end of sternum and the edge approximately equal to sternum length). Epimerae II (61.8) not reaching posterior edge of the shield, but forming weakly developed connections to lateral angle of sternal shield, therefore coxal field II enclosed. Posterior edge of shield straight, 99.3 long; lateral edges 96.9 long. Sternal and ventral shields touching each other. There is a pair of additional triangular shields (72.7 long, 21.8 wide) between lateral edges of sternal and anterior lateral edges of ventral shield. c<sub>3</sub> 9.7 long. Ventral shield 84.8 long, 109.0 wide, separated from genital one by well-visible unsclerotized suture. Punctate pattern of both shields as well as additional shields is better developed than on sternal shield. Coxal fields III and IV enclosed. Ventrum not interrupted, beginning from anterior edge of ventral shield and ending at its posterior edge. Genital shield 48.4 long, 89.6-50.7 wide at anterior edge and at level of narrowest part, respectively. Genital slit 40.7



long. *1a*, *3b* and *4a* represented by conoids; their diameter 7.0, 7.3 and 10.2, respectively. Bases of 3b placed on sclerotized spots. 3a and g represented by setae, 10.4 and 13.3long, respectively. Anal plate 79.9 long, 97.1 wide, bearing well-defined porous sculpture (Fig. 8). Fore suckers (19.4); central one  $(21.9 \times 17.2)$  with 2 pores not touching each other; hind suckers (12.1), anterior edge of lateral suckers (14.0) placed at level of anterior edges of central ones; fore cuticular suckers (17.0) developed, supplied with rounded central sclerites with granular sculpture; paired lateral cuticular suckers  $(28.8 \times 17.7)$  as well as unpaired posterior sucker darker and more sclerotized than the rest surface of anal plate, bearing fine granular pattern. Pores *ih* touching posterior cuticular suckers.

Legs. Length of leg I-II podomeres (femora-tarsi): 36.3, 33.4, 52.6, 79.9; 41.2, 27.4, 28.3, 72.7, respectively. Legs I (Figs 3-4): seta of trochanter short (8.2); vF 26.6; mG 40, cG 9.7, σ 58.1; gT 26.6, hT 29.1, φ 113.8; tarsus I –  $\omega_1$  19.4,  $\omega_2$  12.1, both solenidia placed at proximal edge of tarsus, ba ( $\omega_3$  after Griffiths, 1970) long (37.5), represented by solenidion, widened at base (3.6); wa 33.2; ra 38.8; la enlarged (60.6); f 21.6; e 24.2; d 17.0, d and e near each other; vsc 14.5. Legs II (Figs 5-6): seta of trochanter 27.6 long; vF 50.9; mG 46.7; gT 29.1, hT 26.2; tarsus II - ba (48.0) represented by seta, not widened at base; wa 35.8; ra 36.3; la not enlarged (38.8); f 24.2; e 17.0; d 4.2, d displaced proximal to e; vsc 17.0. Length of claws III-IV more than half of corresponding tarsi. Claw IV 24.2, tarsus IV 32.7 long. Setae d displaced medially on tarsi III-IV. Chaetotaxy and solenidiotaxy of legs I-II as follows: 1-1-2+(1)-2+(1)-8+(3); 1-1-2+(1)-2+(1)-9+(1), respectively.

*Etymology.* The new species is dedicated to Dr. A.B. Egorov (IBPV) who collected the coleopteran host with the holotype of the new species.

Comparison. The new species differs from other species of the genus in the strongly reduced propodosoma of which most part is placed under anterior part of hysterosomal shield; length of hysterosoma/length of free part propodosoma more than 44. The species is closely related to V. schmitzi and V. macrocnemis but differs in the following characters: ba reaches two-thirds of tarsus I (reaches tip of tarsus in V. schmitzi and V. macrocnemis); pores on central suckers of anal plate are separated (touching each other in V. schmitzi and V. macrocnemis).

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#### References

- Černý & Samšińák, 1971. Nadkohorta Acaridiae. In: M. Daniel & V. Černý (eds). Klíč zvířeny ČSSR, 4: 496-529. Prague.
- Bugrov, S.A. 1997. Free-living Astigmata (Acariformes) of the Moscow District fauna. Zool. Zhurn., 76: 147-156. (In Russian).
- Fain, A. 1985. Notes on two genera of mites (Viedebanttia and Nanacarus) (Acari: Astigmata) described by Oudemans. Zool. Meded. Leiden, 59: 275-280.
- Fain, A. & Schwan, T.G. 1984. Three new hypopial nymphs phoretic on fleas parasitic on rodents in California, U. S. A. Bull. Ann. Soc. r. belg. Entomol., 120: 91-97.
- Griffiths, D.A. 1970. A further systematic study of the genus Acarus L., 1758 (Acaridae, Acarina), with a key to species. Bull. British Mus. (nat. Hist.), Zool. Ser., 19: 85-118.
- Griffiths, D.A., Atyeo, W.T., Norton, R.A. & Lynch, C.A. 1990. The idiosomal chaetotaxy of astigmatid mites. J. Zool., 220: 1-32.
- Mahunka, S. 1979. The examination of myrmecophilous Acaroidea mites based on the investigations of Dr. C.W. Rettenmeyer (Acari: Acaroidea). II. Acta zool. hung., 25: 311-356.
- Samšińák, K. 1980. Caloglyphus rodriguezi sp. n., with taxonomic remarks on the tribe Caloglyphini (Acari, Acaridae). Mitt. zool. Mus. Berlin, 56: 201-206.
- Sevastianov, V.D. & Marroch, T.N. 1993. New species of entomophilous mites of the family Acaridae (Sarcoptiformes). Zool. Zhurn., 72: 143-150. (In Russian).
- Tareev, V.N. 1970. Akaroidnye kleshchi (Acaroidea) Primorskogo Kraya [Acaroid mites (Acaroidea) of the Primorsk Territory]: 1-32. Abstract of Ph. D. thesis. Vladivostok. (In Russian).
- Turk, E. & Turk, F. 1957. Systematik und Ökologie der Tyroglyphiden Mitteleuropas. In: H.-J. Stammer (ed.). Beiträge zur Systematik und Ökologie mitteleuropäischer Acarina, 1: 3-226.
- Volgin, V.I. 1951. New species of tyroglyphid mites (Acarina, Tyroglyphidae). Sbornik rabot Inst. prikl. Zool. Fitopatol., 1: 32-36. (In Russian).
- Zachvatkin, A.A. 1941. Tyroglyphoid mites (Tyroglyphoidea). Fauna SSSR. Paukoobraznye, 6: 1-475. Moscow & Leningrad. (In Russian).

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