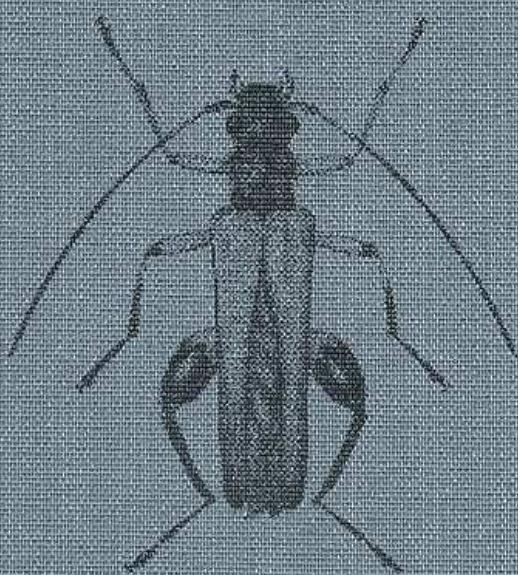


Daniel KUBISZ, Dariusz IWAN, Piotr TYKARSKI

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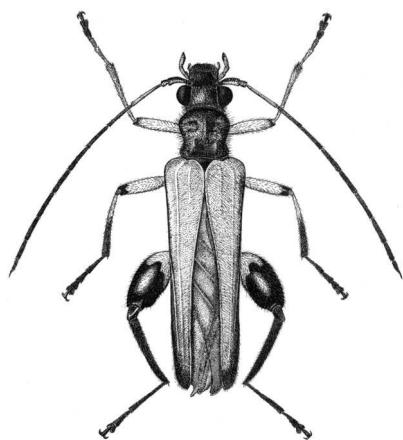
COLEOPTERA POLONIAE

2

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Warszawa 2014

COLEOPTERA POLONIAE



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PUBLISHERS

University of Warsaw – Faculty of Biology

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***Natura optima dux* Foundation, Warsaw**

Coleoptera Poloniae, Volume 2

Website: <http://www.biomap.pl>

Abstracted in: Current Contents*/Agriculture, Biology, and Environmental Sciences;
Entomological Abstracts and Zoological Record

The title partly sponsored by the University of Warsaw,
[Tytuł częściowo sponsorowany przez Uniwersytet Warszawski]

Cover illustration: *Oedemera podagrariae* (Linnaeus, 1767) by Przemysław Szwalko

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ISSN 2391-5285

ISBN 978-83-935107-9-5

Printed in Poland by PAN, Warszawska Drukarnia Naukowa, Warszawa

Coleoptera Poloniae
Tenebrionoidea

This publication should be cited as follows:

Kubisz D., Iwan D., Tykarski P. 2014. Tenebrionoidea (Tetratomidae, Melandryidae, Ripiphoridae, Prostomidae, Oedemeridae, Mycteridae, Pythidae, Aderidae, Scraptiidae). Critical checklist, distribution in Poland and meta-analysis. *Coleoptera Poloniae*, Vol. 2. University of Warsaw – Faculty of Biology, *Natura optima dux* Foundation, Warszawa, 468 pp.

Coleoptera Poloniae: Tenebrionoidea (Tetratomidae, Melandryidae, Ripiphoridae, Prostomidae, Oedemeridae, Mycteridae, Pythidae, Aderidae, Scraptiidae)

Critical checklist, distribution in Poland and meta-analysis

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Abstract

The book catalogues distribution data on occurrence of families of Tenebrionoidea. Having reviewed available distribution data, we consider 84 species to be present in Poland: Aderidae (7 species), Melandryidae (27), Oedemeridae (22), Prostomidae (1), Pythidae (3), Ripiphoridae (3), Scraptiidae (15) and Tetratomidae (6). 27 species were considered doubtful due to misidentifications or a long-term lack of new occurrences, including Mycteridae. Data on distribution of the confirmed taxa cover source references, localities, UTM 10×10 km grid coordinates, dates and collections that hold specimen, accompanied by distribution maps generalized to the UTM grid. A separate chapter gives an overview of Palaearctic distribution of all the discussed taxa, including subspecies when applicable. Detailed taxonomic checklist of the covered groups including synonymy is also provided. The distribution catalogue part is followed by the meta-analysis built upon a database covering all the presented information. A number of analytical and generalization techniques was used, giving synthetic views on research intensity and some other parameters at a species and family level. The publication follows the former volume of the Coleoptera Poloniae series, extending traditional faunistics by links to the database that is available online through the Biodiversity Map and Coleoptera Poloniae websites, served by the Polish Biodiversity Information Network (KSIB).

Key words

Coleoptera, Tenebrionoidea, Tetratomidae, Melandryidae, Ripiphoridae, Prostomidae, Oedemeridae, Mycteridae, Pythidae, Aderidae, Scraptiidae, biodiversity, faunistics, zoogeography, distribution, meta-analysis, checklist, museum collections, Poland.

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TAXONOMY AND SPECIES CHECKLIST

(–) species of not confirmed or doubtful presence in Poland

Family **TETRATOMIDAE** BILLBERG, 1820

Subfamily **TETRATOMINAE** BILLBERG, 1820

Genus ***Tetratoma*** FABRICIUS, 1790

Subgenus ***Abstrulia*** CASEY, 1900

Tetratoma ancora FABRICIUS, 1790

Tetratoma picta NEWMAN, 1835

Tetratoma conjuncta PIC, 1901

Tetratoma obscurior PIC, 1901

Tetratoma pfefferi ROUBAL, 1930

Tetratoma perrisi HOULBERT, 1934

Subgenus ***Tetratoma*** FABRICIUS, 1790

Tetratoma fungorum FABRICIUS, 1790

Dermestes collaris QUENSEL, 1790

Tetratoma dermestoides HERBST, 1792

Boletaria bicolor MARSHAM, 1802

Subfamily **EUSTROPHINAE** GISTEL, 1856

Tribe **Eustrophini** GISTEL, 1856

Genus ***Eustrophus*** ILLIGER, 1802

Eustrophus dermestoides (FABRICIUS, 1792)

Mycetophagus dermestoides FABRICIUS, 1792

Tritoma sicula BAUDI DI SELVE, 1892

Subfamily **HALLOMENINAE** MULSANT, 1856

Genus ***Hallomenus*** PANZER, 1793

Hallominus PAYKULL, 1799
Dinophorus ILLIGER, 1807
Dryala MULSANT, 1856

Hallomenus axillaris (ILLIGER, 1807)

Mordella picea MARSHAM, 1802
Dinophorus axillaris ILLIGER, 1807
Hallomenus fuscus GYLLENHAL, 1810
Dircea dorsalis BECK, 1817
Hallomenus dimidiatus STEPHENS, 1835
Hallomenus scapulatus FAIRMAIRE, 1881

Hallomenus binotatus (QUENSEL, 1790)

Chrysomela binotata QUENSEL, 1790
Elaterooides boleti KUGELANN, 1792
Ips elateroides KUGELANN, 1792
Hallomenus humeralis PANZER, 1793
Hallominus bipunctatus PAYKULL, 1799

Genus ***Mycetoma*** DEJEAN, 1834

Mycetoma suturale (PANZER, 1797)

Dryops suturalis PANZER, 1797
Mycetoma richteri ROUBAL, 1933

Family **MELANDRYIDAE** LEACH, 1815

Subfamily **MELANDRYINAE** LEACH, 1815

Tribe **Dircaeini** MULSANT, 1856

Genus ***Abdera*** STEPHENS, 1832

Carida MULSANT, 1856

Subgenus ***Abdera*** STEPHENS, 1832

(-) ***Abdera biflexuosa*** (CURTIS, 1829)

Elater flexuosus A. G. OLIVIER, 1790
Mordella bifasciata MARSHAM, 1802
Hypulus biflexuosus CURTIS, 1829
Abdera bifasciata STEPHENS, 1832
Dircea griseoguttata FAIRMAIRE, 1850
Abdera undata PERRIS, 1852

(-) ***Abdera quadrifasciata*** (CURTIS, 1829)

Hypulus quadrifasciata CURTIS, 1829

Subgenus ***Caridua*** E. STRAND, 1929

Abdera affinis (PAYKULL, 1799)

Hallominus affinis PAYKULL, 1799

Abdera picea WALKER, 1837

Abdera flexuosa (PAYKULL, 1799)

Hallominus flexuosus PAYKULL, 1799

Hallomenus undatus PANZER, 1799

Genus ***Anisoxya*** MULSANT, 1856

Anisoxya fuscula (ILLIGER, 1798)

Serropalpus fusculus ILLIGER, 1798

Anisoxya tenuis ROSENHAUER, 1847

Anisoxya mustela ABEILLE DE PERRIN, 1869

Genus ***Dircea*** FABRICIUS, 1798

Dircea australis FAIRMAIRE, 1856

Dircea quadriguttata DUFTSCHMID, 1812

Dircea quadrisignata MULSANT, 1856

Dircea imitata OLEXA, 1955

Dircea quadriguttata (PAYKULL, 1798)

Hypulus quadriguttatus PAYKULL, 1798

Dircea quadriguttata FABRICIUS, 1798

Dircea guttata STIERLIN, 1898

Dircea shibatai HAYASHI, 1960

Genus ***Phloiotrya*** STEPHENS, 1832

Subgenus ***Phloiotrya*** STEPHENS, 1832

Phloeotrya AGASSIZ, 1846

Phloiotrya rufipes (GYLLENHAL, 1810)

Dircea rufipes GYLLENHAL, 1810

Dircea testaceipennis PIC, 1900

(-) ***Phloiotrya subtilis*** (REITTER, 1897)

Dircea subtilis REITTER, 1897

Genus ***Wanachia*** SCHULZE, 1912

Caridina SEIDLITZ, 1898

Wanachia triguttata (GYLLENHAL, 1810)

Dircea triguttata GYLLENHAL, 1810

Abdera scutellaris MULSANT, 1856

Abdera suturalis BAUDI DI SELVE, 1877

Tribe **Hypulini** SEIDLITZ, 1875

Genus **Hypulus** PAYKULL, 1798

Mystaxus SEMENOV, 1900

Hypulus bifasciatus (FABRICIUS, 1792)

Notoxus bifasciatus FABRICIUS, 1792

Ptilinus alni D. H. SCHNEIDER, 1792

Helops fasciatus PANZER, 1793

Hypulus queruginosus (QUENSEL, 1790)

Elater queruginosus QUENSEL, 1790

Elater blekingensis D. H. SCHNEIDER, 1792

Notoxus dubius FABRICIUS, 1792

Hypulus undulatus PIC, 1925

Tribe **Melandryini** LEACH, 1815

Genus **Melandrya** FABRICIUS, 1801

Subgenus ***Melandrya*** FABRICIUS, 1801

Melandrya barbata (FABRICIUS, 1787)

Helops barbatus FABRICIUS, 1787

Melandrya flavigaster DUFTSCHMID, 1812

Melandrya caraboides (LINNAEUS, 1760)

Chrysomela caraboides LINNAEUS, 1760

Helops serratus FABRICIUS, 1775

Tenebrio rufibarbis SCHALLER, 1783

Melandrya tibialis VITURAT, 1895

Melandrya luridipes PIC, 1907

Melandrya coeruleascens PETRI, 1912

Subgenus ***Paramelandrya*** NIKITSKY, 2002

Melandrya dubia (SCHALLER, 1783)

Tenebrio dubius SCHALLER, 1783

Helops canaliculatus FABRICIUS, 1787

Melandrya caraboides LATREILLE, 1818

Melandrya rufipes CHEVROLAT, 1833

Melandrya goryi LAPORTE, 1840

Melandrya alternans MOTSCHULSKY, 1872

Melandrya niponica LEWIS, 1895

Genus ***Phryganophilus*** C. R. SAHLBERG, 1833

Longemelandrya PIC, 1953

Phryganophiloides MORISHIMA, 1988

Phryganophilus auritus MOTSCHULSKY, 1845

Phryganophilus nigriventris HAMPE, 1850
Phryganophiloides elegans MORISHIMA, 1988

Phryganophilus ruficollis (FABRICIUS, 1798)

Cantharis analis FABRICIUS, 1792
Dircea ruficollis FABRICIUS, 1798
Lymexylon paradoxus PAYKULL, 1799
Melandrya ruficollis LEWIS, 1895
Phryganophilus rosti HUBENTHAL, 1905

Tribe **Orchesiini** MULSANT, 1856

Genus ***Orchesia*** LATREILLE, 1807

Subgenus ***Clinocara*** C. G. THOMSON, 1859

Orchesia blandula BRANCSIK, 1874

Orchesia carpathica REITTER, 1878
Orchesia transsylvanica REITTER, 1878

Orchesia fasciata (ILLIGER, 1798)

Serropalpus fasciatus ILLIGER, 1798
Dircea trifasciata ZETTERSTEDT, 1838
Orchesia laterufa PIC, 1927

(-) ***Orchesia grandicollis*** ROSENHAUER, 1847

Orchesia laticollis L. REDTENBACHER, 1849

Orchesia minor WALKER, 1837

Orchesia sepicola ROSENHAUER, 1847
Orchesia tetratoma C. G. THOMSON, 1864

Orchesia undulata KRAATZ, 1853

Hallomenus fasciatus GYLLENHAL, 1810

Subgenus ***Orchesia*** LATREILLE, 1807

Orchesia fusiformis SOLSKY, 1871

Orchesia acicularis REITTER, 1886
Orchesia nadeshdae SEMENOV, 1898
Orchesia obscuricolor PIC, 1954

Orchesia micans (PANZER, 1793)

Megatoma picea HERBST, 1792
Hallomenus micans PANZER, 1793
Orchesia clavicornis LATREILLE, 1804
Orchesia australis GUILLEBEAU, 1887
Orchesia reyi GUILLEBEAU, 1887
Orchesia subimpressa GUILLEBEAU, 1887
Orchesia abeillei GUILLEBEAU, 1888
Orchesia corsica GUILLEBEAU, 1888

Subgenus ***Orchesteria*** GUILLEBEAU, 1887

Orchesia luteipalpis MULSANT et GUILLEBEAU, 1857

Tribe **Serropalpini** LATREILLE, 1829

Genus ***Serropalpus*** HELLENIUS, 1786

Serropalpus barbatus (SCHALLER, 1783)

Mordella barbata SCHALLER, 1783

Serropalpus striatus HELLENIUS, 1786

Lymexylon biguttatus SCHELLENBERG, 1802

Serropalpus niponicus LEWIS, 1895

Tribe **Xylitini** C. G. THOMSON, 1864

Genus ***Dolotarsus*** JACQUELIN DU VAL, 1863

Dolotarsus lividus (C. R. SAHLBERG, 1833)

Dircea livida C. R. SAHLBERG, 1833

Dircea ephippia SCHAUM, 1859

Dolotarsus rufipes JACQUELIN DU VAL, 1863

Phryganophilus suturus GREDLER, 1866

Genus ***Xylita*** PAYKULL, 1798

Xylita laevigata (HELLENIUS, 1786)

Serropalpus laevigatus HELLENIUS, 1786

Elater buprestoides FABRICIUS, 1792

Dircea discolor FABRICIUS, 1798

Dircea decolorata RANDALL, 1858

Xylita robusta MOTSCHULSKY, 1872

Xylita producta HATCH, 1965

Tribe **Zilorini** NIKITSKY, 2007

Genus ***Zilora*** MULSANT, 1856

(-) ***Zilora elongata*** J. R. SAHLBERG, 1881

(-) ***Zilora ferruginea*** (PAYKULL, 1798)

Xylita ferruginea PAYKULL, 1798

Zilora obscura (FABRICIUS, 1794)

- Parnus obscurus* FABRICIUS, 1794
Helops sericeus J. STURM, 1807
Mycetochara bifoveolata DUFOUR, 1851
Zilora ferruginea MULSANT, 1856
Zilora eugeniae GANGLBAUER, 1891

Subfamily **OSPHYINAE** MULSANT, 1856

Genus ***Conopalpus*** GYLLENHAL, 1810

Conopalpus testaceus (A. G. OLIVIER, 1790)

- Melyris testaceus* A. G. OLIVIER, 1790
Conopalpus flavicollis GYLLENHAL, 1810
Conopalpus nigricornis GERMAR, 1821
Conopalpus vigorsi STEPHENS, 1832
Conopalpus ruficollis NEWMAN, 1833
Conopalpus thoracicus BAUDI DI SELVE, 1877
Conopalpus obscuriceps PIC, 1925

Genus ***Osphya*** ILLIGER, 1807

- Nothous* A. G. OLIVIER, 1811
Pelecina ILLIGER, 1807

Osphya bipunctata (FABRICIUS, 1775)

- Cantaris bipunctata* FABRICIUS, 1775
Nothus clavipes A. G. OLIVIER, 1811
Nothus praeustus A. G. OLIVIER, 1811
Dryops clavipes GYLLENHAL, 1817
Nothus bimaculata STEPHENS, 1832
Nothus anceps STEPHENS, 1839
Nothus uralensis MOTSCHULSKY, 1845
Osphya obscuripennis PIC, 1897
Osphya innotata PIC, 1898
Osphya vittipennis SEIDLITZ, 1898
Osphya impunctata BOUSKELL, 1899
Osphya maculata BOUSKELL, 1899
Osphya quadripunctata BOUSKELL, 1899
Osphya apicentorata PIC, 1908
Osphya testaceithorax PIC, 1909

Family **RIPIPHORIDAE** GEMMINGER et HAROLD, 1870

Subfamily **PELECOTOMINAE** SEIDLITZ, 1875

Genus **Pelecotoma** FISCHER VON WALDHEIM, 1809

Pelecotoma fennica PAYKULL, 1799

Ripiphorus finnicus PAYKULL, 1799

Pelecotoma latreillei FISCHER VON WALDHEIM, 1823

Pelecotoma mosquense FISCHER VON WALDHEIM, 1809

Pelecotoma zoufali FLEISCHER, 1916

Subfamily **RIPIDIINAE** GERSTAECKER, 1855

Genus **Ripidius** THUNBERG, 1806

Symbius SUNDEVALL, 1831

Rhipidius AGASSIZ, 1846

Pararhipidius COIFFAIT 1947

Ripidius quadriceps (ABEILLE DE PERRIN, 1872)

Rhipidius quadriceps ABEILLE DE PERRIN, 1872

Rhipidius apicipennis KRAATZ, 1891

Rhipidius kabylianus CHOBAUT, 1902

Rhipidius parisiensis LESNE, 1902

Rhipidius boissyi ABEILLE DE PERRIN, 1909

Rhipidius amori BOLÍVAR Y PIETAIN, 1917

Rhipidius denisi CHOBAUT, 1919

Rhipidius maroccanus CHOBAUT, 1922

Rhipidius vilarrubiae ESPAÑOL, 1942

Pararhipidius burdigalensis COIFFAIT, 1947

Rhipidius tigrani IABLOKOV-KHNZORIAN, 1957

Subfamily **RIPIPHORINAE** GEMMINGER et HAROLD, 1870

Tribe **Macrosiagonini** REITTER, 1911

Genus **Macrosiagon** HENTZ, 1830

Emenadia LAPORTE, 1840

Siagonadia REITTER, 1910

(-) **Macrosiagon bimaculata** (FABRICIUS, 1787)

Mordella tricuspidata GMELIN, 1790-

Rhipiphorus carinthiacus PANZER, 1794

Rhipiphorus bicolor A. G. OLIVIER, 1795

Mordella larvata SCHRANK VON PAULA, 1789

Rhipiphorus litoratus FABRICIUS, 1798

Rhipiphorus quadrimaculatus GYLLENHAL, 1817

Rhipiphorus binotatus FISCHER VON WALDHEIM, 1829
Rhipiphorus apicalis STEVEN, 1832
Rhipiphorus binotatus FISCHER VON WALDHEIM, 1832
Rhipiphorus nigrithorax STEVEN, 1832
Rhipiphorus angustulus MOTSCHULSKY, 1849
Macrosiagon pallidipes REITTER, 1898

(-) ***Macrosiagon praeusta* (GEBLER, 1829)**

Rhipiphorus praeustus GEBLER, 1829
Rhipiphorus sulcatus MÉNÉTRIÉS, 1832
Rhipiphorus sulcatus FISCHER VON WALDHEIM, 1832
Rhipiphorus rufipennis CHEVROLAT, 1834
Rhipiphorus fulvipennis KÜSTER, 1846
Rhipiphorus apicalis KÜSTER, 1848

Genus ***Metoecus*** DEJEAN, 1834

Ripidastes GISTEL, 1848
Cyttaroecus SCHILDER, 1923

***Metoecus paradoxus* (LINNAEUS, 1760)**

Mordella paradoxa LINNAEUS, 1760
Mordella erythrogaster FRÖLICH, 1792
Ripiphorus angulatus PANZER, 1803
Rhipiphorus affinis STEVEN, 1832
Metoecus pictus GISTEL, 1857
Metoecus tricolor GISTEL, 1857
Metoecus abdominalis GRADL, 1882
Metoecus apicalis GRADL, 1882
Metoecus flavoniger GRADL, 1882
Metoecus macularis GRADL, 1882
Metoecus nigrescens GRADL, 1882
Metoecus nigriventris GRADL, 1882
Metoecus notiventris GRADL, 1882
Metoecus semiflavus GRADL, 1882
Myiodes antoniae REITTER, 1895
Metoecus gradli SCHILSKY, 1900
Metoecus atripennis PIC, 1916
Metoecus luteonotatus PIC, 1916
Metoecus multinotatus PIC, 1916
Metoecus nigrolineatus PIC, 1916
Metoecus theresae PIC, 1916
Metoecus trinotatus PIC, 1916
Metoecus viturati PIC, 1916
Metoecus basalis SCHILDER, 1924
Metoecus dicipiens SCHILDER, 1924
Metoecus humeralis SCHILDER, 1924
Metoecus obscuripennis SCHILDER, 1924

Family **PROSTOMIDAE** C. G. THOMSON, 1859

Genus ***Prostomis*** LATREILLE, 1819

Megagnathus DEJEAN, 1821

Prostomis mandibularis (FABRICIUS, 1801)

Trogosita madibularis FABRICIUS, 1801

Prostomis brunneus FURSOV, 1935

Prostomis elburica FLEISCHER, 1919

Cucujus maxillosus P. W. J. MÜLLER, 1805

Prostomis strandi ROUBAL, 1936

Family **OEDEMERIDAE** LATREILLE, 1810

Subfamily **CALOPODINAE** A. COSTA, 1852

Genus ***Calopus*** FABRICIUS, 1775

Calopus serraticornis (LINNAEUS, 1758)

Cerambyx serraticornis LINNAEUS, 1758

Calopus pretneri J. MÜLLER, 1929

Subfamily **OEDEMERINAE** LATREILLE, 1810

Tribe **Ditylini** MULSANT, 1858

Genus ***Chrysanthia*** W. L. E. SCHMIDT, 1844

Chrysanthia geniculata geniculata W. L. E. SCHMIDT, 1846

Chrysanthia geniculata W. L. E. SCHMIDT, 1846

Chrysanthia viridis W. L. E. SCHMIDT, 1846

Chrysanthia geniculata L. HEYDEN, 1877

Chrysanthia cupreomicans WESTHOFF, 1881

Chrysanthia nigricornis WESTHOFF, 1881

Chrysanthia saturata WESTHOFF, 1881

Chrysanthia viridissima (LINNAEUS, 1758)

Cantharis viridissima LINNAEUS, 1758

Cantharis viridis DE GEER, 1775

Necydalis thalassina FABRICIUS, 1792

Chrysanthia cuprina REY, 1892

Chrysanthia korbi REITTER, 1894

Chrysanthia diversipes PIC, 1932

Genus ***Ditylus*** FISCHER VON WALDHEIM, 1817

Mimetus ESCHSCHOLTZ, 1818

(-) ***Ditylus laevis laevis*** (FABRICIUS, 1787)

Helops laevis FABRICIUS, 1787

Ditylus helopoides FISCHER VON WALDHEIM, 1817

Mimetus unicolor ESCHSCHOLTZ, 1818

Tribe **Nacerdini** MULSANT, 1858

Genus ***Anogcodes*** DEJEAN, 1834

Anoncodes W. L. E. SCHMIDT, 1844

Pachychirus L. REDTENBACHER, 1845

Lethonymus MARSEUL, 1857

Oedechira MOTSCHULSKY, 1872

Peronocnemis FAIRMAIRE, 1886

Anoncodina SEIDLITZ, 1899

Anogcodes fulvicollis (SCOPOLI, 1763)

Cantharis fulvicollis SCOPOLI, 1763

Necydalis obscura GYLLENHAL, 1810

Anoncodes medius GREDLER, 1866

Nacerda longicollis ROUBAL, 1936

Nacerda therondi MÉGUIGNON, 1948

Anogcodes melanurus (FABRICIUS, 1787)

Necydalis melanurus FABRICIUS, 1787

Necydalis ustulatus FABRICIUS, 1787

Necydalis fasciatus VILLIERS, 1789

Ditylus rufus FISCHER VON WALDHEIM, 1820

Anoncodes scutellaris WALTL, 1839

(-) ***Anogcodes ruficollis*** (FABRICIUS, 1781)

Necydalis ruficollis FABRICIUS, 1781

Necydalis coerulescens ROSSI, 1790

Anogcodes rufiventris (SCOPOLI, 1763)

Cantharis rufiventris SCOPOLI, 1763

Leptura bipartita SCHRANK VON PAULA, 1776

Necydalis melanocephalus FABRICIUS, 1794

Oedemera dorsalis A. G. OLIVIER, 1811

(-) ***Anogcodes seladonius seladonius*** (FABRICIUS, 1792)

Necydalis seladonia FABRICIUS, 1792

Oedemera dispar DUFOUR, 1841

Anoncodes amoenus W. L. E. SCHMIDT, 1846

Anoncodes viridipes W. L. E. SCHMIDT, 1846

Anoncodes meridionalis A. COSTA, 1852

Nacerdes aurosus FAIRMAIRE, 1863

Anogcodes ustulatus (SCOPOLI, 1763)

- Cantharis ustulata* SCOPOLI, 1763
Leptura ferruginea SCHRANK VON PAULA, 1776
Necydalis hybrida ROSSI, 1794
Necydalis adustus PANZER, 1795
Necydalis collaris PANZER, 1795
Ditylus melanocephalus FISCHER VON WALDHEIM, 1829
Oedemera axillaris MÉNÉTRIÉS, 1832
Anogcodes flaviventris FALDERMANN, 1837
Oedemera paradoxa FALDERMANN, 1837
Oedechira flavipennis MOTSCHULSKY, 1872
Anoncodes zimmermanni GISTEL, 1857
Anoncodes inlateralis PIC, 1930

Genus **Nacerdes** DEJEAN, 1834

- Nacerda* STEPHENS, 1839
Nacerdoscuta PIC, 1915

Subgenus **Nacerdes** DEJEAN, 1834

- Nacerdes melanura** (LINNAEUS, 1758)
- Cantharis melanura* LINNAEUS, 1758
Cantharis nigripes FABRICIUS, 1781
Cantharis lepturoides THUNBERG, 1784
Cantharis testacea GEOFFROY, 1785
Necydalis notata FABRICIUS, 1792
Cantharis acuta MARSHAM, 1802
Oedemera analis A. G. OLIVIER, 1811
Necydalis erminea GERMAR, 1817
Oedemera apicalis SAY, 1835
Ditylus rufus BRULLÉ, 1839
Nacerdes sardea W. L. E. SCHMIDT, 1846
Xanthochroa italicica CHEVROLAT, 1877
Nacerdes zoufali REITTER, 1907

Subgenus **Xanthochroa** W. L. E. SCHMIDT, 1844

- Patiala* LEWIS, 1895
Patialomorpha NAKANE, 1954
Axanthochroa ŠVIHLA, 1986
Asiochroa ŠVIHLA, 1998

Nacerdes carniolica carniolica (GISTEL, 1834)

- Oedemera carniolica* GISTEL, 1834
Oedemera blossevillei GUÉRIN-MÉNEVILLE, 1838

Tribe **Asclerini** SEMENOV, 1894

Genus ***Ischnomera*** STEPHENS, 1832

Asclera DEJEAN, 1834

Ascleronia SEIDLITZ, 1899

Chromasclera SEIDLITZ, 1899

Ischnomera caerulea (LINNAEUS, 1758)

Cantharis caerulea LINNAEUS, 1758

Necydalis coerulescens FABRICIUS, 1775

Oedemera quadrinervosa LATREILLE, 1804

Oedemera nigripes A. G. OLIVIER, 1811

Ischnomera unicolor MELSHEIMER, 1846

Ischnomera opacocerulea REITTER, 1911

Asclera oblitterata PIC, 1926

Asclera carinithorax ROUBAL, 1936

Ischnomera cinerascens (PANDELLE, 1867)

Asclera cinerascens PANELLÉ, 1867

Ischnomera cyanea (FABRICIUS, 1792)

Necydalis cyanea FABRICIUS, 1792

Asclera viridescens PIC, 1920

Asclera graeca DAHLGREN, 1976

Ischnomera sanguinicollis (FABRICIUS, 1787)

Necydalis sanguinicollis FABRICIUS, 1787

Necydalis flavicollis PANZER, 1794

Genus ***Probosca*** W. L. E. SCHMIDT, 1846

Subgenus ***Probosca*** W. L. E. SCHMIDT, 1846

(–) ***Probosca unicolor*** (KÜSTER, 1847)

Chitona unicolor KÜSTER, 1847

Probosca plumbea SUFFRIAN, 1848

(–) ***Probosca virens*** (FABRICIUS, 1792)

Callidium virens FABRICIUS, 1792

Probosca viridana W. L. E. SCHMIDT, 1846

Probosca nigrofemorata PIC, 1898

Probosca purpureomicans PIC, 1913

Probosca hispanica PIC, 1920

Probosca boiteli PIC, 1936

Tribe **Oedemerini** LATREILLE, 1810

Genus **Oedemera** A. G. OLIVIER, 1789

Subgenus **Oedemera** A. G. OLIVIER, 1789

Stenolytra DILWYN, 1829

Oedemerina A. COSTA, 1852

Oedemerastra SEIDLITZ, 1899

Oedemerella SEIDLITZ, 1899

Oedemeronia SEIDLITZ, 1899

Oedemera croceicollis (GYLLENHAL, 1827)

Necydalis croceicollis GYLLENHAL, 1827

Oedemera sarmatica F. F. MORAWITZ, 1861

Oedemera femorata (SCOPOLI, 1763)

Cantharis femorata SCOPOLI, 1763

Necydalis flavescens LINNAEUS, 1767

Necydalis praterrana SCHRANK VON PAULA, 1781

Oedemera similis W. L. E. SCHMIDT, 1846

Oedemera fallax SEIDLITZ, 1899

Oedemera flavipes (FABRICIUS, 1792)

Necydalis flavipes FABRICIUS, 1792

Oedemera aenea A. G. OLIVIER, 1795

Necydalis clavipes FABRICIUS, 1801

Oedemera pusilla A. COSTA, 1852

Oedemera delagrangei PIC, 1923

Oedemera lurida lurida (MARSHAM, 1802)

Necydalis lurida MARSHAM, 1802

Oedemera viridula SEIDLITZ, 1899

Oedemera monticola ŠVIHLA, 1978

(-) **Oedemera nobilis** (SCOPOLI, 1763)

Cantharis nobilis SCOPOLI, 1763

Necydalis caerulea LINNAEUS, 1767

Necydalis ceramboides FORSTER, 1771

Oedemera magica DALLA TORRE, 1880

Oedemera subviolacea PIC, 1901

Oedemera gallica PIC, 1920

Oedemera viridescens PIC, 1920

Oedemera sirgueyi PIC, 1927

Oedemera podagrariae podagrariae (LINNAEUS, 1767)

Necydalis podagrariae LINNAEUS, 1767

Cantharis fulva GEOFFROY, 1785

Necydalis testacea FABRICIUS, 1792

Necydalis melanocephala PANZER, 1795

Necydalis simplex DONOVAN, 1801

Oedemera incerta MULSANT, 1858

Oedemera sericans MULSANT, 1858
Oedemera femoralis SEIDLITZ, 1899
Oedemera sebastiani PIC, 1901
Oedemera flavicrus SEIDLITZ, 1899
Oedemera discoidalis PIC, 1921
Oedemera distinctipes PIC, 1921
Oedemera diversipes PIC, 1921
Oedemera inlateralis PIC, 1921
Oedemera limbaticollis PIC, 1921
Oedemera reducta PIC, 1921

***Oedemera pthysica* (SCOPOLI, 1763)**

Cantharis pthysica SCOPOLI, 1763
Necydalis femorata PANZER, 1795
Oedemera subulata A. G. OLIVIER, 1795
Necydalis marginata FABRICIUS, 1798
Oedemera discoidalis LATREILLE, 1804
Oedemera stenoptera FALDERMANN, 1837
Oedemera angusticollis A. COSTA, 1852
Oedemera vittata J. FRIVALDSZKY, 1877

(–) ***Oedemera rufofemorata rufofemorata* GERMAR, 1824**

Oedemera rufofemorata GERMAR, 1824
Oedemera femorata BRULLÉ, 1832
Oedemera brevicornis W. L. E. SCHMIDT, 1846
Oedemera cyanescens W. L. E. SCHMIDT, 1846

***Oedemera subrobusta* (NAKANE, 1954)**

Oedemerina subrobusta NAKANE, 1954

(–) ***Oedemera tristis* W. L. E. SCHMIDT, 1846**

***Oedemera virescens virescens* (LINNAEUS, 1767)**

Cantharis virescens LINNAEUS, 1767
Necydalis striata HERBST, 1784
Oedemera abdominalis A. G. OLIVIER, 1795
Oedemera cupreomicans REITTER, 1905

Subgenus ***Oncomera*** STEPHENS, 1829

Oncomerella REITTER, 1911
Oncomerina SEIDLITZ, 1899

(–) ***Oedemera femoralis femoralis* (A. G. OLIVIER, 1803)**

Dryops femorata FABRICIUS, 1792
Oncomera femoralis A. G. OLIVIER, 1803
Necydalis calopoides GERMAR, 1817
Oncomera acutiventris ABEILLE DE PERRIN, 1896

Subgenus ***Stenaxis*** W. L. E. SCHMIDT, 1844

(–) ***Oedemera annulata* GERMAR, 1824**

Family **MYCTERIDAE** BLANCHARD, 1845

Genus ***Mycterus*** CLAIRVILLE, 1798

Rhinomacer FABRICIUS, 1781

(-) ***Mycterus curculioides*** (FABRICIUS, 1781)

Rhinomacer curculioides FABRICIUS, 1781

Curculio coriaceus THUNBERG, 1791

Mycterus griseus CLAIRVILLE, 1798

Mycterus gracilior FAIRMAIRE, 1874

Family **PYTHIDAE** SOLIER, 1834

Genus ***Pytho*** LATREILLE, 1796

Pytholus RAFINESQUE, 1815

Enoptes GISTEL, 1848

Pytho abieticola J. R. SAHLBERG, 1875

Pytho niger REITTER, 1911

Pytho depressus (LINNAEUS, 1767)

Tenebrio depressus LINNAEUS, 1767

Tenebrio lignarius DE GEER, 1775

Cucujus coeruleus HERBST, 1782

Cucujus coeruleus FABRICIUS, 1787

Cucujus castaneus FABRICIUS, 1792

Cucujus coeruleus FABRICIUS, 1792

Cucujus festivus FABRICIUS, 1792

Cucujus planus HERBST, 1799

Pytho atriceps PIC, 1912

Pytho basipennis PIC, 1912

Pytho kolwensis C. R. SAHLBERG, 1833

Pytho sahlbergi PIC, 1912

Family **ADERIDAE** WINKLER, 1927

Genus ***Aderus*** STEPHENS, 1829

Hylophilus BERTHOLD, 1827

Xylophilus LATREILLE, 1829

Phomalus CASEY, 1895

Aderus populneus (CREUTZER, 1796)

Notoxus populneus CREUTZER, 1796

Lytta boleti MARSHAM, 1802

Xylophilus biskrensis PIC, 1893

Genus **Anidorus** MULSANT et REY, 1866

Anidorus MULSANT et REY, 1866

(-) ***Anidorus lateralis*** (GREDLER, 1866)

Xylophilus lateralis GREDLER, 1866

Xylophilus tirolensis GREDLER, 1866

Anidorus nigrinus (GERMAR, 1842)

Xylophilus nigrinus GERMAR, 1842

Xylophilus patricius ABEILLE DE PERRIN, 1872

Euglenes nadeshdae SEMENOV, 1899

Xylophilus alpestris PIC, 1899

Genus **Cobososia** COLLADO et ALONZO-ZARAZAGA, 1996

(-) ***Cobososia pruinosa pruinosa*** (KIESENWETTER, 1861)

Xylophilus pruinosis KIESENWETTER, 1861

Xylophilus testaceus BAUDI DI SELVE, 1877

Xylophilus obscurus PIC, 1892

Hylophilus montandoni PIC, 1925

Hylophilus moreanus PIC, 1925

Genus **Euglenes** WESTWOOD, 1830

Xylophilus CURTIS, 1830

Euglenes oculatus (PAYKULL, 1798)

Anthicus oculatus PAYKULL, 1798

Lytta nigricollis MARSHAM, 1802

Euglenes pygmaeus (DE GEER, 1775)

Cerambyx pygmaeus DE GEER, 1775

Notoxus fulvus A. G. OLIVIER, 1795

Notoxus melanocephalus CREUTZER, 1796

Anthicus ferrugineus PAYKULL, 1798

Euglenes fennicus MANNERHEIM, 1843

Anidorus lokvenci ROUBAL, 1938

Genus **Phytobaenus** R. F. SAHLBERG, 1834

Phytobaenus amabilis amabilis R. F. SAHLBERG, 1834

Phytobaenus amabilis R. F. SAHLBERG, 1834

Xylophilus bisbimaculatus HAMPE, 1850

Pseudeuglenes PIC, 1897

Genus **Pseudanidorus** PIC, 1893

Pseudanidorus pentatomus (THOMSON, 1864)

Xylophilus pentatomus THOMSON, 1864

Euglenes quinquetomus SEIDLITZ, 1891

Euglenes rossicus SEMENOV, 1899

Tanilotes CASEY, 1895

Genus ***Vanonus*** CASEY, 1895

Vanonus brevicornis brevicornis (PERRIS, 1869)

Xylophilus brevicornis (PERRIS, 1869)

Hylophilus omissus PIC, 1948

Family **SCRAPTIIDAE** MULSANT, 1856

Subfamily **SCRAPTIINAE** MULSANT, 1856

Tribe **Scaptiini** MULSANT, 1856

Genus ***Scaptia*** LATREILLE, 1807

Scaptia fuscula P. W. J. MÜLLER, 1821

Scaptia nigricans STEPHENS, 1832

Scaptia minuta MULSANT, 1856

Scaptia nigriceps HEYDEN, 1870

Scaptia algirica PIC, 1898

Hallomenus innata KANGAS, 1959

Subfamily **ANASPIDINAE** MULSANT, 1856

Tribe **Anaspidini** MULSANT, 1856

Genus ***Anaspis*** GEOFFROY, 1762

Subgenus ***Anaspis*** GEOFFROY, 1762

Plesianaspis A. COSTA, 1854

Anaspis arctica ZETTERSTEDT, 1828

Anaspis ruficeps ZETTERSTEDT, 1840

Anaspis nigriceps SCHILSKY, 1895

Anaspis imitata SCHILSKY, 1899

Anaspis tenerbaumi PIC, 1928

Anaspis bohemica SCHILSKY, 1899

Anaspis forticornis SCHILSKY, 1899

Anaspis fortipes REITTER, 1911

Anaspis norvegica MÜNSTER, 1924

(-) ***Anaspis fasciata*** (FORSTER, 1771)

Mordella fasciata FORSTER, 1771

Mordella humeralis FABRICIUS, 1775

Anaspis bicolor GEOFFROY, 1785

Mordella biguttata ROSSI, 1794

Anaspis geoffroi P. W. J. MÜLLER, 1821

Anaspis quadrinotata STEPHENS, 1832

Anaspis quadripustulata STEPHENS, 1832

Anaspis scapularis STEPHENS, 1832
Anaspis subfasciata STEPHENS, 1832
Anaspis cruciata A. COSTA, 1854
Anaspis discicollis A. COSTA, 1854
Anaspis vulcanica A. COSTA, 1854
Anaspis quadrimaculata A. COSTA, 1854
Anaspis bipunctata RAGUSA, 1898
Anaspis bisbimaculata PIC, 1910
Anaspis sicula PIC, 1910
Anaspis biinterrupta PIC, 1931

***Anaspis frontalis* (LINNAEUS, 1758)**

Mordella frontalis LINNAEUS, 1758
Mordella lateralis GYLLENHAL, 1810
Mordella flavifrons ESCHSCHOLTZ, 1818
Mordella punctata ESCHSCHOLTZ, 1818
Anaspis verticalis FALDERMANN, 1837
Anaspis assimilis SNELLEN VAN VOLLENHOVEN, 1854
Anaspis flavoatra LETZNER, 1857
Anaspis maculicollis MOTSCHULSKY, 1860
Anaspis abollata GOZIS, 1882
Anaspis hopffgarteni SCHILSKY, 1895

***Anaspis kiesenwetteri* EMERY, 1876**

Anaspis picea SCHILSKY, 1895
Anaspis melanaria SCHILSKY, 1899

(-) ***Anaspis lurida* STEPHENS, 1832**

Mordella fusca SCHRANK VON PAULA, 1781
Mordella testacea MARSHAM, 1802
Anaspis subtestacea STEPHENS, 1832
Plesianaspis analis A. COSTA, 1854
Plesianaspis flava A. COSTA, 1854
Plesianaspis unicolor A. COSTA, 1854

(-) ***Anaspis maculata* (GEOFFROY, 1785)**

Mordella melanopa FORSTER, 1771
Mordella bimaculata ROSSI, 1792
Mordella nigricollis MARSHAM, 1802
Mordella obscura MARSHAM, 1802
Mordella pallida MARSHAM, 1802
Mordella bipunctata BONELLI, 1812
Anaspis exoleta A. COSTA, 1854
Anaspis testacea SNELLEN VAN VOLLENHOVEN, 1854
Silaria picta HAMPE, 1871
Anaspis innotata SCHILSKY, 1892
Anaspis florenceae DONISTHORPE, 1928

(-) ***Anaspis pulicaria* A. COSTA, 1854**

Anaspis lateralis STEPHENS, 1832
Anaspis forcipata MULSANT, 1856
Anaspis curtii ROUBAL, 1912

Anaspis ruficollis (FABRICIUS, 1792)

Mordella thoracica HERBST, 1784
Mordella ruficollis FABRICIUS, 1792
Anaspis alpicola EMERY, 1876
Anaspis bickhardti SCHILSKY, 1906

(–) ***Anaspis silvatica*** GABRIEL, 1916

Anaspis thoracica (LINNAEUS, 1758)

Mordella thoracica LINNAEUS, 1758
Anaspis lateralis C. G. THOMSON, 1864
Anaspis confusa EMERY, 1876
Anaspis septentrionalis CHAMPION, 1891
Anaspis gerhardii SCHILSKY, 1892
Anaspis latipalpis SCHILSKY, 1895
Anaspis scutellata SCHILSKY, 1899
Anaspis martialis PIC, 1918
Anaspis notatithorax PIC, 1918
Anaspis marginicollis H. LINDBERG, 1925
Anaspis obscuricolor STCHEGOLEVA-BAROVSKAYA, 1931
Anaspis occipitalis STCHEGOLEVA-BAROVSKAYA, 1931
Anaspis schilskyana HELLÉN, 1935

Subgenus ***Larisia*** EMERY, 1876

Anaspis palpalis (GERHARDT, 1876)

Silaria palpalis GERHARDT, 1876
Anaspis impressa GUILLEBEAU, 1893
Anaspis affinis SCHILSKY, 1895
Anaspis eucineticolor ROUBAL, 1935

Subgenus ***Nassipa*** EMERY, 1876

Anaspis costai EMERY, 1876

Anaspis fuscescens STEPHENS, 1832
Anaspis fulvipes COSTA, 1854
Plesianaspis thoracica A. COSTA, 1854
Anaspis skalitzkyi SCHILSKY, 1895

Anaspis flava (LINNAEUS, 1758)

Mordella flava LINNAEUS, 1758
Anaspis pectoralis SCHILSKY, 1899
Anaspis scutellaris SCHILSKY, 1899
Anaspis unicolor SCHILSKY, 1899
Anaspis fauconneti PIC, 1918

Anaspis melanostoma A. COSTA, 1854

Anaspis monilicornis MULSANT, 1856

***Anaspis rufilabris* (GYLLENHAL, 1827)**

Mordella clavicornis FORSTER, 1771
Mordella atra FABRICIUS, 1792
Mordella nigra ROSSI, 1792
Mordella rufilabris GYLLENHAL, 1827
Anaspis obscuripes SCHILSKY, 1895
Anaspis hudsoni DONISTHORPE, 1909
Anaspis testaceomarginata ROUBAL, 1935

Subgenus ***Silaria*** MULSANT, 1856

***Anaspis brunnipes* (MULSANT, 1856)**

Silaria brunnipes MULSANT, 1856
Anaspis fuscipes MULSANT, 1856
Anaspis brunnifrons COUCKE, 1891

(-) ***Anaspis quadrimaculata* GYLLENHAL, 1817**

Mordella bipustulata BONELLI, 1812
Silaria quadripustulata P. W. J. MÜLLER, 1821

***Anaspis varians varians* (MULSANT, 1856)**

Mordella frontalis COSTA, 1854
Silaria varians MULSANT, 1856
Anaspis collaris MULSANT, 1856
Anaspis juvenilis SCHILSKY, 1899

Genus ***Cyrtanaspis*** EMERY, 1876

***Cyrtanaspis phalerata* (GERMAR, 1847)**

Anaspis phalerata GERMAR, 1847
Cyrtanaspis obscura SCHILSKY, 1895
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<i>Anaspis (Larisia) palpalis</i> (GERHARDT, 1876)	p. 252
<i>Anaspis (Nassipa) costai</i> EMERY, 1876	p. 254
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INTRODUCTION

In 2012, the catalogue „Coleoptera Poloniae: Tenebrionoidea (Tenebrionidae, Boridae). Critical checklist, distribution in Poland and meta-analysis” appeared, authored by Dariusz IWAN, Daniel KUBISZ and Piotr TYKARSKI. The present book continues the index of beetles belonging to the superfamily Tenebrionoidea occurring in Poland, comprising families of Tetratomidae, Melandryidae, Ripiphoridae, Prostomidae, Oedemeridae, Mycteridae, Pythidae, Aderidae, Scraptiidae.

The most important catalog work of beetles of Poland that has appeared so far is “Katalog Fauny Polski (Catalogus Faunae Poloniae)”, further referred to as KFP, issued in the end of XXth century. The work included, among others, taxa discussed in this book. According to the earlier developed scheme, it contained data on taxonomy, distribution, bibliography and biology of the species which were accounted in literature to the fauna of Poland in its present geographical borders. The families presented herein were described in volumes 13, 14, 22 of “Katalog Fauny Polski (Catalogus Faunae Poloniae)” (BURAKOWSKI et al. 1986, 1987, 2000). Here is a comparison of those and present studies (the number of species occurring in Poland – CS, „confirmed species”; the number of species mistakenly reported from Poland – DS, „doubtful species”, * – former Anaspidae was included to Scraptiidae:

	KFP		Present paper	
	CS	DS	CS	DS
Tetratomidae	2	–	6	–
Melandryidae	32	2	27	6
Ripiphoridae	2	3	3	2
Prostomidae	1	–	1	–
Oedemeridae	22	5	22	10
Mycteridae	1	–	–	1
Pythidae	3	–	3	–
Aderidae	5	2	7	2
Scraptiidae	1	–	15	6
(Anaspidae)	20	1	*	*

Coleoptera Poloniae: Tenebrionoidea

The classification and taxonomic nomenclature used by the authors of KFP were modified in later works.

The taxonomic position of genera included in it has been changed (in some cases, many times) during recent years, due to different opinions of entomologists on the high-level systematics of the order Coleoptera. Species belonging to the genera *Scaptia* and related ones were classified as a separate family of Scaptiidae or included in Tenebrionidae or Melandryidae. Beetles belonging to the genera *Anaspis*, *Cyrtanaspis*, *Pentaria* and several others, not occurring in Poland, were long treated as a subfamily Anaspidinae within Mordellidae, related with them, and this classification remains in most of keys and catalogues used until now. The taxonomy of that group was presented this way, *inter alia*, in the faunistic survey of species of Central Europe (ERMISCH 1956) and the most commonly used identification key (ERMISCH 1969). The present taxonomic classification was applied only in supplement to the key (LOHSE and LUCHT 1992). Subsequently, both subfamilies were given a status of separate families of Anaspidae and Scaptiidae; this view is shared by Katalog Fauny Polski (BURAKOWSKI et al. 1987), or Wykaz zwierząt Polski (MROCKOWSKI and STEFAŃSKA 1991). However, according to the newest and widely accepted taxonomy of beetles (LAWRENCE and NEWTON 1995), those groups are subfamilies of the family Scaptiidae. This has been suggested for quite a long time (CROWSON 1967; FRANCISCOLO 1972; WATT 1987), but has been widely accepted only recently. Polish Scaptiidae *sensu lato* have been previously described in a short key (BOROWIEC and TARNAWSKI 1983), comprising in fact only the genus *Scaptia*.

We adopted taxonomy following "Catalogue of Palaearctic Coleoptera" (LÖBL and SMETANA 2008) – the families were presented by several authors: Tetratomidae (NIKITSKY 2008), Melandryidae (NIKITSKY and POLLOCK 2008), Ripiphoridae (BATELKA 2008), Prostomidae (SCHAWALLER 2008), Oedemeridae (ŠVIHLA 2008), Mycteridae (LÖBL 2008), Pythidae (POLLOCK 2008), Aderidae (NARDI 2008), Scaptiidae (LEBLANC et al. 2008). The number of taxa occurring in Poland (current investigations), in relation to Palearctics, is the following for particular families (species group name – total number or species and subspecies):

	Genera			Species group name		
	Palaearctic	Poland		Palaearctic	Poland	
Tetratomidae	11	4	36%	55	6	11%
Melandryidae	44	15	34%	192	27	14%
Ripiphoridae	15	3	20%	53	3	6%
Prostomidae	1	1	100%	9	1	11%
Oedemeridae	39	6	15%	466	22	5%
Pythidae	2	1	50%	7	3	43%
Aderidae	14	6	43%	104	7	7%
Scaptiidae	11	3	28%	210	15	7%

Introduction

After issuance of KFP (1986, 1987) most of faunistic data were presented as single reports in works of narrow scope. Three synthetic studies are important, comprising five families:

Tetratomidae and Melandryidae. The paper by KUBISZ et al. (2010) based on institutional and private collection data (nearly 2000 voucher specimens) summarized distributions of 37 species of Tetratomidae and Melandryidae currently known to occur in Poland. Two taxa were reported as new in Poland: *Orchesia grandicollis* and *Dircea australis*;

Oedemeridae and Scraptiidae. In 2006, KUBISZ published a monograph on both families, comprising 24 species of the family Oedemeridae and 22 species of Scraptiidae. The paper includes data on biology and distribution, together with history of studies, descriptions, zoogeographical analysis and identification keys for all the taxa at all ranks.

Aderidae. JAŁOSZYŃSKI et al. (2013) presented data on distribution, morphology, and biology of all species of Aderidae occurring in Poland, based on over 370 specimens from private and institutional Polish collections. *Cobosobia pruinosa* (KIESENWETTER) was removed from Polish fauna.

Mycterus curculioides (FABR.) is the only representative of the family Mycteridae reported from the territory of modern Poland. The records from Upper and Lower Silesia from the 19th century (among others, by German researchers KELCH, ROGER, LETZNER, GERHARDT; no voucher specimens in collections) and from Puławy (specimens collected in 1912 by Sz. TENENBAUM) have not been confirmed, which is why the species (and the family as a whole) has been included in the group of species, the current presence of which in Poland is doubtful.

The list and summary of the whole data presented in the present book, together with detailed statistics of particular species and families, are set forth in the chapter „Meta-analysis of occurrence data”.

The data published in the current volume, similarly as in the case of the previous one (IWAN et al. 2012), is integrated into a database of the project *Biodiversity Map*¹ (TYKARSKI 2011a). The project is run by the University of Warsaw as an initiative of the Polish Biodiversity Information Network (KSIB)², providing IT tools for faunistic investigations in Poland. The KSIB database system supports the long-term program *Coleoptera Poloniae*³ aimed at integration of data on beetles of Poland. The data and maps of species distribution are available online.

¹ www.biomap.pl

² Krajowa Sieć Informacji o Bioróżnorodności, www.ksib.pl

³ coleoptera.biomap.pl

SPECIES WITH CONFIRMED PRESENCE IN POLAND

Family **TETRATOMIDAE** BILLBERG, 1820
Subfamily **TETRATOMINAE** BILLBERG, 1820

Genus ***Tetratoma*** FABRICIUS, 1790
Subgenus ***Abstrulia*** CASEY, 1900

Tetratoma ancora FABRICIUS, 1790

Tetratoma picta NEWMAN, 1855
Tetratoma conjuncta PIC, 1901
Tetratoma obscurior PIC, 1901
Tetratoma pfefferi ROUBAL, 1930
Tetratoma perrisi HOULBERT, 1934

Distribution in Poland (Fig. 1)4

Baltic Coast: Gdańsk-Oliwa (Oliva) [CF33], *** (BERCIO and FOLWACZNY 1979); Koszalin (Koszalin) [WA70], *** (LÜLLWITZ 1916); Sopot (Zoppot) [CF43], *** (BERCIO and FOLWACZNY 1979).

Pomeranian Lake District: Wejherowo (Neustadt) [CF25], *** (LENTZ 1879, BERCIO and FOLWACZNY 1979).

Masurian Lake District: “Dęby w Krukach Pasłęckich” nat. res. [DE18], IV–XI 2002, *** (BYK and BYK 2004).

Wielkopolska-Kujawy Lowland: ***

Mazovian Lowland: ***

Podlasie Lowland: ***

Białowieża Primeval Forest: f. comp. 493D [FD83], 2 V 1988, ISEA (KUBISZ 1995); f. comp. 316/317 [FD94], 2 V 1991, AC (KUBISZ et al. 2010); f. comp. 399C [FD94], 5 V 1987, 1991, 1995–1996, 1998–1999, ISEA (KUBISZ 1995, KUBISZ 2004a, KUBISZ et al. 2010); f. comp. 317C [FD95], 1988–1991, ISEA (KUBISZ 1995);

f. comp. 288C/318A [FD95], 1988–1991, 1995–1996, 1998–1999, ISEA (KUBISZ 1995, KUBISZ 2004a); Hajnówka f. div. [a.FD74], V–X 2000, *** (BOROWSKI 2001, BYK 2001b); Białowieża Primeval Forest, without precise locality, *** (BURAKOWSKI et al. 1987); Białowieża Primeval Forest, without precise locality, 1998–2001, *** (BYK et al. 2006).

Lower Silesia: Wrocław (Breslau) [a.XS46], *** (LETZNER 1871, GERHARDT 1890b, 1910).

Trzebnica Hills: ***

Upper Silesia: Zawadzkie (Zawadzki) [CB20], V–VI 1929, *** (NOWOTNY and POLENTZ 1933).

Kraków-Wieluń Upland: “Góra Chełm” nat. res. [CA88], 1996–1997, AC (SZAFRANIEC and SZOŁTYS 1997); Kraków-Sikornik [DA14], 21 VI 1986, ISEA (KUBISZ et al. 2010); Kraków, Skaly Panieńskie [DA14], *** (KULCZYŃSKI 1873); Ojcowski N.P., Mt. Chełmowa Góra [DA16], 24 V 1969, ISEA (PAWŁOWSKI et al. 1994); Ojcowski N. P., Sąspowska valley [DA16], 8 V 1989, ISEA (KUBISZ and PAWŁOWSKI 1998); Ujazd [DA15], 12 IV 1920, ISEA (KUBISZ et al. 2010), *** (BURAKOWSKI et al. 1987).

Małopolska Upland: “Grabowiec” nat. res. [DA79], 28 IV 1990, AC (KUBISZ et al. 2010); Klonów [DA47], 7 V 2006, ISEA (KUBISZ et al. 2010).

Świętokrzyskie Mts.: Trzemoszna f. distr. [DB57], 2006, *** (BOROWSKI 2007).

Lublin Upland: ***

Roztocze Upland: Krasnobród [FB50], *** (BURAKOWSKI et al. 1987); Nart [FB40], 21 IV 1986, AC (KUBISZ et al. 2010).

Sandomierz Lowland: Niepołomice [DA44], *** (BURAKOWSKI et al. 1987).

Western Sudetes: Duszniki-Zdrój (Reinerz) [WR98], VI 1914, *** (KOLBE 1915), ***, MNHW (KUBISZ et al. 2010), *** (BURAKOWSKI et al. 1987).

Eastern Sudetes: Łądek-Zdrój (Bad Landeck) [XR37], VII 1919, *** (SCHOLZ 1929); „Puszcza Śnieżnej Białki” nat. res. (Saalwiesen) [XR37], IX 1926, *** (KOLBE 1928).

West Beskid Mts.: Mt. Babia Góra [CV99], Puszcza Czatożajska Forest, 26 VI 1963, 1 XI 1964, *** (PAWŁOWSKI 1967), 27 VI 1997, ISEA (KUBISZ et al. 2010); Mt. Barania Góra [CV59], VI, *** (KOTULA 1873); Mt. Czantoria (Czantory) [CA40], *** (REITTER 1870a, BURAKOWSKI et al. 1987); Hala Łabowska vicinity [DV78], 9 III 2001, AC (KUBISZ et al. 2010); Krynica-Kopciowa [DV97], 17 V 1989, 24 V 1989, 7 VI 1989, ISEA (KUBISZ et al. 2010); Łapanów [DA42], 20 II 1915, ISEA (KUBISZ et al. 2010), *** (BURAKOWSKI et al. 1987); Mt. Prehyba [DV77], *** (BURAKOWSKI et al. 1987); Mt. Radziejowa [DV77], 30 VI 1987, ISEA (KUBISZ et al. 2010); Mt. Szymbielnia (Kamitzer Platte) [CA51], 7 VII, *** (KOLBE 1908); “Barania nebst Obchodzita und Skrzyczna”, *** (NOWICKI 1873).

Nowy Targ Basin: ***

East Beskid Mts.: Brylińce [FA10], IV–IX, *** (TRELLA 1923); Mt. Helicha [FA21], 1 XI 1886, ISEA (KUBISZ et al. 2010); Krasiczyn [FA21], IV–IX, *** (TRELLA 1923); “Modrzyna” nat. res. [EV57], 8 VII 1987, ISEA (KUBISZ et al. 2010);

Species with confirmed presence in Poland

Prałkowce [FA21], IV–IX, *** (TRELLA 1923), 7 V 1879, 18 V 1884, ISEA (KUBISZ et al. 2010); Przemyśl [FA21], ***, NHC (KUBISZ et al. 2010); Przemyśl-Pikulice [FA21], IV–IX, *** (TRELLA 1923); Mt. Turnica [FV19], IV–IX, *** (TRELLA 1923, 1938); Wapowce [FA11], 1878, ISEA (KUBISZ et al. 2010).

Bieszczady Mts.: Dwernik [FV15], 3 XI 1967, MIZ (KUBISZ et al. 2010); Mt. Hyrlata [EV94], 21 X 1968, ISEA (KUBISZ et al. 2010); Mt. Magura Stuposiańska [FV24], *** (BURAKOWSKI et al. 1987); Polonina Caryńska [FV23], 19 V 1967, ISEA (KUBISZ et al. 2010); Mt. Rozki [EV94], *** (BURAKOWSKI et al. 1987); Smolnik [FV25], *** (BURAKOWSKI et al. 1987); Ustrzyki Górnne [FV23], 24–27 V 1996, AC, 3 IV 1968, MIZ (KUBISZ et al. 2010), *** (BURAKOWSKI et al. 1987); Wetlina [FV04], 25 XI 1953, MIZ, 6 IX 1963, ISEA (KUBISZ et al. 2010); Wetlina PGR [FV04], 22–24 VII 1994, AC (KUBISZ et al. 2010); Widełki [FV24], *** (BURAKOWSKI et al. 1987).

Pieniny Mts.: ***

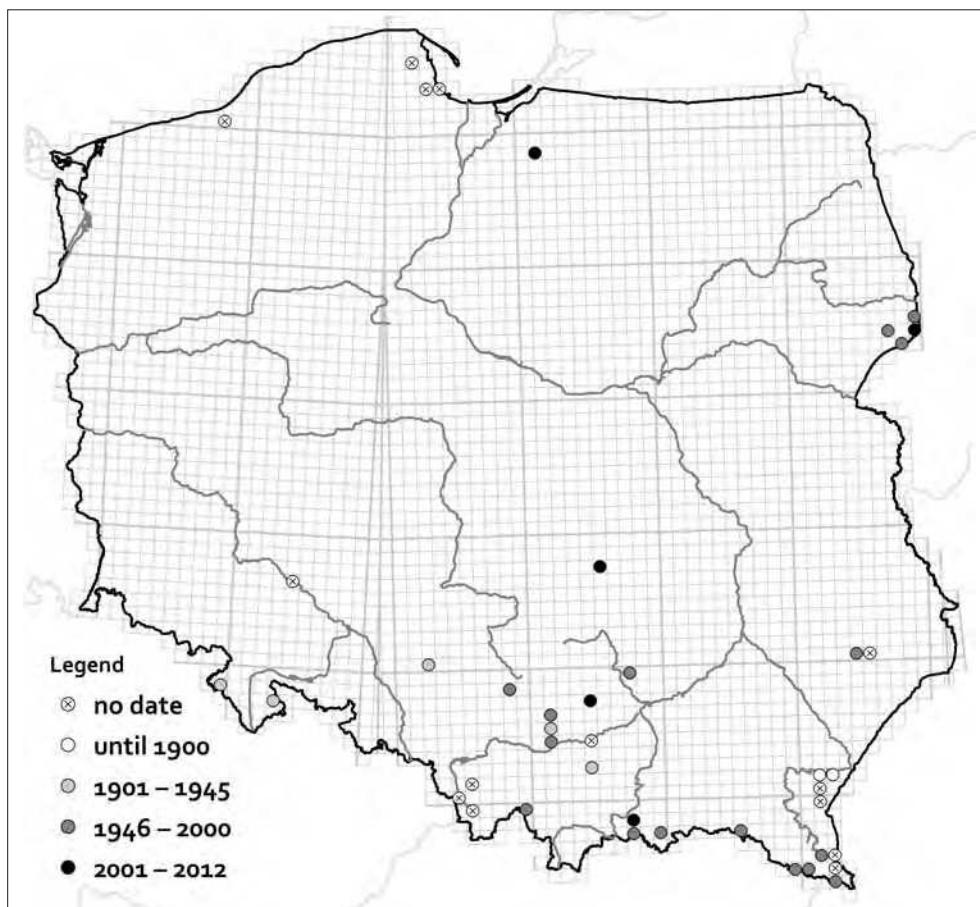


Fig. 1. Occurrence of *Tetratoma ancora* in Poland.

Tatra Mts.: ***

General: Góry Kłodzkie Mts. (Glatzer Gebirge) (GERHARDT 1910).

Comments

KFP: 3756. *Tetratoma ancora* FABRICIUS, 1790.

A rare species, observed mainly in the southern part of Poland, especially in the mountains and foothills (up to 2000 m above sea level), as well as in Białowieża Primeval Forest. A xylomycetophage, connected with broadleaved trees.

For the Palaearctic distribution see Fig. 202.

Subgenus ***Tetratoma*** FABRICIUS, 1790

Tetratoma fungorum FABRICIUS, 1790

Dermestes collaris QUENSEL, 1790

Tetratoma dermestoides HERBST, 1792

Boletaria bicolor MARSHAM, 1802

Distribution in Poland (Fig. 2)

Baltic Coast: Gdańsk (Danzig) [a.CF42], *** (LENTZ 1879, BERCIO and FOLWACZNY 1979); Koszalin (Köslin) [WA70], *** (LÜLLWITZ 1916).

Pomeranian Lake District: Szczecin-Niebuszewo [VV71], 3 I 2006, AC (KUBISZ et al. 2010); Zakrzewo vic. [XV41], road to Wersk, 18 III 2007, AC (KUBISZ et al. 2010); Zalew Koszycki [XU19], 2000–2008, *** (RUTA 2009); Zdrojowa Góra f. div., f. comp. 173/204 [XU19], 29 III 2002, AC (KUBISZ et al. 2010).

Masurian Lake District: “Dęby w Krukach Pasłęckich” nat. res. [DE18], IV–XI 2002, *** (BYK and BYK 2004); Łażek [DE64], 10 XI 2008, ISEA (KUBISZ et al. 2010); Ostróda (Osterode) [DE35], *** (KUGELANN 1794, LENTZ 1857, 1879, BERCIO and FOLWACZNY 1979).

Wielkopolska-Kujawy Lowland: Biedrusko vic. [XU32], military range, 22 I 1998, 18 X 1998, AC (KUBISZ et al. 2010); Boguszyniec [CC48], 15 XII 1988, ISEA (KUBISZ et al. 2010); Dzierawy [CC38], 16 X 1996, ISEA (KUBISZ et al. 2010); Gołuchów [YT04], 5 II 1984, 16 XII 1989, ISEA (KUBISZ et al. 2010); Gruszecka (Birnbäume) [XT50], *** (LETZNER 1871, GERHARDT 1890b, 1910); Karszew [CC57], 18 XII 1999, AC (KUBISZ et al. 2010); Koło [CC48], 28 I 1990, 14 II 1990, ISEA (KUBISZ et al. 2010); Lusowo [XU11], 20 II 1995, 1 III 1998, 21 IV 2003, AC (KUBISZ et al. 2010); Mokrz [WU84], 24 X 2007, AC (KUBISZ et al. 2010); Osiek Mały [CC48], 24 XII 1989, ISEA (KUBISZ et al. 2010); Piła-Górne [XU19], old cemetery, 21 II 1998, AC (KUBISZ et al. 2010); Piła-Kalina vic. [XU28], 16 III 2002, AC (KUBISZ et al. 2010); Piła-Koszyce [XU19], 17 II 2001, AC (KUBISZ et al. 2010);

Poznań [a.XU30], ***(SZULCZEWSKI 1922), ***, NHC (KUBISZ et al. 2010); Poznań-Cytadela [XU31], 1994–2001, ***(JAŁOSZYŃSKI and KONWERSKI 2001); Poznań-Rataje [XU30], Warta river valley, 11 XI 1997, AC (KUBISZ et al. 2010); Promno [XU51], Dębiniec lake vic., 16 I 1999, AC (KUBISZ et al. 2010); Tarnowa [CC37], 20 IX 1987, ISEA (KUBISZ et al. 2010); Żagań [WT21], 5 XII 1986, ISEA (KUBISZ et al. 2010).

Mazovian Lowland: Kampinoski N. P., „Sieraków” nat. res. [DC89], 4 VII and 8 IX 2009, 8 IV 2010, AC (MARCZAK 2010).

Podlasie Lowland: ***

Białowieża Primeval Forest: ***

Lower Silesia: Chwałęcin (Quanzendorf) [XS32], ***(GERHARDT 1910); Dunino [WS76], IX 1913, MNHW (KUBISZ et al. 2010), ***(BURAKOWSKI et al. 1987); Głębocko [XS72], 9 XI 2008, AC (KUBISZ et al. 2010); Legnica (Lieggnitz) [WS87], ***(LETZNER 1871, GERHARDT 1890b, 1910), ***, MNHW (KUBISZ et al. 2010); Lesieńiec [WS27], ***(BURAKOWSKI et al. 1987); Malczyce [XS07], V 1928, MNHW (KUBISZ et al. 2010), ***(BURAKOWSKI et al. 1987); Trzmielów [WS79], ***, MNHW (KUBISZ et al. 2010); Tułowice [XS80], 6 X 1995, AC (KUBISZ et al. 2010); Wrocław (Breslau) [a.XS46], ***(LETZNER 1871, GERHARDT 1890b, 1910); Wrocław-Kotowice [XS37], 18 III 1990, 19 X 1990, AC (KUBISZ et al. 2010); Wrocław-Leśnica (Deutsch Lissa) [XS36], ***(KOLBE 1928); Wrocław-Zakrzów [XS46], 15 IX 1991, AC (KUBISZ et al. 2010); Zakrzów-Kotowice [XS55], 21 IV 1996, AC (KUBISZ et al. 2010); Zimna Woda [WS78], ***, MNHW (KUBISZ et al. 2010).

Trzebnica Hills: ***

Upper Silesia: Kluczbork [CB05], 10 IX 1990, AC, 23 II 1991, AC, 13 X 1996, AC (KUBISZ et al. 2010); „Lipowiec” nat. res. [CA84], 1996–1997, AC (SZAFRANIEC and SZOŁTYS 1997); Kluczbork f. div., Lasowice Wielkie f. distr., f. comp. 108 [CB04], 29 IX 2001, AC (KUBISZ et al. 2010); Kluczbork f. div., Święciny f. distr., f. comp. 240 [YS04], 6 IV 1999, AC (KUBISZ et al. 2010); Kluczbork f. div., Zameczek f. distr., f. comp. 137 [BB94], 28 XI 1999, AC (KUBISZ et al. 2010); Kluczbork f. div., Zofiówka f. distr., f. comp. 126 [CB15], 4 XI 1999, AC (KUBISZ et al. 2010); „Segiet” nat. res. [CA48], 1996–1997, AC (SZAFRANIEC and SZOŁTYS 1997); Tarnowskie Góry [CA49], 15 IV 1985, AC, ISEA (KUBISZ et al. 2010).

Kraków-Wieluń Upland: Ojcowski N.P., Mt. Chełmowa Góra [DA16], 2 IV 2006, AC (KUBISZ et al. 2010); “Sokole Góry” nat. res. [CB72], 8 XII 1999, AC (KUBISZ et al. 2010).

Małopolska Upland: Jasień f. distr. [DC31], 2010–2011, ***(BOROWSKI et al. 2013, RUTKIEWICZ et al. 2013); Tunel [DA28], X 1984, ISEA (KUBISZ et al. 2010); „Żądlowice” nat. res. [DC51], 2010–2011, ***(BOROWSKI et al. 2013).

Świętokrzyskie Mts.: Chełmowa Góra f. distr. [EB03], 2006, ***(BOROWSKI 2007, BYK 2007); Cisów f. distr. [DB92], 2006, ***(BOROWSKI 2007, RUTKIEWICZ 2007); Dębno f. distr. [a.DB93], 2006, ***(MOKRZYCKI 2007); Świętokrzyskie Mts., without precise locality, 2006, ***(MOKRZYCKI 2011).

Lublin Upland: ***

Roztocze Upland: "Nart" nat. res. [FB40], 24 IX 1987, AC (KUBISZ et al. 2010).

Sandomierz Lowland: Puszczka Niepołomicka Forest, "Dębina" nat. res. [DA53] (M. BARAN and T. KLEJDYSZ pers. com.).

Western Sudetes: Brachów [WS76], *** (BURAKOWSKI et al. 1987).

Eastern Sudetes: ***

West Beskid Mts.: Mt. Babia Góra [CV99], Puszczka Czatożańska Forest, 30 V, 29 XI 1963, *** (PAWŁOWSKI 1967), 16–20 IX 2002, AC (KUBISZ et al. 2010); Chybie (Chybi) [CA42], *** (REITTER 1870a); Hala Łabowska vicinity [DV78], 10 III 2001, AC (KUBISZ et al. 2010); Roztoka Mała [DV78], *** (BURAKOWSKI et al. 1987).

Nowy Targ Basin: ***

East Beskid Mts.: Brylińce [FA10], III–IV, X–XI, *** (TRELLA 1923); Mt. Bukowy Garb [FA12], *** (TRELLA 1936); Mt. Cergowa [EV58], 21 IV 1971, ISEA (KUBISZ

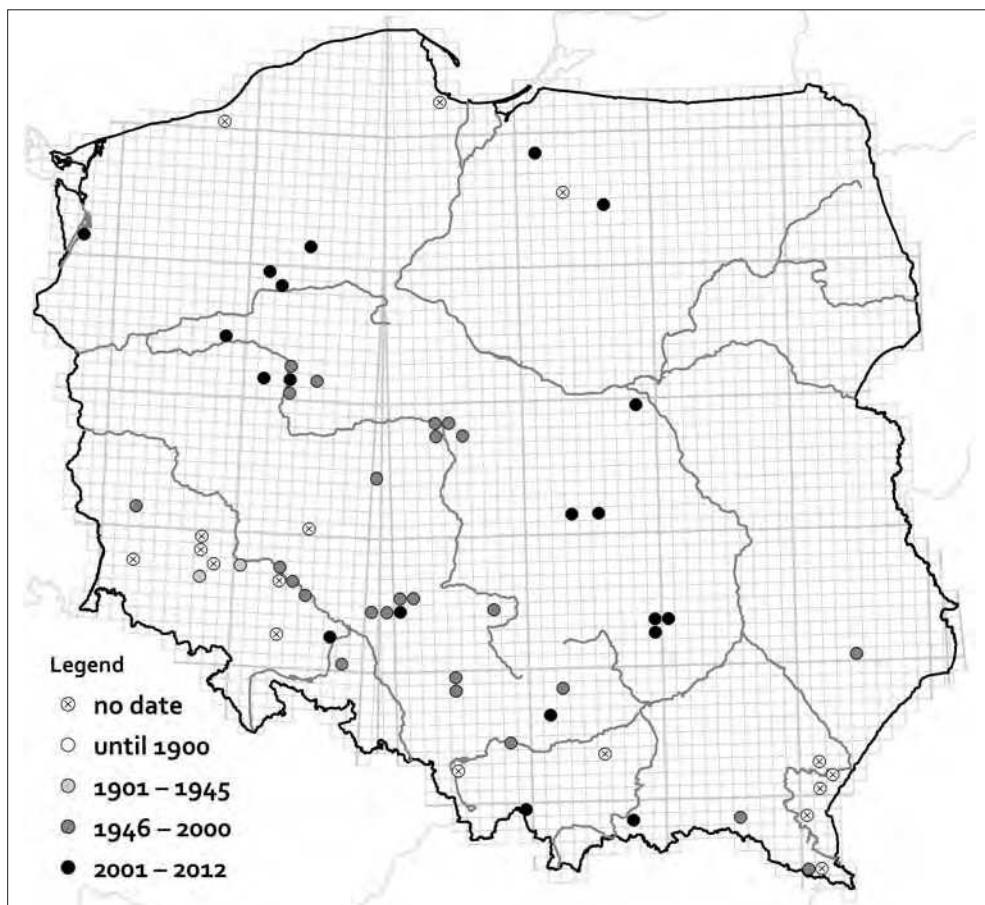


Fig. 2. Occurrence of *Tetratoma fungorum* in Poland.

Species with confirmed presence in Poland

et al. 2010); Mazury [FA21], III–IV, X–XI, *** (TRELLA 1923); Prałkowce [FA21], III–IV, X–XI, *** (TRELLA 1923).

Bieszczady Mts.: Ustrzyki Górne [FV14], *** (BURAKOWSKI et al. 1987); Wetlina [FV04], 9 IX 1963, ISEA (KUBISZ et al. 2010); Mt. Wielka Rawka [FV08], *** (BURAKOWSKI et al. 1987).

Pieniny Mts.: ***

Tatra Mts.: ***

Comments

KFP: 3757. *Tetratoma fungorum* FABRICIUS, 1790.

A xylomycetophage, rather commonly occurring in Poland, although in most cases single individuals are collected. In the present work, the first report from Sandomierz Lowland is included (M. BARAN and T. KLEJDYSZ pers. com.).

For the Palaearctic distribution see Fig. 203.

Subfamily **EUSTROPHINAE** GISTEL, 1856

Tribe **Eustrophini** GISTEL, 1856

Genus ***Eustrophus*** ILLIGER, 1802

Eustrophus dermestoides (FABRICIUS, 1792)

Mycetophagus dermestoides FABRICIUS, 1792
Tritoma sicula BAUDI DI SELVE, 1892

Distribution in Poland (Fig. 3)

Baltic Coast: Gdańsk-Jelitkowo (Gleńkau) [CF43], *** (LENTZ 1879, HORION 1956, BERCIO and FOLWACZNY 1979); Gdańsk-Oliwa (Oliva) [CF33], *** (HORION 1956, BERCIO and FOLWACZNY 1979); Koszalin (Köslin) [WA70], *** (LÜLLWITZ 1916).

Pomeranian Lake District: Kujan vic. [XV41], f. comp. 117, 27 VI 1999, AC (KUBISZ et al. 2010); Szczecin [a.VV72], *** (BURAKOWSKI et al. 1987).

Masurian Lake District: Ostróda (Osterode) [DE35], *** (LENTZ 1879, HORION 1956, BERCIO and FOLWACZNY 1979).

Wielkopolska-Kujawy Lowland: Głębowice vic. [XT20], Łacha river valley, 31 III 2007, AC (KUBISZ et al. 2010); Głogów (Glogau) [WT72], *** (LETZNER 1871, GERHARDT 1890b, 1910); Grodzisk Wielkopolski [WT98], *** (SZULCZEWSKI 1922); Kalisz [BC93], 21 IX 1982, ISEA (KUBISZ et al. 2010); „Krajkowo” nat. res. [XT38], 2–5 VII 2005, *** (MOKRZYCKI et al. 2008); Milicz [XT50], 7 VI 1997, AC (KUBISZ

Coleoptera Poloniae: Tenebrionoidea

et al. 2010); Nowa Sól [WT43], *** (BURAKOWSKI et al. 1987); Obrzycko vic. [XU04], Warta river valley, 19 VI 1997, AC (KUBISZ et al. 2010); Obrzycko [XU04], 23 V 2006, AC (KUBISZ et al. 2010); Poznań-Rataje [XU30], 20 VIII 2003, AC (KUBISZ et al. 2010); Poznań [a.XU30], *** (SCHUMANN 1904, SZULCZEWSKI 1922), ***, NHC (KUBISZ et al. 2010); Rogalin [XT38], 16 V 1998, AC (KUBISZ et al. 2010); Ruszków [CC38], 26 I 2003, AC (KUBISZ et al. 2010); Toruń [CD37], *** (BURAKOWSKI et al. 1987).

Mazovian Lowland: Łomianki [DC99], *** (BURAKOWSKI et al. 1987); Marcelin [EC09], 19 V 1894, USMB (KUBISZ et al. 2010); Serock [ED01], *** (BURAKOWSKI et al. 1987); Szumin [ED33], 10 VIII 1985, ISEA (KUBISZ et al. 2010); Świdry Małe [EC17], *** (BURAKOWSKI et al. 1987); Warszawa [EC08], 5 IX 1895, 7 VI 1897, USMB (KUBISZ et al. 2010); Warszawa-Bielany [DC99], *** (BURAKOWSKI et al. 1987), 14 VI 1895, USMB (KUBISZ et al. 2010); Warszawa-Łazienki [EC08], *** (BURAKOWSKI et al. 1987); Warszawa-Morysin [EC07], *** (BURAKOWSKI et al. 1987); Warszawa-Ogród Zoologiczny [EC08], *** (BURAKOWSKI et al. 1987).

Podlasie Lowland: ***

Białowieża Primeval Forest: f. comp. 186B [FD85], 4 V 1989, ISEA (KUBISZ et al. 2010); Białowieża [FD94], ca. 1880, ISEA (KUBISZ et al. 2010); Białowieża N. P., without precise locality, *** (KARPIŃSKI 1949, BURAKOWSKI et al. 1987), V–X 2000, *** (BOROWSKI 2001, BYK 2001a, BYK et al. 2006).

Lower Silesia: Brzeg (Brieg) [XS73], *** (LETZNER 1871, GERHARDT 1890b, 1910); Dunino [WS76], *** (BURAKOWSKI et al. 1987), ***, MNHW (KUBISZ et al. 2010); Jutrzyna (Marienau) [XS52], *** (LETZNER 1871, GERHARDT 1890b, 1910); Krotoszyce (Kroitsch) [WS76], *** (GERHARDT 1910); Legnica (Liegnitz) [WS87], *** (LETZNER 1871, GERHARDT 1890b); Malczyce [XS07], *** (BURAKOWSKI et al. 1987), 30 VI 1921, ***, MNHW (KUBISZ et al. 2010); Oława [XS64], 28 V 1993, AC (KUBISZ et al. 2010); Pątnów [WS67], *** (BURAKOWSKI et al. 1987), ***, MNHW (KUBISZ et al. 2010); Wrocław-Świniary [XS37], 27 IV 1991, AC (KUBISZ et al. 2010); Wrocław-Szczytniki (Breslau-Scheitnig) [XS46], *** (LETZNER 1871, GERHARDT 1890b, 1910); Wrocław [XS46], Park Szczytnicki, 7 VI 1985, AC, 19 V 1998, AC (KUBISZ et al. 2010); Wrocław [XS46], V 1938, ***, MNHW (KUBISZ et al. 2010); Wrocław-Wojnów [XS56], 8 IV 2006, AC (KUBISZ et al. 2010).

Trzebnica Hills: Oborniki Śląskie [XS38], 24 IV 2008, AC (KUBISZ et al. 2010); Twardogóra (Festenberg) [XS79], *** (LETZNER 1871, GERHARDT 1890b, 1910).

Upper Silesia: Blachownia-Ostrowy [CB52], 12 VII, *** (LGOCKI 1908); “Łęczak” nat. res. [CA05], 1996–1997, AC (SZAFRANIEC and SZOLTYS 1997); Rudy Raciborskie (Rauden) [CA16], *** (KELCH 1852, ROGER 1856, LETZNER 1871, GERHARDT 1890b, 1910); Zawadzkie [CB20], 29 V 1929, USMB (KUBISZ et al. 2010); Ziemięcice [CA38], 29 V 1992, ISEA (KUBISZ et al. 2010).

Kraków-Wieluń Upland: Kraków-Bodzów [DA14], *** (BURAKOWSKI et al. 1987); Kraków [DA24], Botanical Garden, *** (BURAKOWSKI et al. 1987), 17 IX 1892, 22 VI 1936 – 10 VI 1938, ISEA (KUBISZ et al. 2010); Kraków-Dąbie [DA24], *** (BURAKOWSKI et al. 1987), 12 VII 1892, ISEA (KUBISZ et al. 2010); Kraków-

SPECIES WITH DOUBTFUL OR NOT CONFIRMED PRESENCE IN POLAND

Family MELANDRYIDAE LEACH, 1815

Abdera biflexuosa (CURTIS, 1829)

Elater flexuosus A. G. OLIVIER, 1790

Mordella bifasciata MARSHAM, 1802

Hypulus biflexuosus CURTIS, 1829

Abdera bifasciata STEPHENS, 1832

Dircaea griseoguttata FAIRMAIRE, 1850

Abdera undata PERRIS, 1852

Reported occurrence data

General: Silesia (Schlesien) (SCHILSKY 1888, SEIDLITZ 1891).

Comments

KFP: *Abdera biflexuosa* (CURTIS, 1829).

DIETL (1899) stated that the species was erroneously reported by SEIDLITZ (1891) from Nadelwald Schlesien – the data in fact pertain to *Cyrtanaspis phalerata* (GERMAR, 1847).

For the Palaearctic distribution see Fig. 100.

Abdera quadrifasciata (CURTIS, 1829)

Hypulus quadrifasciata CURTIS, 1829

Reported occurrence data

Lower Silesia: Barszów (Barschau bei Glogau) [WT80], *** (LETZNER 1883).

Comments

KFP: *Abdera quadrifasciata* (CURTIS, 1829).

The species inhabits mainly southern and western Europe, and has never been reliably reported from Poland. The only locality from where it has been found (Barszów near Lubin) does not exist any more; at present, there is water reservoir Żelazny Most, built in the 1970s. SEIDLITZ (1898) was the first to question the report by LETZNER (together with a note by LENTZ (1879) from East Prussia) as a „fatal error” in identification of *Cyrtanaspis phalerata*. Further general reports on its occurrence in Silesia and in former East Prussia (e.g., SCHILSKY 1909, REITER 1911, ŁOMNICKI 1913) were based only on these two incorrect notes.

For the Palaearctic distribution see Fig. 102.

Orchesia grandicollis ROSENHAUER, 1847

Orchesia laticollis L. REDTENBACHER, 1849

Reported occurrence data

Eastern Sudetes: Mt. Śnieżnik Kłodzki (Glatzer Schneeberg) [XR37], VII 1893, *** (GERHARDT 1895c, 1910).

Tatra Mts.: Jaszczyrówka vic. [DV26], 12 XI 1872, NHC (KUBISZ et al. 2010); Tatra Mts., without precise locality, *** (KOTULA 1873, NOWICKI 1873).

Comments

KFP: –. *Orchesia grandicollis* ROSENHAUER, 1847.

A montane species; records come from the Tatra Mts. (KOTULA 1873, KUBISZ et al. 2010), however, its present occurrence in Poland requires confirmation. Larvae and adults can be found in rotten wood of trunks and branches of broadleaved trees, under decaying leaves and in the moss growing on old tree.

For the Palaearctic distribution see Fig. 180.

Phloiotrya subtilis (REITTER, 1897)

Dircea subtilis REITTER, 1897

Reported occurrence data

Białowieża Primeval Forest: Hajnówka f. div. [a.FD74], V–X 2000, *** (BOROWSKI 2001).

Species with doubtful or not confirmed presence in Poland

West Beskid Mts.: Cieszyn vic. [a.CA21], *** (REITTER 1911).

East Beskid Mts.: Przemyśl [FA21], *** (REITTER 1897).

Comments

KFP: 3775. *Phloiotrya subtilis* (REITTER, 1897).

A species described by REITTER (1897) on a basis of a single specimen of female, collected by M. RYBIŃSKI – *locus typicus*: Przemyśl (no voucher specimens in the collections of ISEA).

For the Palaearctic distribution see Fig. 188.

Classified in the category CR on the Polish Red List (PAWŁOWSKI et al. 2002).

***Zilora elongata* J. R. SAHLBERG, 1881**

Reported occurrence data

Białowieża Primeval Forest: Stara Białowieża [FD84], 28 VII 1987, MIZ (KUBISZ et al. 2010).

Comments

KFP: absent.

In the publication by KUBISZ et al. (2010), the species is cited as “*Zilora cf. elongata* SAHLBERG, 1881”, with the following comment: “*Zilora elongata* is a boreal species, distributed from Primorski Kray to Europe, where it has been hitherto reported to occur in the north-eastern areas, from Russia to Belarus (NIKITSKY and POLLOCK 2008). Two specimens collected in the Białowieża Primeval Forest show several features that correspond with those given in the original description of *Z. elongata* (SAHLBERG 1881) and in the identification key in SEIDLITZ 1898. However, due to problems discussed in the comment for *Z. obscura* (see above), reliable determination is not possible. *Zilora elongata* was reported to develop in a rotten wood of coniferous trees infested by fungi *Trichaptum abietinum* (DICKS.) RYVARDEN and *Trichaptum fuscoviolaceum* (EHRENB.) RYVARDEN (Polyporaceae) (NIKITSKY 1992)”.

Reported locations of the species are shown together with other species of *Zilora* at Fig. 31.

For the Palaearctic distribution see Fig. 207.

***Zilora ferruginea* (PAYKULL, 1798)**

Xylita ferruginea PAYKULL, 1798

Reported occurrence data

Wielkopolska-Kujawy Lowland: Gołaszyn vic. [XU23], military range, 24 X 2006, AC (KUBISZ et al. 2010); Mokrz [WU84], 1 VI 1996, 5 V 2003, 8 V 2003, 21 IV 2004, AC (KUBISZ et al. 2010).

Upper Silesia: Tarnowskie Góry [CA49], 24 III 1984, ISEA (KUBISZ et al. 2010).

Roztocze Upland: Kosobudy f. distr. [FB41], 21 VII–8 VIII 1902, *** (TENENBAUM 1913).

Bieszczady Mts.: Mt. Rożki [EV94], 3 VI 1970, MIZ (BURAKOWSKI 1971).

Comments

KFP: 3781. *Zilora ferruginea* (PAYKULL, 1798).

In the publication by KUBISZ et al. (2010), the species is cited as “*Zilora cf. ferruginea* (PAYKULL, 1798)”, with the following comment: “This rare species of still unclear taxonomic status has been previously recorded only from the Bieszczady Mts. The record from Roztocze cited in KFP (as TENENBAUM 1913, 1923) is uncertain. Szymon TENENBAUM did not collect this species himself, and his record refers to an unpublished manuscript by J. HORNTZIL (TENENBAUM 1913: 3, 47); this information was repeated in TENENBAUM (1923). Several localities where specimens showing features of *Z. ferruginea* were collected are listed above. However, unclear and apparently variable diagnostic characters and taxonomic problems make reliable determinations impossible”.

Reported locations of the species are shown together with other species of *Zilora* at Fig. 31.

For the Palaearctic distribution see Fig. 208.

Family RIPIPHORIDAE GEMMINGER et HAROLD, 1870

Macrosiagon bimaculata (FABRICIUS, 1787)

Mordella tricuspidata GMELIN, 1790

Rhipiphorus carinthiacus PANZER, 1794

Rhipiphorus bicolor A. G. OLIVIER, 1795

Mordella larvata SCHRANK VON PAULA, 1789

Rhipiphorus lituratus FABRICIUS, 1798

Rhipiphorus quadrimaculatus GYLLENHAL, 1817

Rhipiphorus binotatus FISCHER VON WALDHEIM, 1829

Rhipiphorus apicalis STEVEN, 1832

Rhipiphorus binotatus FISCHER VON WALDHEIM, 1832

Rhipiphorus nigrithorax STEVEN, 1832

Rhipiphorus angustulus MOTSCHULSKY, 1849

Macrosiagon pallidipes REITTER, 1898

META-ANALYSIS OF THE OCCURRENCE DATA

Introduction

The rich material presented in this volume deserves a summary of information at species and family levels. The purpose of this chapter is to extract some parameters, obscured in the main catalogue part, describing history, intensity and completeness of entomological research on the particular group of insects. Beside its basic review role, this part may be useful for planning future research activities.

Methods

The distribution data were converted into a database, further augmented with GIS extensions necessary for geographic analyses. As some source papers did not provide complete detailed information on occurrence dates and/or localities of specimens used, and filling these gaps would be too time consuming, we decided not to consider in this chapter records meant as species-date-locality units. For this reason, the presented tables and general maps do not contain counts of records or specimens.

Granularity of the location information was diverse, from quite accurate village names, through grid-based UTM 10×10 km squares, to a more or less inaccurate regional level. A basis for spatial operations in this chapter was UTM grid, and centroid coordinates of UTM grid squares were assigned to larger units, where necessary. This procedure was used for calculations of the number of districts and regions (see below) from which occurrence of species was reported. Should a centroid fall outside a country border line, then the UTM square was assigned to a region nearest to its centroid.

In the presented analyses, we use the following terms to describe distribution of each taxon: „coverage” as an estimate of real area occupied, expressed as a number of UTM squares; „extent” as a measure of spread, a proxy of which is a number of regions. The term „regions” used here refers to lower-level areas of a proposed regionalisation system (TYKARSKI 2011b), based on the physiogeographical regionalization of Poland by KONDRAKCI (2002), planned to become a successor of the inaccurate KFP regions. The purpose of their use in the analysis was to provide a convenient way for approximations of the physical extent of species distribution in the country, not connected to an artificial administrative division and sufficiently detailed for analyses of distribution at the country level (Fig. 85A, B). In the current and the previous volume (IWAN et al. 2012) we made use of several methods of mapping species distributions, keeping the KFP division (Fig. 85A) for traditionally fashioned listing occurrence sites in the catalogue part, UTM grid coordinates for the localities themselves and species maps, and different systems in meta-analyses. The Biodiversity Map project, its website and on-line tools made it possible to integrate existing mapping systems used in Polish faunistics, which was not possible before. These achievements let us easily switch between or overlay regional divisions, provided that accurate enough occurrence locality data are available.

The maps and spatial calculations for this volume were prepared with ArcGIS Desktop 10.0.

Research dynamics

The first published reports on occurrence of the families discussed herein pertained to *Pyro despressus* and *Tetratoma fungorum*, both written by KUGELANN in 1794. Together with notices by ILLIGER (1798) on *Serropalpus barbatus* and *Orchesia fasciata*, these records came from vicinities of Ostróda in the north of Poland. Not much later, in 1806, another German author, WEIGEL, mentioned *Anaspis maculata* and *Oedemera nobilis* from Silesia. Due to historical circumstances, the first half of the century of entomological research in Poland was dominated by German authors. The first Polish entomologist documenting occurrence of the tenebrionoids was NOWICKI, who discovered, among others, *Melandrya barbata* in 1864.

The number of species in most of the eight Tenebrionoidea families known from Poland apparently stabilised in the beginning or 20th century or still earlier, and maintained it almost to the end of the 20th century (Fig. 86). The

exception was probably the most taxonomically difficult Scaptiidae, showing a significant increase in the total number of species known from Poland in 1920–30. In almost all groups except Prostomidae and Pythidae, new species were recorded in the turn of the 21st century. This fact must have been

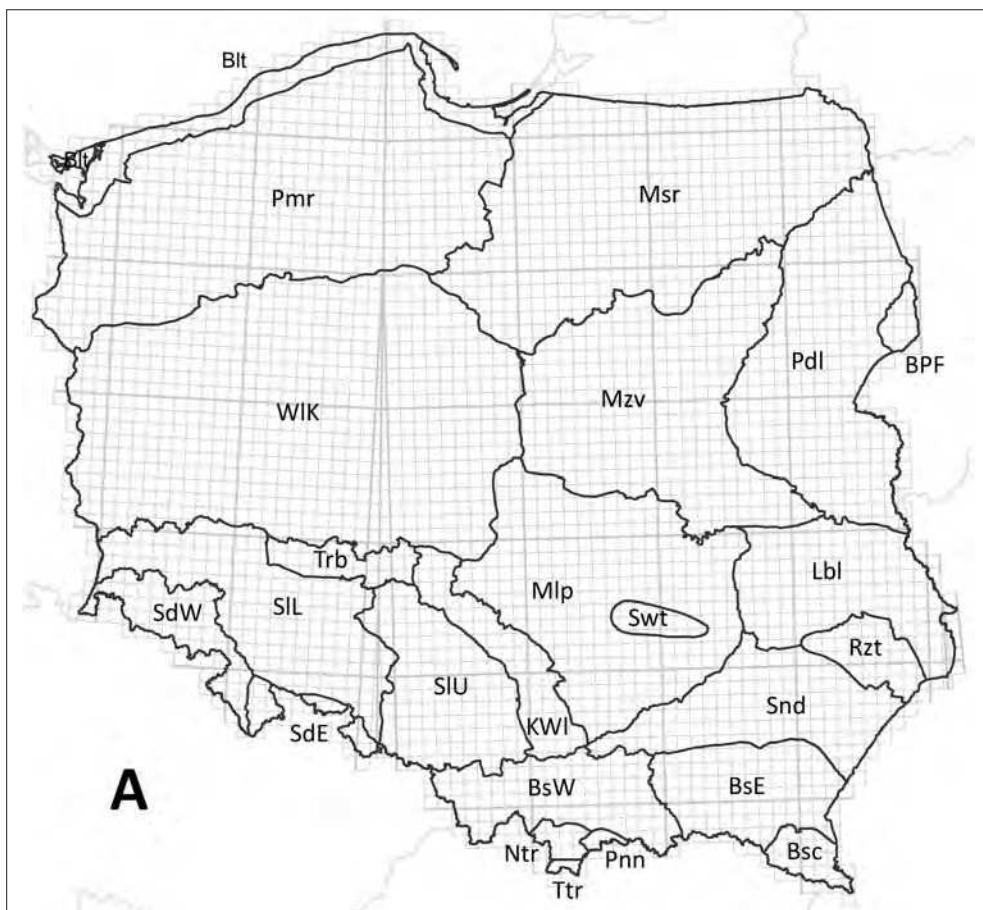


Fig. 85A. Regionalisation systems and grids used in this volume. UTM grid in light gray. Small squares in most cases have a width of 10 km. Regional division (black) of Poland introduced in the Catalogus Faunae Poloniae (KFP), referred to in the text as KFP regions. Blt – Baltic Coast, BPF – Białowieża Primeval Forest, Bsc – Bieszczady Mts., BsE – East Beskid Mts., SdE – Eastern Sudetes, KWl – Kraków-Wieluń Upland, SIL – Lower Silesia, Lbl – Lublin Upland, Mlp – Małopolska Upland, Msr – Masurian Lake District, Mzv – Mazovian Lowland, Ntr – Nowy Targ Basin, Pnn – Pieniny Mts., Pdl – Podlasie Lowland, Pmr – Pomeranian Lake District, Rzt – Roztocze Upland, Snd – Sandomierz Lowland, Swt – Świętokrzyskie Mts., Ttr – Tatra Mts., Trb – Trzebnica Hills, SIU – Upper Silesia, BsW – West Beskid Mts., SdW – Western Sudetes, WIK – Wielkopolska-Kujawy Lowland.

connected with a boost of entomological activity reflected by a two- or three-fold increase in the number of published papers which occurred at that time (Fig. 86, dotted lines). The intensive growth of publishing activity was correlated with increase in a number of occurrence localities reported in publications, which started in 70' and greatly enhanced at the end of

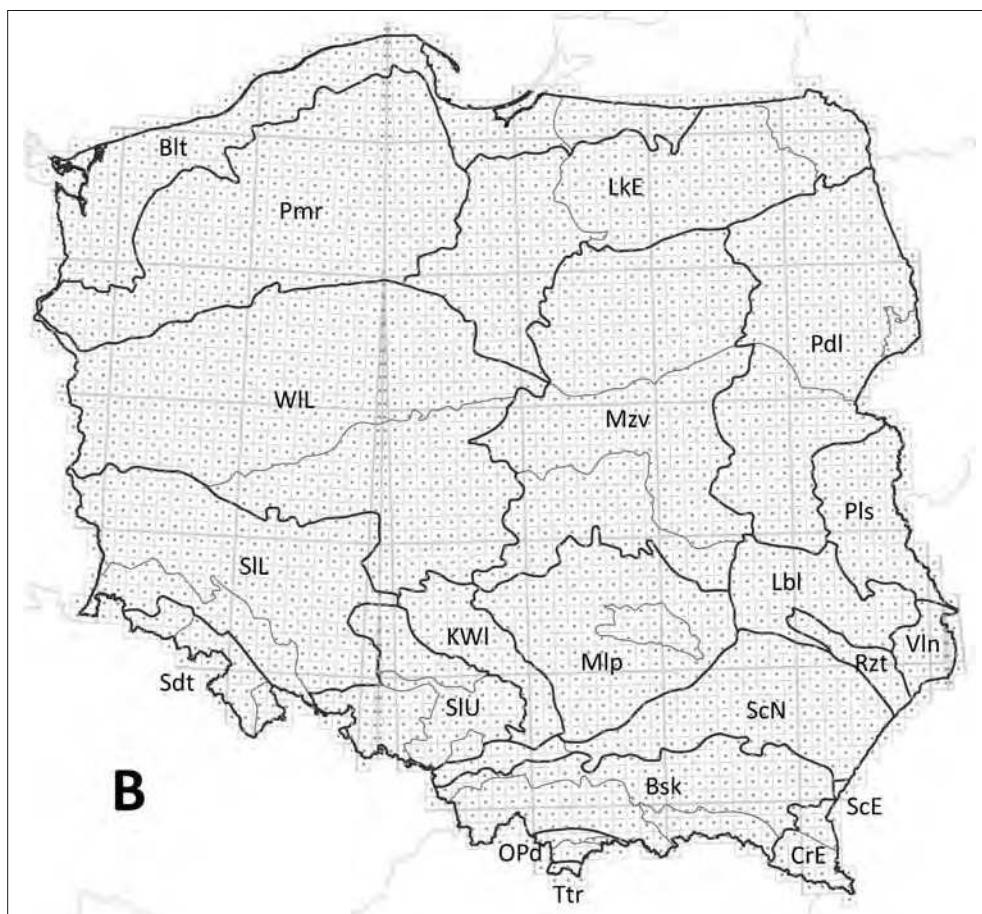


Fig. 85B. Regional division (main units in black, subunits in dark grey) used for estimating within-country extent of distribution of species, calculated by assigning centroids (dots) of UTM squares of species occurrences to a containing unit (for details see Methods). Blt – Baltic Coastlands, Bsk – Beskid Mts. and Foothills, CrE – Eastern Carpathians, KWl – Kraków-Wieluń Upland, Lbl – Lublin Upland, LkE – Eastern Lakelands, Mlp – Małopolska Upland, Mzv – Mazovia, OPd – Orawa-Podhale, Pdl – Podlasie, Pls – Polesie, Pmr – Pomeranian Lakeland, Rzt – Roztocze, ScE – Eastern Subcarpathia, ScN – Northern Subcarpathia, Sdt – Sudety Mts., SIL – Lower Silesia, SIU – Upper Silesia, Ttr – Tatra Mts., Vln – Volhynian Upland, WIL – Wielkopolska-Lubusz.

20th century (Fig. 87). The total count of UTM squares for records of the analysed families varied from ca. 650 for Oedemeridae (being still ca. 20 per species on average) to as few as 9 for Prostomis and 45 for Ripiphoridae.

The recent increase of species counts and publication activity does not necessarily reflect the increase of actual research efforts. Apart from taxonomical changes, clarifications and divisions of earlier described species, this may be partially the effect of extensive queries in materials stored in entomological collections in Poland that revealed data collected earlier (e.g. KUBISZ 2006). Nevertheless, entomological research *sensu stricto*, so necessary for recognition of today's state of Polish fauna, is doubtlessly intensified in the current period.

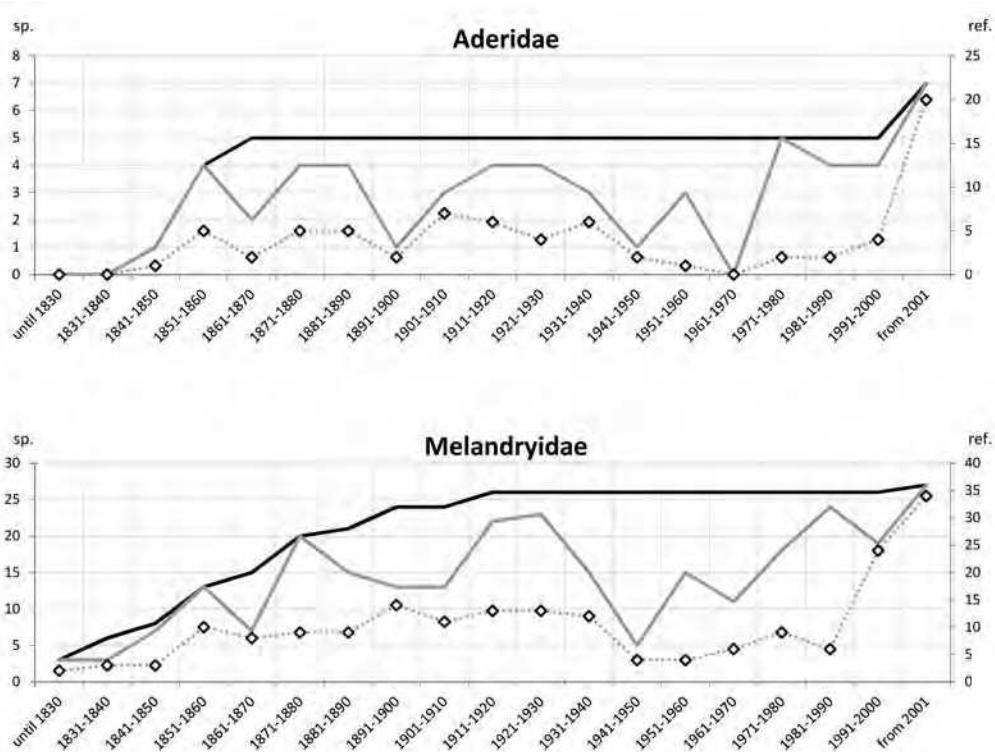


Fig. 86A. Changes over time in number of species and number of publications on tenebrionoid families. Abbreviations: cum. sp. cnt. – cumulative count of species, sp. cnt. – count of species per period, ref. cnt. – count of publications per period.

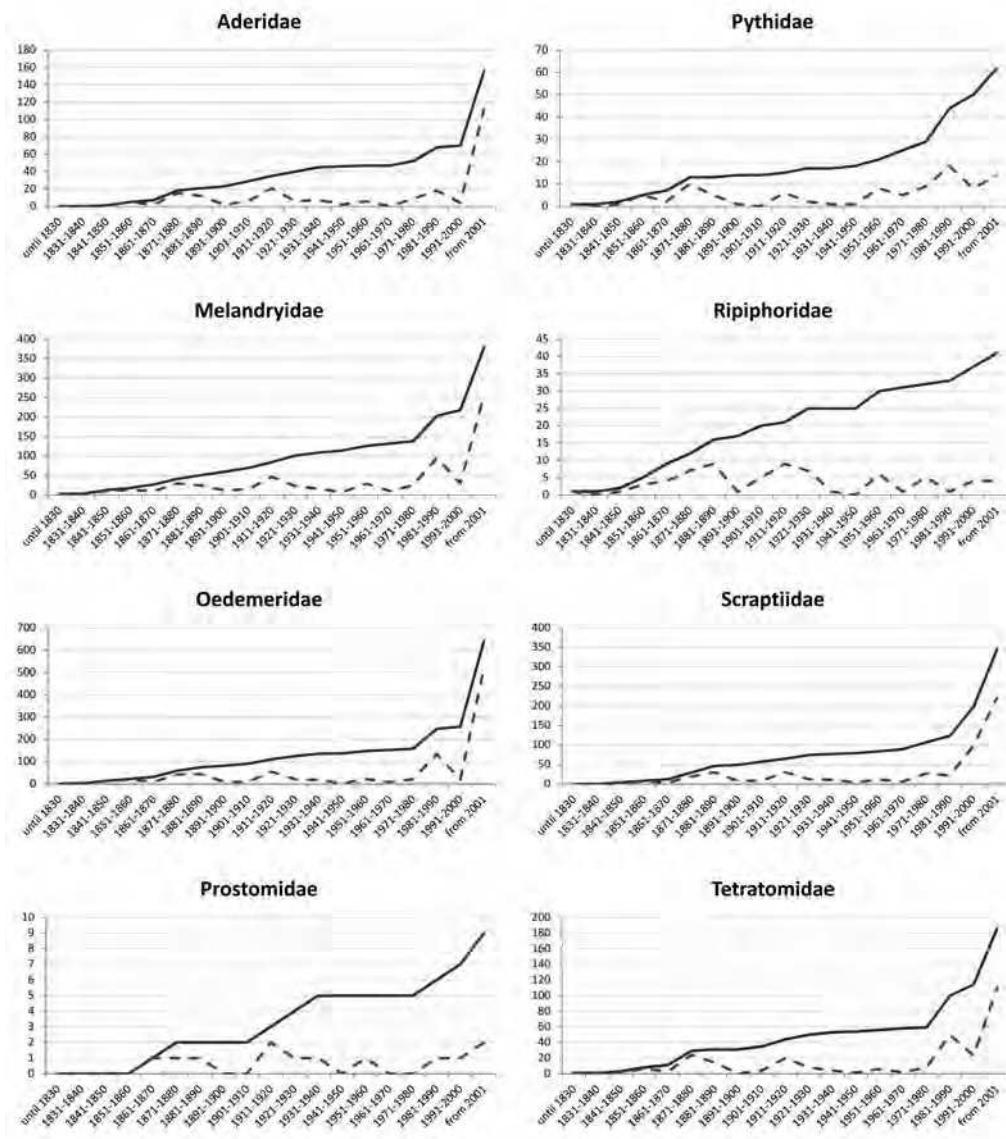


Fig. 87. Changes over time in number of UTM squares correspondent to original localities of tenebrionoid studies in the analysed sources. Abbreviations: cum. UTM sq. cnt. – cumulative count of UTM squares, UTM sq. cnt. – count of UTM squares per period.

Source publications and entomological collections versus sites of data origin

The data came from 887 UTM squares, which is ca. 26% of the total 3384 grid squares for Poland. The extent of exploration of individual localities was very unequal, depending also on a family. In each case, there are squares with no data on time (Figs. 88a–94a, for Prostomidae and its single representative in Poland, see Fig. 37). This is very distinct in more species-rich families like Oedemeridae (Fig. 90) or Scraptiidae (Fig. 93). Fortunately, a considerable part of these inaccuracies is of rather technical nature, resulting from the fact that some authors of source papers did not publish these data although they exist on labels of voucher specimens. The missing data can be acquired and

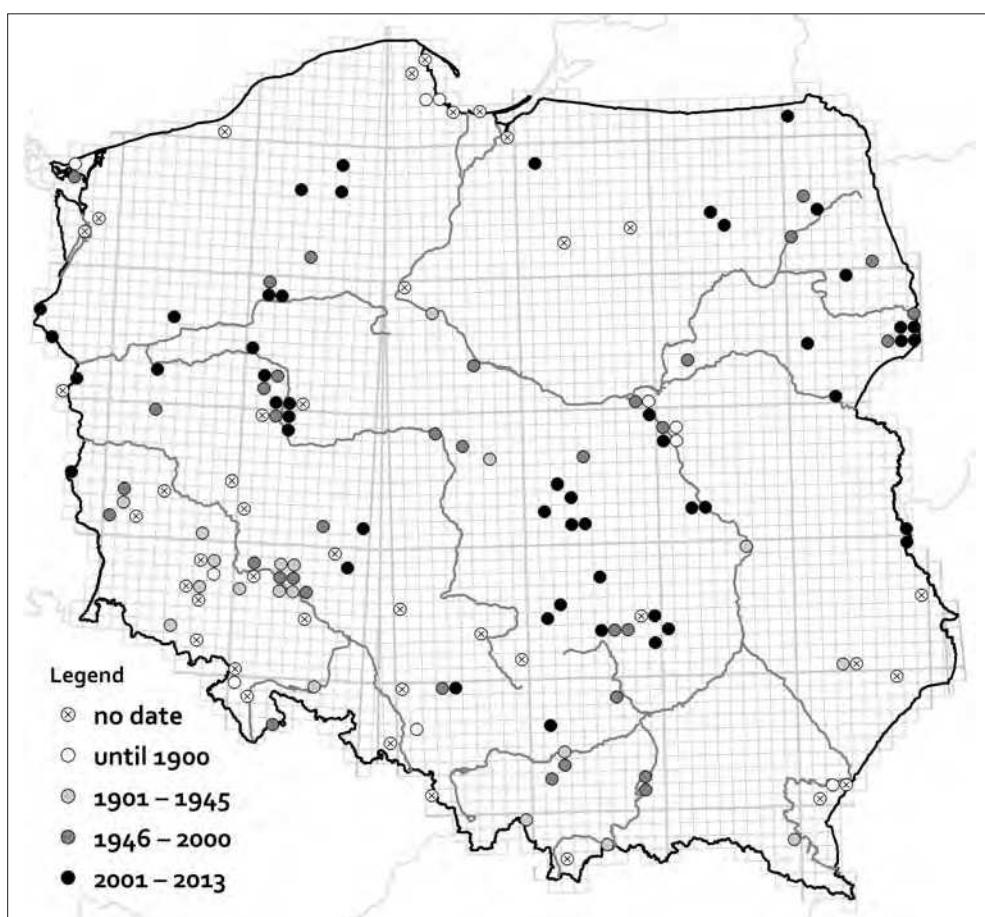


Fig. 88a. Year of last published research on Aderidae per utm square.

followed by the collection of MIZ (Table 1), 75 and 72 respectively, although it is highly probable that there are more species scattered through private collections, awaiting to be revealed. It is also quite possible that a thorough query in institutional collections would also yield additional data. Only ISEA and private collections hold species absent from the rest of the analysed collection resources (at least based on published data): *Anaspis arctica*, *A. bohemica*, *A. costai*, *A. kiesenwetteri*, *A. melanostoma*, *A. palpalis*, *A. ruficollis*, *Nacerdes carniolica*, *Orchesia fusiformis*, *Pelecotoma fennica*, *Prostomis mandibularis*, *Pseudanidorus pentatomus*, and *Ripidius quadriceps* (Table 2).

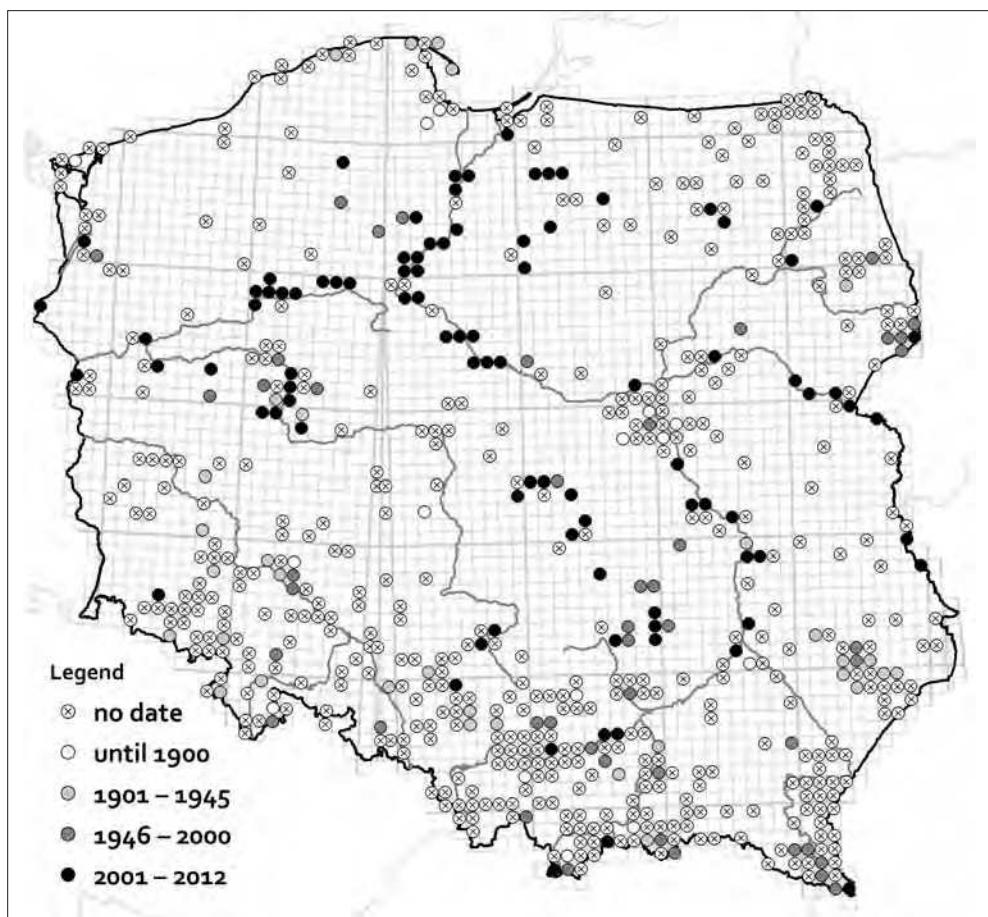


Fig. 90a. Year of last published research on Oedemeridae per utm square.

Table 1. Summary of information obtained from collections in the analyzed material.

ISEA – Institute of Systematics and Evolution of Animals PAS in Cracow, MIZ – Museum and Institute of Zoology PAS in Warsaw, Private – private collections, MNHW – Museum of Natural History, Wrocław University, Wrocław, NHC – Natural History Collections, A. Mickiewicz University, Poznań, USMB – Upper Silesian Museum in Bytom.

Collection	Species cnt.	Unique Species cnt.	UTM square cnt.	Unique UTM square cnt.
Private	72	6	241	96
ISEA	75	7	500	310
MIZ	56	–	149	51
MNHW	28	–	37	21
NHC	10	–	5	–
USMB	40	–	80	29

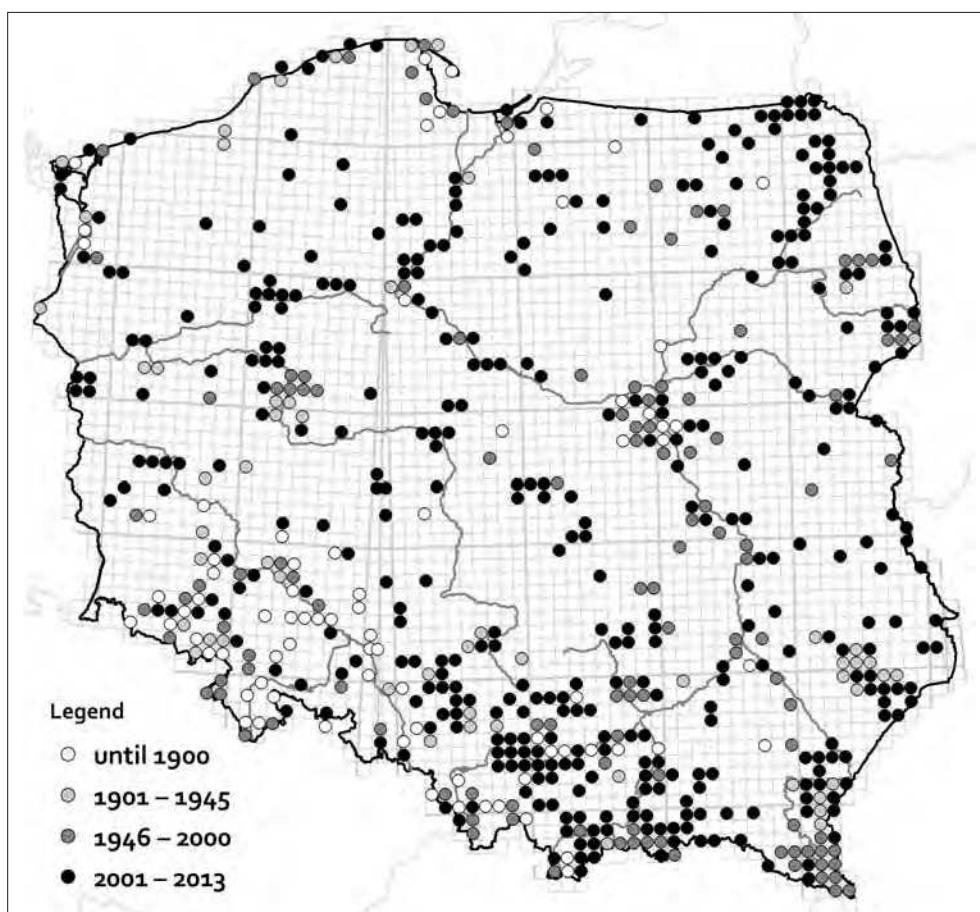


Fig. 90b. Year of the first publication on Oedemeridae per utm square.

Table 2. Summary of data on species of the discussed families occurring in Poland, based on the analysed material.
 Column headers: UTM – number of UTM 10×10 km squares, districts – count of districts, regions – count of regions (see Methods), p. first yr – first year of publication, p. last yr – last year of publication, refs – references count, r. first yr – year of the first record, r. last yr – year of the last record, collections – collections holding specimens of the species; P – private collections, I – ISEA, M – MIZ, U – USMB, Po – NHC, Wr – MNHW

species	UTM	districts	subareact	p.first yr	p.last yr	refs	r.first yr	r.last yr	collections
Aderidae									
<i>Aderus populeus</i>	52	38	18	1856	2013	22	1898	2010	PIMUWr
<i>Anidorus nigrinus</i>	77	61	21	1850	2013	34	1854	2013	PIMWr
<i>Euglenes oculatus</i>	28	21	12	1854	2013	21	1854	2010	PIMUWr
<i>Euglenes pygmaeus</i>	36	31	18	1856	2013	34	1865	2007	PIMWr
<i>Phytobaenus amabilis amabilis</i>	8	7	6	1863	2013	13	1900	2001	PIMU
<i>Pseudanidorus pentatomus</i>	4	3	3	2002	2013	2	1994	2007	A
<i>Vanonus brenicornis brevicornis</i>	3	3	3	2008	2013	2	2000	2011	P
Melandryidae									
<i>Abdera affinis</i>	49	42	21	1846	2013	27	1854	2010	PIMPoWr
<i>Abdera flexuosa</i>	49	42	24	1853	2013	33	1884	2010	PIMUWr
<i>Anisoxya fuscula</i>	22	21	8	1866	2010	19	1888	2008	PM
<i>Conopalpus testaceus</i>	39	34	18	1895	2010	16	1894	2008	PIMUWr
<i>Dircea australis</i>	3	1	1	2010	2010	1	1986	1993	P
<i>Dircea quadriguttata</i>	4	3	3	1879	2010	10	1790	2000	PIM
<i>Dolotarsus linioides</i>	26	18	14	1875	2013	22	1869	2010	P
<i>Hypulus bifasciatus</i>	56	42	20	1837	2013	30	1854	1999	PIMU PoWr
<i>Hypulus quercinus</i>	23	20	15	1833	2013	25	1832	2010	PIMU Po
<i>Melandrya barbata</i>	15	14	13	1864	2012	16	1878	2011	P
<i>Melandrya caraboides</i>	42	34	15	1830	2010	33	1829	2003	PIMPoWr
<i>Melandrya dibia</i>	47	33	21	1854	2013	27	1854	2010	PIMU
<i>Orchesia blandula</i>	26	16	11	1883	2010	9	1872	2004	PIMWr

The numbers of species represented by specimens held in collections are to some extent reflected also in the number of UTM squares visited by collectors of the specimens, 500 for ISEA, 241 for private and for 149 for the MIZ collection (Fig. 97, Table 1). The ISEA collection turned out to be the most successful in gathering specimens from locations that were visited by no one else (310 unique UTM squares, which is more than one third of the total UTM count in the analysed material).

Only data from the MNHW collection turned out to be biased to areas surrounding a city hosting the collection. Sampling spots of the

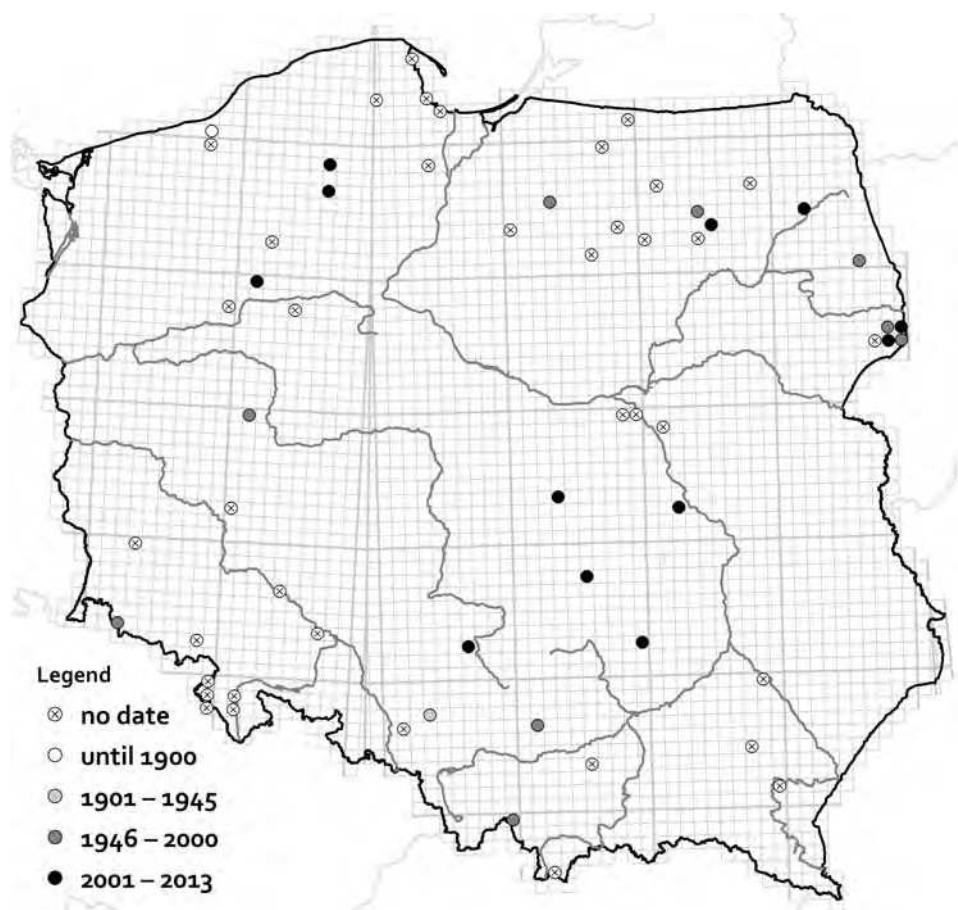


Fig. 91a. Year of last published research on Pythidae per utm square.

other collections were more or less spread over the country. There is, however, a clear tendency that some localities must have been especially attractive for collectors, as specimens from them are held in 3 or more collections. The other reason may be an exchange of specimens between collections. The group of such places includes Ojców National Park, the cities and vicinities of Kraków and Warszawa, some locations near Zakopane (Tatras), Zwierzyniec (Roztocze Upland) and Przemyśl (East Beskid).

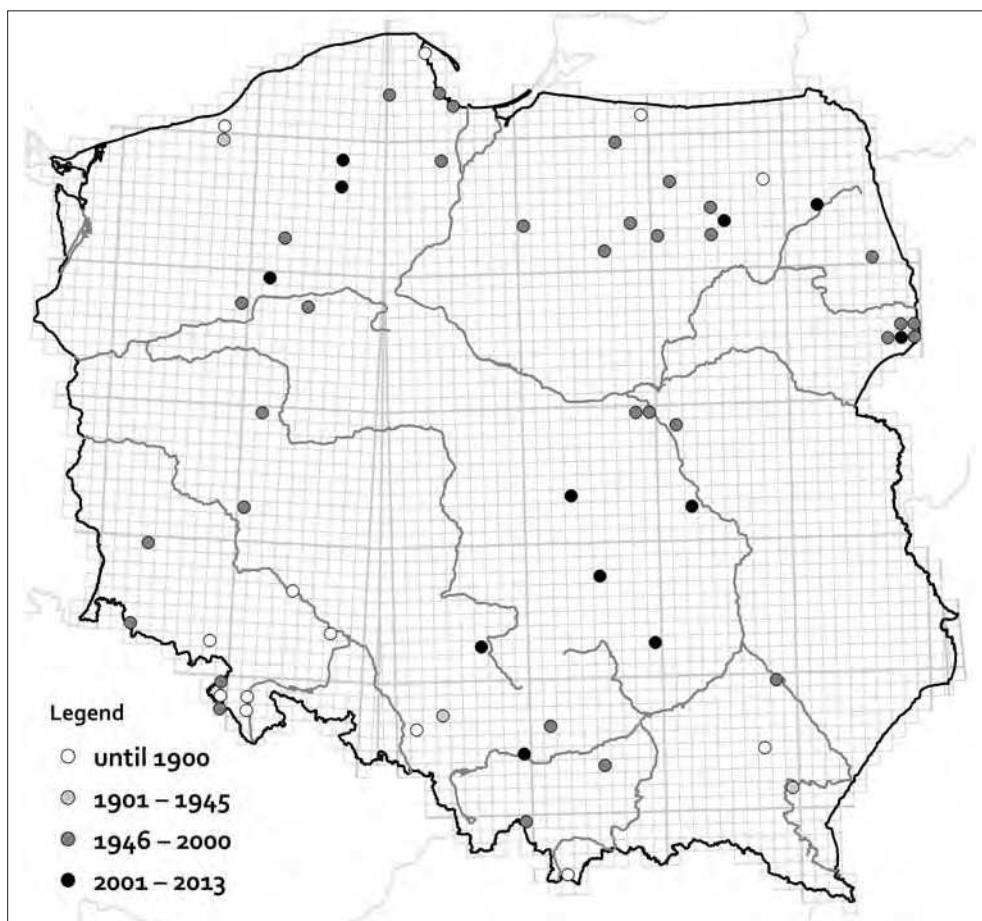


Fig. 91b. Year of the first publication on Pythidae per utm square.

although occurring frequently. In both cases, the number of the reporting publications may be relatively low. An opposite example may be less obvious – a case of species, limited to a few habitats/areas, documented in relatively many papers. By definition, a rare insect gives few chances to be observed and reported, and such papers cannot be numerous, although well known. How can one distinguish between such „well known rarities” and representatives of taxonomically difficult groups with obscure ecology that did not attract attention of entomologists except for a few specialists or invasive species seen at the beginning of their march through the country? The best clarification

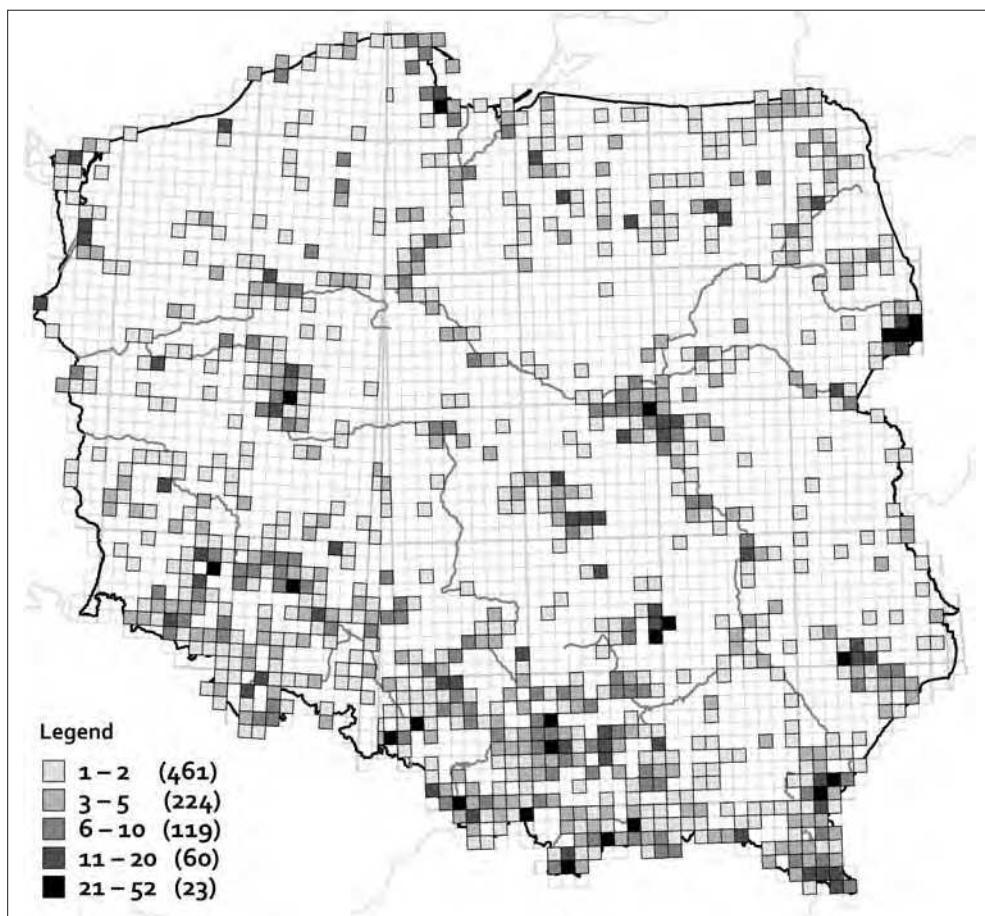


Fig. 95. Total number of species in the analyzed material per UTM square. Number of UTM squares for each class given in parentheses.

would be to compare changes in the known extent of a species over time. Therefore the relationship illustrated in Fig. 98A–C, as well as most of the presented species statistics, should be treated as a snapshot of the process of accumulation of knowledge on the species distribution.

At the family level (Table 3) the highest coverage and number of publications was found for Melandryidae (298 UTM squares, 192 publications), the lowest for Prostomidae (5 UTM squares, 13 papers).

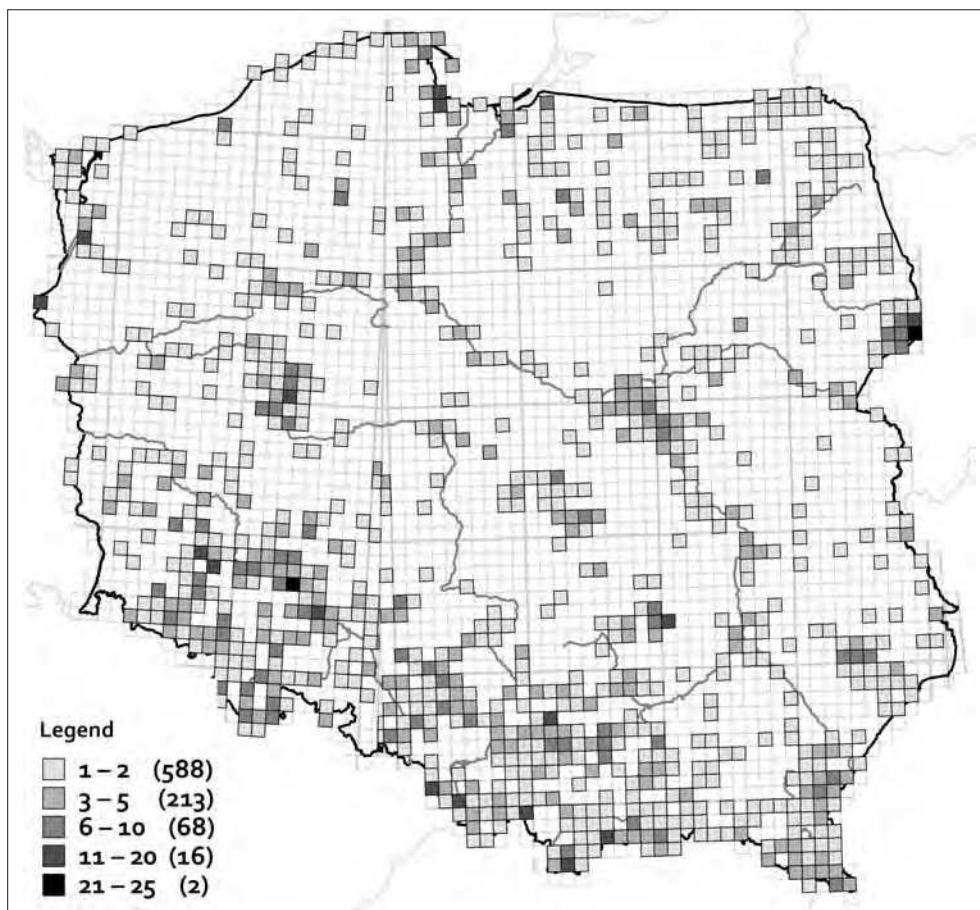


Fig. 96. Intensity of research on tenebrionids in Poland expressed as number of publications per UTM square. Number of UTM squares for each class given in parentheses.

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Table 3. Summary of data at the family level.

Column headers: UTM – number of UTM 10×10 km squares, p. first yr – first year of publication, p. last yr – last year of publication, refs – references count, r. first yr – year of the first record, r. last yr – year of the last record.

Family	UTM	p. first yr	p. last yr	refs	r. first yr	r. last yr
Aderidae	116	1854	2013	74	1854	2013
Melandryidae	298	1798	2013	192	1790	2011
Oedemeridae	185	1833	2013	165	1832	2011
Prostomidae	5	1922	2003	13	1920	1999
Pythidae	24	1794	2013	51	1790	2013
Ripiphoridae	21	1830	2007	38	1780	2006
Scaptiidae	187	1856	2013	136	1830	2011
Tetratomidae	126	1856	2013	73	1855	2011

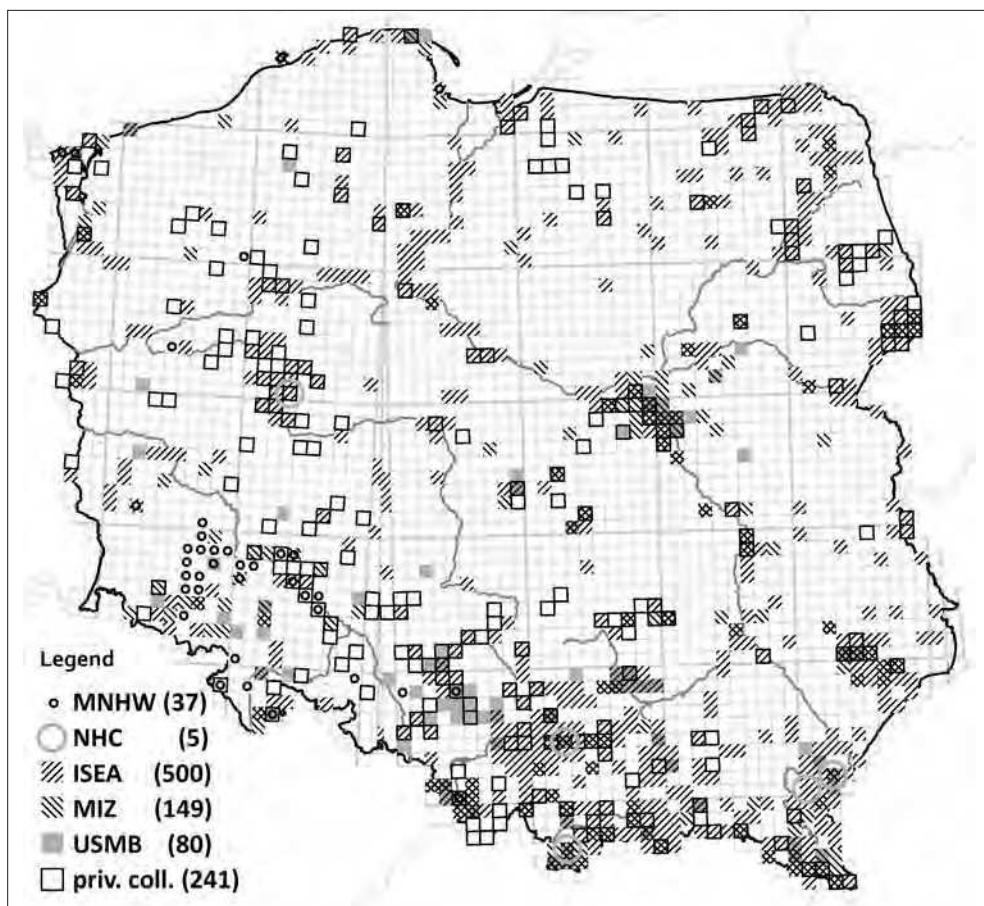


Fig. 97. Distribution of original collection localities of tenebrionid specimens from entomological collections used. Number of UTM squares for each class given in parentheses.

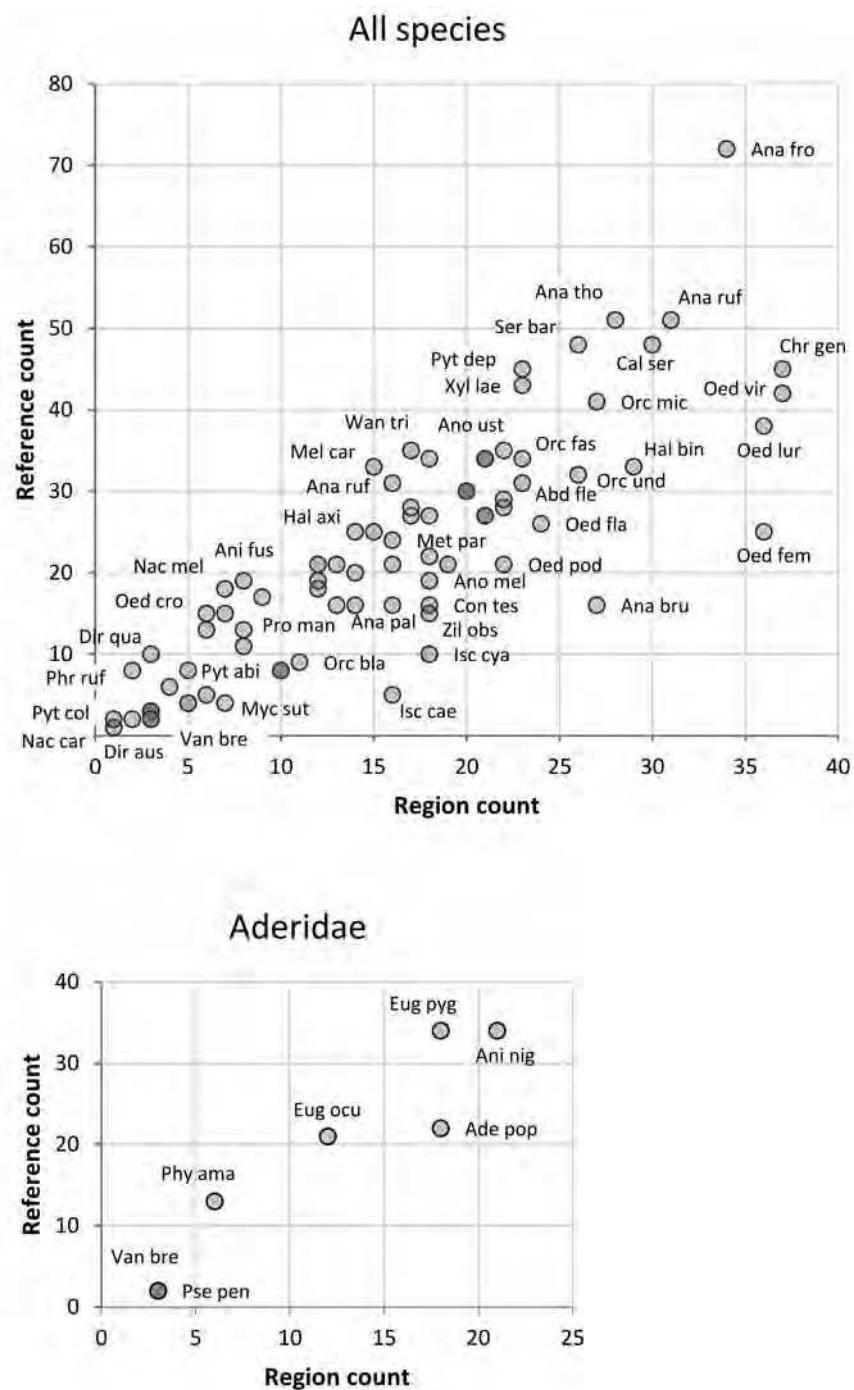


Fig. 98a. Relationship between publication count and extent of distribution in Poland, measured by number of regions of occurrence.

GENERAL DISTRIBUTION

Maps presented in this part have been made just as a supplementary material showing a general outline of distributions of the species in Palaearctic. They are not intended to present the accurate shape of ranges; in fact there is no data allowing to precisely delimit the geographical range of any of the species. As biogeography of species is not the main subject of this catalogue, we have used a simplified solution and depicted only countries or regions. In most cases, it is sufficient for presentation of the extent of the species distributions. Visualization of occurrence countries instead of giving a simple text list helps also to show the possible gaps in data, which is evident in case of cosmopolitan species.

For subspecific taxa, the maps include Palaearctic ranges of remaining subspecies. This additional information may be helpful when considering current occurrence data and chances for range changes.

The maps for *Abdera biflexuosa*, *A. quadrifasciata*, *Anaspis pulicaria*, *A. fasciata*, *A. silvatica*, *Cobosia pruinosa pruinosa*, *Ditylus laevis laevis*, *Mycterus curculiooides*, *Oedemera tristis*, *Phloiotrya subtilis*, and *Zilora ferruginea* show corrections to the information in CPC (LÖBL and SMETANA 2008), marking the lack of reliable published data about presence of these taxa in Poland on the date of publication. For *Dircea australis*, *Nacerdes carniolica carniolica*, *Orchesia micans*, *Prostomis mandibularis*, *Pseudanidorus pentatomus*, and *Vanonus brevicornis brevicornis*, we marked presence of the species, correcting the omission in CPC. Non-native parts of distribution of species, noted in CPC as ‘invasive’, were given a separate symbol.

light gray	□	– land borders
gray	■	– Palaearctic borders
black	■	– distribution of a main taxon
black dots	■■■■	– non-native distribution of a main taxon
black crosses (for Poland)	■■■■	– presence in Poland confirmed
white crosses on black (for Poland)	■■■■	– presence in Poland doubtful
hatches		– distribution of other Palaearctic subspecies



Fig. 99. Palaearctic distribution of *Abdera affinis*.



Fig. 100. Palaearctic distribution of *Abdera biflexuosa*.

General distribution



Fig. 101. Palaeartic distribution of *Abdera flexuosa*.



Fig. 102. Palaeartic distribution of *Abdera quadrifasciata*.



Fig. 103. Palaearctic distribution of *Aderus populneus*.



Fig. 104. Palaearctic distribution of *Anaspis arctica*.

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INDEX OF GEOGRAPHICAL NAMES

Names are followed by UTM coordinates given in brackets. Parentheses are used where there is an alternative name. Former German names of places are given in italics.

- Agatówka [EB 73]
- Albendorf* (Wambierzyce) [XR09]
- Aleksandrów [FA 39]
- Allmoyen* (Jelmuń) [EE06]
- Althammer* (Stara Kuźnia) [CA17]
- Althöfchen* (Stary Dworek) [WU22]
- Alwernia [CA94]
- Andreashütte* (Zawadzkie) [CB20]
- Andrzejówka [DV86]
- Annaberg* (Góra Św. Anny, mt.) [BA99]
- Antałówka [DV26]
- Arkadia [DC37]
- Augustów [FE36]
- Auras* (Uraz) [XS27]

- Babia Góra, mt. [CV99]
- Babice [FA01]
- Bachanowo [FF11]
- Bachorzewo [CD83]
- Bachotek [CE91]
- Bad Landeck* (Łądek-Zdrój) [XR37]
- Bagno Ławki [FE00]
- Bakończyce [FA21]
- Baligród [EV96]
- Bałdowo [CE59]
- Bandrów [FV27]
- Barania Góra, mt. [CV59]

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- Baranowo [XU21]
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Bardo (*Wartha*) [XR29]
Barschau bei Glogau (Barszów) [WT80]
Barszów (*Barschau bei Glogau*) [WT80]
Bartenstein (Bartoszyce) [DF81]
Bartne [EV29]
Bartnia Góra [EV19]
Bartoszyce (*Bartenstein*) [DF81]
Barwinek [EV57]
Bellinchen (Bielinek) [VU46]
Bellinchen a. d. Oder (Bielinek) [VU46]
Bellinchen/Oder (Bielinek) [VU46]
Bereźki [FV24]
Beskid Wołosacki, mt. [FV23]
Beuthen O.S. (Bytom) [CA57]
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Bezednia [FB41]
Będkowice [DA15]
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Biała [XU08]
„Biała Góra” nat. res. [DA28]
„Biała Góra” nat. res. [FA79]
Biała Góra, mt. [DV57]
Biała Góra [CE67]
Biała Góra (*Weissenberg*) [CE67]
Białe Skaly, mt. [DV57]
Białyka [DA00]
Białyka valley [DV35]
Białopole [FB95]
Białowieża [FD94]
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„Bielinek” nat. res. [VU46]

- Bielszowice [CA47]
Bircza [FA00]
Birkenau (Gliwice-Brzezinka) [CA37]
Birnbäumel (Gruszczka) [XT50]
Bischoffskoppe (Biskupia Kopa, mt.) [XR76]
Biskupia Kopa, mt. (*Bischoffskoppe*) [XR76]
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