

Some results of Study on the Nitidulidae from Namibia and Adjacent Territories. Part 1 (Coleoptera, Cucujoidea, Nitidulidae)¹

ALEXANDER G. KIREJTSHUK

With 110 figures in the text

Abstract. The paper deals with a preliminary consideration of the Namibian fauna of the family Nitidulidae, including some species recorded from adjacent territories, i.e. Angola, Zambia, Botswana, Zimbabwe and Republic of South Africa. The data on 56 species are given in the paper, amongst which 31 species have been collected from Namibia, but others can be expected to be collected there. The synonymy and systematic position for some species of the genera *Carpophilus* STEPHENS, 1830, *Meligethes* STEPHENS, 1830, *Lordites* ERICHSON, 1843, are discussed. 7 species of the *Meligethes* subgenus *Clypeogethes* are described as new: *M. (C.) annae*, locus typicus: Republic of South Africa, Natal, Cathedral Peaks Forest Station; *M. (C.) antila*, locus typicus: RSA, Natal, Nqutu; *M. (C.) arcopenis*, l.t.: Namibia, Grootfontein, Farm Hurisib; *M. (C.) asignifer*, l.t.: Namibia, Bushmanland, Klein Dobe; *M. (C.) bisignifer*, l.t.: Namibia, Abachaus; *M. (C.) mitis*, l. t.: Namibia, env. Windhoek; *M. (C.) opacitorsurm*, l.t.: Namibia, Okahandja. Information on distribution of the considered species is given in a maximally wide scope. The notes to some species are provided available bionomical comments.

Key words: Insecta, Coleoptera, Nitidulidae; new species, lectotype designation, new synonymy, key, faunistics; Afrotropical region, Southern Africa, Namibia, Angola, Botswana, Zambia, Zimbabwe, and Republic of South Africa.

1. Introduction

This paper was planed as a result of study on the specimens collected during field works in Namibia of M. UHLIG and other participants of the expeditions of the Museum of Natural History of the Humboldt-University (Berlin) and of H. ROER from the Museum Alexander Koenig (Bonn). Besides, other data of study of the specimens from Namibia and adjacent territories deposited in collections of various museums are included in the paper, except materials published in some of my previous works (KIREJTSHUK, 1987; 1988; 1990; 1994; 1995). In order to make the text more short the writer have omitted the synonymy not connected with data from Africa and many records for the species under consideration from this continent but from localities far from Namibia are listed with very short indications of country and museum depository in abbreviations mentioned in the text.

¹ Ergebnisse der entomologischen Afrika-Expeditionen des Museums für Naturkunde Berlin. 21. Beitrag.
Results of the entomological expeditions of the Museum of Natural History Berlin to Africa. 21st contribution.

2. Abbreviations

CAS	– Californian Academy of Sciences, Los Angeles;
CMO	– Canadian Museum of Nature, Ottawa;
CNC	– Canadian National Collection (Biosystematics Research Institute), Ottawa;
DEI	– Deutsches Entomologisches Institut, Eberswalde;
IZW	– Instytut Zoologiczny PAN, Warszawa;
MNHUB	– Museum für Naturkunde der Humboldt-Universität, Berlin;
MAK	– Zoologisches Forschungsinstitut und Museum Alexander Koenig, Bonn;
MNP	– Museum National d’Histoire Naturelle, Paris;
MAT	– Musée Royal de l’Afrique Centrale, Tervuren;
NHL	– Natural History Museum in London (formerly – British Museum (Natural History));
NMW	– Naturhistorisches Museum in Wien;
NRS	– Naturhistoriska Riksmuseet, Stockholm;
RNL	– Rijkmuseum van Natuurlijke Historie, Leiden;
SMS	– Staatliches Museum für Naturkunde in Stuttgart;
SMD	– Staatliches Museum für Tierkunde, Dresden;
SMWN	– State Museum of Namibia, Windhoek;
TMB	– Termesztudományi Múzeum, Budapest;
TMP	– Transvaal Museum, Pretoria;
ZIN	– Zoological Institute of the Russian Academy of Sciences, St. Petersburg;
ZMH	– Zoologiska museet, Helsingfors Universitet;
ZMK	– Zoologisk Museum, København;
ZML	– Zoological Museum at Lund University;
ZMM	– Zoological Museum at Manchester University;
ZMU	– Zoological Museum at Moscow University;
ZSM	– Zoologische Staatssammlung, München.

3. List of species recorded from Namibia and adjacent territories

Epuraeinae

- Epuraea (Haptoncus) luteola* ERICHSON, 1843
Epuraea (Haptoncus) ocularis FAIRMAIRE, 1849
Epuraea (Haptoncurina) motschulskyi REITTER, 1873
Epuraea (Polinexa) decellei JELINEK, 1979
Epuraea (Polinexa) inexpectata JELINEK, 1979
Parepuraea simoni simoni (GROUVELLE, 1895)
Taenioncus (Taenioncus) longior (GROUVELLE, 1912)

Carpophilinae

- Carpophilus (Carpophilus) bifenestratus* MURRAY, 1864
Carpophilus (Carpophilus) dolens MURRAY, 1864
Carpophilus (Carpophilus) hemipterus (LINNEUS, 1758)
Carpophilus (Carpophilus) marginellus MOTSCHULSKY, 1858
Carpophilus (Carpophilus) obsoletus ERICHSON, 1843
Carpophilus (Carpophilus) quadrisignatus ERICHSON, 1843
Carpophilus (Myothorax) dimidiatus (FABRICIUS, 1792)
Carpophilus (Myothorax) fumatus BOHEMAN, 1851
Carpophilus (Myothorax) nepos MURRAY, 1864 (= *freemani* DOBSON, 1956)
Carpophilus (Myothorax) zeaphilus DOBSON, 1969
Carpophilus (Ecnomorphus) apicipennis FAIRMAIRE, 1869 **comb. n.**
Carpophilus (Ecnomorphus) deplanatus (BOHEMAN, 1851)
Urophours (Anophorus) humeralis (FABRICIUS, 1798)
Urophours (Anophorus) picinus (BOHEMAN, 1851)

Amphicrossinae

Amphicrossus namibiensis KIREJTSHUK, 1987

Meligethinae

- Pria (Pria) angustula* COOPER, 1982
Pria (Pria) cinerascens ERICHSON, 1843
Pria (Pria) magna REITTER, 1872
Pria (Pria) notata COOPER, 1982
Pria (Pria) palpata KIREJTSHUK, 1989
Pria (Pria) robiginosa COOPER, 1982
Pria (Pria) testacea GROUVELLE, 1908 (1909)
Pria (Pria) vicina GROUVELLE, 1908/1909
Meligethes (Lariopsis) arcuatus REITTER, 1872
Meligethes (Lariopsis) haagii REITTER, 1872
Meligethes (Lariopsis) pulchellus REITTER, 1872
Meligethes (Lariopsis) variabilis REITTER, 1872
Meligethes (Chromogethes) illustris GROUVELLE, 1899
Meligethes (Clypeogethes) annae **sp. n.**
Meligethes (Clypeogethes) antlia **sp. n.**
Meligethes (Clypeogethes) arcopenis **sp. n.**
Meligethes (Clypeogethes) asignifer **sp. n.**
Meligethes (Clypeogethes) bisignifer **sp. n.**
Meligethes (Clypeogethes) imitans KIREJTSHUK, 1988
Meligethes (Clypeogethes) rugifer SPORNRAFT et KIREJTSHUK, 1993
Meligethes (Clypeogethes) mitis **sp. n.**
Meligethes (Clypeogethes) opacidorsum **sp. n.**
Meligethes (Clypeogethes) verniceus KIREJTSHUK, 1990

Nitidulinae

- Annachramus distinctus* (GROUVELLE, 1899)
Taracta varia (GROUVELLE, 1898)
Stelidota didyma REITTER, 1875
Lordites (Lordites) costipennis (BOHEMAN, 1851), **comb. n.**
Lordites (Lordites) tibialis (BOHEMAN, 1851), **comb. n.**
Aethina (Circopes) peringueyi GROUVELLE, 1908/1909, **comb. n.**
Aethina (Ithya) hirsutula REITTER, 1873, **comb. n.**
Anister hintoni JELINEK, 1981b
Monafricus major (GROUVELLE, 1899)

4. List of the studied specimens and descriptions of species from Namibia and adjacent territories

Epuraeinae

Epuraea (Haptoncus) luteola ERICHSON, 1843

GILLOGLY, 1982: tropicopolitan; AUDISIO, 1982: Sierra Leone; KIREJTSHUK, 1992 and others: pantropical species, recorded from Japan, Northern Korea and China (synonymy); AUDISIO, 1993: also Italy and Macaronesia.

Specimens examined: some hundreds of specimens from different African countries, including Angola (MAT, ZIN), Guinea (TMB, ZIN), Ivory Coast (SMS, ZIN), Togo (SMS), Nigeria (SMS, ZIN), Cameroon (MNHUB, ZIN), Equatorial Guinea (MNHUB, ZIN), Fernando-

Poo(MNHUB, ZIN), Ethiopie (TMB, ZIN, ZMM), Tanzania (MNHUB), Kenya (MNHUB, TMB, ZIN), Malawi (MAT, ZIN), Madagascar (MAT, MNHUB, ZIN), Réunion (MNHUB), Seychelles (ZIN), Uganda (CMC).

Notes. This species is a mycetophagous form with a wide scope of trophics, adhered to different substracts of dead plants; imagines of it are not infrequent visitors of flowers.

***Eपुरaea (Haptoncus) ocularis* FAIRMAIRE, 1849 (figs. 1–3)**

(= *Eपुरaea bisignata* BOHEMAN, 1851; *Eपुरaea bifasciata* KRAATZ, 1895).

BOHEMAN, 1851: South Africa; KRAATZ, 1895: Togo; GILLOGLY, 1982: E Africa, Madagascar, Sri Lanka, Indonesia, Micronesia, Hawaii; KIREJTSHUK, 1992: also Korea, central and southern China, Australia and other tropical territories of Eastern Hemisphere.

Specimens examined: Namibia: 1 (MNHUB) – “East Caprivi: Katima Mulino, lux, 19°29'S/24°17'E, 3–8. III. 1992, leg. M. Uhlig”; Republic of South Africa: Holotype *Eपुरaea bisignata* Boh., female (NRS) – “Caffraria, J. Wahlb.,” “typus”, “*Eपुरaea bisignata* Boh. Lectotypus 1851–1968, Dr. Endrödy-Younga”; 1 female (SMS) – “Natal, St. Lucia, 29–31. 10. 1981, leg. J. Klapperich”; and also some dozens of specimens from different African countries, including Angola (MAT, ZIN), Guinea (Budapest, ZIN), Togo (MNHUB, ZIN), Cameroon (MNHUB, ZMK), Zaire (MAT, ZIN), Ethiopie (ZIN, ZMN), Kenya (MNHUB, TMB, ZIN), Tanzania (MNHUB, ZIN), Malawi (MAT, ZIN), Seychelles (MAT, ZIN).

Notes: This species has a mode of life similar to that of previous species.

The type specimen of *E. bisignata* and another specimen from Republic of South Africa have a comparatively strongly pigmented (dark pitchy brown to almost black) body, with lighter (bright reddish) fore part of head, mandibles, antennae, legs, explanate sides of pronotum and elytra and a spot on disk or also with base on elytra (underside has an intermediate coloration – chestnut brownish). A similar aberrant coloration is unusual for most specimens from different parts of areal, though this extreme in coloration occurs among the specimens originated from southern islands of the Polynesian region (but all specimens from the Australian region have a more or less normal colour). Therefore the writer prefer after JELINEK (1979) to regard this form as a kind of populational variety.

***Eपुरaea (Haptoncurina) motschulskyi* REITTER, 1873**

JELINEK, 1979: Guinea, Sierra Leone, Nigeria, Zaire, Rwanda, Burundi, Tanzania, Zambia, Republic of South Africa, Madagascar; JELINEK, 1978: Bhutan, Nepal, China, Japan; KIREJTSHUK, 1992: Tropics of Asia, Africa, Madagascar, Australia and Pacific insular systems.

Specimens examined: Namibia: 3 exx (MAK); Republic of South Africa: 8 (RNS) – “Durban, Mjöberg, Aug.,” “det Sjöberg dispersus Grouv.”; and also some hundreds specimens from Cameroon (MAT, STD, ZIN), Togo (MNHUB, ZIN), Nigeria (SMS), Sudan (TMB, ZIN), Ethiopie (MAT, TMB, ZIN), Uganda (MAT, ZIN), Kenya (TMB, ZIN), Rwanda (MAT), Tanzania (DEI, MAT, MNHUB, ZIN), Mali (MAT).

Notes. This species is associated with flowers and fruits of different plants (AUDISIO, 1982; JELINEK, 1992).

***Eपुरaea (Polinexa) decellei* JELINEK, 1979**

(= *Eपुरaea (Eपुरaea) decellei* JELINEK, 1979; *Eपुरaea (Polinexa) decellei*: KIREJTSHUK, 1989).

JELINEK, 1979: Cameroon, Zaire, Uganda, Tanzania, Zimbabwe.

Specimens examined: Angola: 2 (NHL, ZIN) – “(A 36) Chianga, 21–24. III. 1972”, “at light”; and also some specimens from Togo (DEI, ZIN), Cameroon (MNHUB, ZIN), Ghana (CNC, ZIN), Gambia (ZIN, ZML), Senegal (ZIN, ZML), Tanzania (TMB, ZIN).

Notes. One of studied male from Angola has abnormally convex pronotum with more arched and comparatively widely explanate side, though other characters, including those in genitalia, are as those in most specimens of the species under consideration.

This species in contrast to other species of this subgenus has an appearance similar to that in some anthophagous representatives from the subgenus *Haptoncurina* JELINEK, 1979, genus *Pareपुरaea* JELINEK, 1979 and tribe Taenioncini KIREJTSHUK, in press.

***Epuraea (Polinexa) inexpectata* JELINEK, 1979**

(= *Epuraea (Epuraea) inexpectata* JELINEK, 1979; *Epuraea (Polinexa) decellei*: KIREITSHUK, 1989).

JELINEK, 1979: Nigeria, Zaire.

Specimens examined: Angola: 1 (NHL) – "(A 36) Chianga, 21–34. III. 1972", "at light"; some specimens also from Zaire (MAT, ZIN).

Notes. This species is characterized by oval and convex body having some resemblance to anthophagous species of *Meligethinus* GROUVELLE, 1906, *Pria* STEPHENS, 1830, and *Apria* GROUVELLE, 1919, and it could be supposed that these species share a similar mode of life.

***Parepuraea simoni simoni* (GROUVELLE, 1895)**

(= *Epuraea simoni* GROUVELLE, 1895; *Parepuraea simoni simoni*: JELINEK, 1979)

JELINEK, 1979: "Rhodesia, South Africa".

Specimens examined: Angola: 10 (NHL, ZIN) – "(A 30) 7 mls W Gabela, 16–18. III. 1972, at light"; 1 (ZIN) – "(A 26) Salazar, I.I.A.A., 9–15. III. 1972, at light"; Zambia: 1 (IZW) – "N. Rhodesia, Abercorn, XI. 1946, Dr. W. Eichler"; 1 (ZIN) – "N. Rhodesia, XI. 1946, Dr. W. Eichler".

Notes. This species is quite widely distributed ranging from West and Central Africa (Liberia, Cameroon, Ivory Coast, Zaire, and additional specimens from Togo (MNHUB, ZIN) and Nigeria (CMC, ZIN, ZMM) – *P. simoni minor* JELINEK, 1979), through Equatorial province of Zaire, including additional material from the Upper Zaire (11 (FMC, ZIN) – "B. Congo, Bambesa, Oriente Prov., Bas-Uele Distr., 20-VI-1958", "Purchased ex Ch. de Wyngaert (Brussels)"), (*P. s. major* JELINEK, 1979), to Ethiopia, East Africa, Rwanda and southern Katanga (*P. s. intermedia* JELINEK, 1979), reaching Zimbabwe and Republic of South Africa (*P. s. simoni*). The information taken from labels of museum specimens belonging to different species of the *Parepuraea* JELINEK, 1979, allows a supposition on association of this group with flowers (at least, imagines and larvae of one of the species from Madagascar are collected on inflorescence of a representative from the Araceae).

***Taenioncus (Taenioncus) longior* (GROUVELLE, 1912)**

(= *Carpophilus longior* GROUVELLE, 1912; *Taenioncus longior*: KIREITSHUK, 1984).

WILLIAMS & al., 1983: Republic of the Congo; KIREITSHUK, 1984: Zaire, Ethiopia.

Specimens examined: Namibia: 1 ex (MAK) – "Nyangana/Okavango, 14–22. 1. 1985, H. Roer"; and also more than two hundreds specimens from Republic of South Africa (SMS, ZIN), Guinea (ZIN), Ivory Coast (SMS, ZIN), Ghana (SMS), Zaire (MAT, ZIN), Gambia (ZML, ZIN), Ethiopia (MAT, ZIN), Tanzania (MNHUB, ZIN).

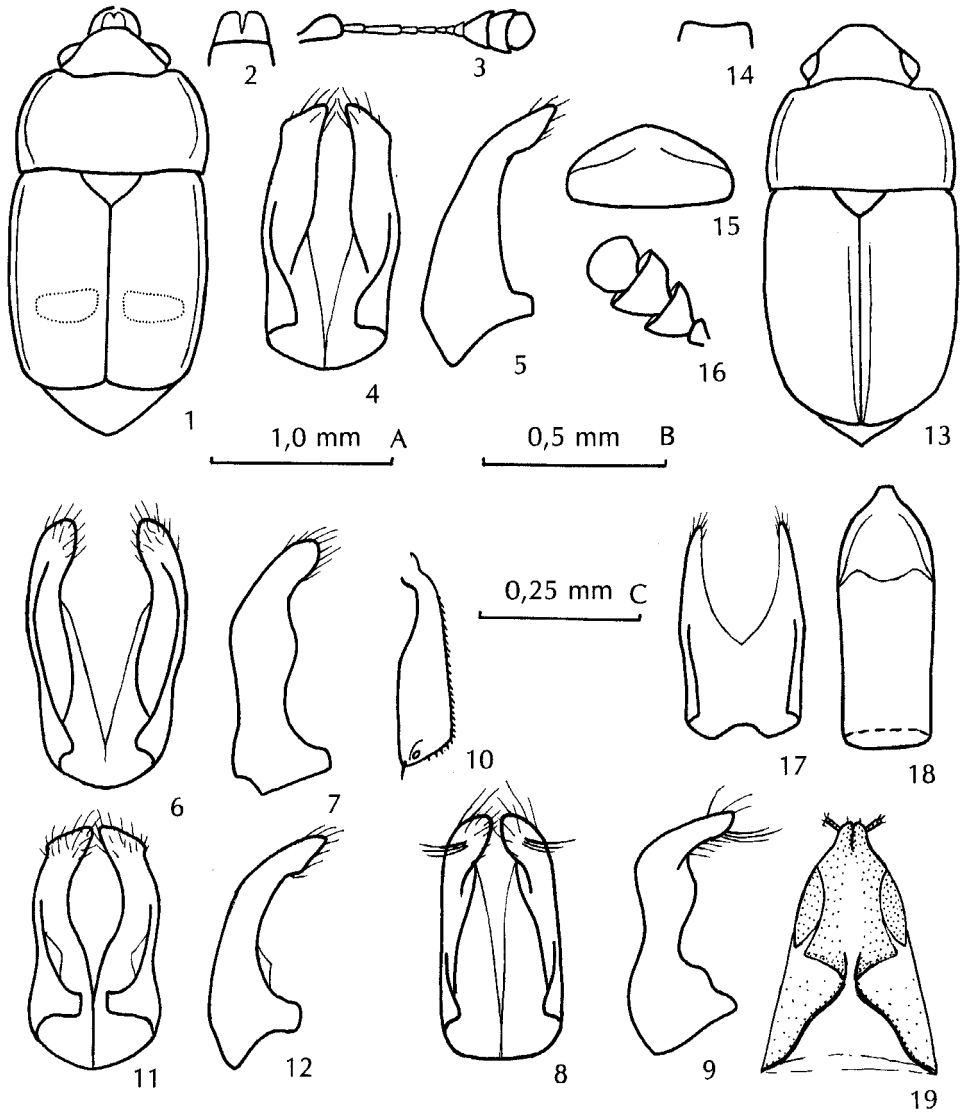
Notes. Bionomy of this species remains still unknown. However, the indo-malayan members of the subgenus are associated with flowers and soft fruits (probably, with larval development in fructifications at decaying stage).

Carpophilinae***Carpophilus (Carpophilus) bifenestratus* MURRAY, 1864 – fig. 4–5**

(= *Carpophilus tersus* WOLLASTON, 1865 (KIREITSHUK, in litteris); ? *Carpophilus bisignatus* BOHEMAN, 1851).

GROUVELLE, 1908: India, West Bengal, "Mahe"; Sri Lanka; WILLIAMS & al., 1983: also Malgasy Republic, Canary islands; AUDISIO et KIREITSHUK, 1988: also Afrotropical region, Mediterranean, including Kazakhstan; KIREITSHUK, in litteris: Thailand, Vietnam.

Specimens examined: Namibia: 3 (MNHUB, ZIN) – "b. Grootfontain, Farm Hurisib, 8–9. X. 1991, leg. U. Göllner", "Oleander"; Republic of South Africa: Holotype *C. bisignatus*, female (NRS) – "Caffraria, J. Wahlb.", "type", "Lectotypus 1851–1968, Dr. S. Endrödy-Younga"; 1 (NRS) – "Cap B. Spei", "Drege"; and also more than a hundred specimens from Guinea (ZIN), Ghana (MAT, ZIN), Nigeria (TMB, ZSM), Equatorial Guinea (MNHUB, ZIN), Sudan (TMB), Ethiopia (ZMM), Uganda (CNC, ZMH, ZIN), Tanzania (DEI, TMB, ZIN), Madagascar (DEI, MNHUB, ZIN).



Figs. 1–19. Genera *Epuraea*, *Carpophilus* and *Pria* (orig.). – 1–3 *E. (Haptoncus) ocellaris* (holotype *E. bisignata* syn. n.); 1 body shape with outline of explanate sides of pronotum and elytra and dotted contour of reddish spot on elytral disk, dorsal; 2 fore part of head with labrum, dorsal; 3 antenna; 4–5 *C. (Carpophilus) bifenestratus*; 4 tegmen, ventral; 5 idem., lateral; 6–7 *C. (Carpophilus) binotatus* MURRAY, 1864; 6 tegmen, ventral; 7 idem., lateral; 8–9 *C. (Carpophilus) dolens*; 8 tegmen, ventral; 9 idem., lateral; 10–12 *C. (Myothorax) truncatus*; 10 male hind tibia; 11 tegmen, ventral; 12 idem., lateral; 13–19 *P. (Pria) angustula*; 13 body with contour of explanate pronotal sides and subsutural lines, dorsal; 14 fore edge of frons; 15 mentum; 16 male antennal club; 17 tegmen, ventral; 18 penis trunk, dorsal; 19 ovipositor, ventral; Scales: A – to figs. 1, 13; B – to figs. 2, 3, 14; C – to figs. 4–12, 15–19.

Notes. This species is quite common in Afro-Madagascarean regions and manifests a considerable variability maintaining a more or less stable structure in male genitalia (see KIREJTSHUK, 1995). Unfortunately, the type *C. bisignatus* is scarcely distinguishable from females of both *C. (C.) bifenestratus* and other species of this group, though it is more similar to the first by a comparatively slender body (see KIREJTSHUK, 1994). This species differs from *C. (C.) binotatus* MURRAY, 1864 with a more restricted afro-tropical area mainly in genital structure of male (see KIREJTSHUK, 1995 and figs. 6–7), comparatively more robust and more convex body.

***Carpophilus (Carpophilus) dolens* MURRAY, 1864 – figs. 8–9**

WILLIAMS & al., 1983: Senegal, Equatorial Guinea.

Specimens examined: Zambia: 1 (IZW) – “N. Rhodesia, 3. I. 1947, Dr. W. Eichler”; Equatorial Guinea (MNHUB) – “Span. Guinea, Makomo, Campogt, 6. IV. 06, G. Tessmann S.G.” (det. S. Endrödy-Younga), Uganda (CMC, ZIN), Tanzania (NRS).

Notes. This species is rather similar to *C. (C.) obsoletus*, but differs from it in somewhat more oval body, light antennae and legs, widely rounded pygidial apex in males and structure of aedeagus. This name is a probable senior synonym of *C. tumidulus* GROUVELLE, 1899, however, their synonymy should be defined after a comparison of the types.

***Carpophilus (Carpophilus) hemipterus* (LINNEUS, 1758)**

KIREJTSHUK, 1992; Audisio, 1993 and others: cosmopolitan.

Specimens examined: Republic of South Africa: 1 (MNHUB) – “gymnura N., Pr. b. sp.”; 1 (MNHUB) – “N. Transvaal Lootpansberg; Mp. hôte Magd. Knothe S., V. 1901”; 3 (TMB, ZIN) – “Transvaal, Pretoria, singled, 24. XII. 1980, leg. S. Endrödy”; and also some hundreds specimens from Republic of South Africa (CMO, NRS, TMB, TMP, ZIN), Senegal (MAT, ZIN) Gambia (TMB, ZIN, ZML), Cameroon (MNHUB), Equatorial Guinea (MNHUB), Kongo (TMB, ZIN), Zaire (MAT, TMB, ZIN), Sudan (TMB), Ethiopia (MNHUB, TMB, ZIN, ZMM), Uganda (ZIN, ZMK), Kenya (MNHUB, TMB, ZIN, ZMK), Rwanda (MAT, ZIN), Burundi (MAT, ZIN), Tanzania (MNHUB), Madagascar (ZIN), Seychelles (ZIN).

Notes. This species is connected with decomposed plant substrates of different consistence in natural and artificial conditions.

***Carpophilus (Carpophilus) marginellus* MOTSCHULSKY, 1858**

KIREJTSHUK, 1992: cosmopolitan.

Specimens examined: Republic of South Africa: 3 (STD, ZIN) – “Kapland”; and some specimens from Guinea (ZIN), Ghana (MAT, ZIN), Seychelles (ZIN).

This species is well characterized by unicoloured chestnut body with subparallelsided pronotum in distal half, rectilinear caudal marginal lines behind mid coxal cavities and peculiar structure of aedeagus. It is very widely distributed (perhaps, mainly due to delivery of man) occurring in natural conditions under bark, in fruits and other substrates of plant origin. However, this species is one of usual inhabitants in different places with stored products.

***Carpophilus (Carpophilus) obsoletus* ERICHSON, 1843**

(= *Carpophilus immaculatus* LUCAS, 1849).

GILLOGLY, 1962: United States, West Indies, South America, Egypt, West Africa, Madagascar, Sri Lanka, Malaya, Philippines, China, Japan, W Caroline Is.; JELINEK, 1981a: Oriental and Ethiopian regions, Iran; WILLIAMS & al., 1983: in addition to the mentioned above – Africa, New Guinea; KIREJTSHUK, 1992: in natural conditions in tropics of the East Hemisphere.

Specimens examined: Namibia: 3 (MNHUB, ZIN) – “Bushmanland: Kleine Dobe, 19°25'S/20°21'E, 19–21. II. 92, leg. M. Uhlig”; and also some hundreds specimens from Ghana (MAT, ZIN), Guinea (ZIN), Equatorial Guinea (MNHUB, ZIN), Gambia (ZIN, ZML), Ethiopia (ZIN), Tanzania (MNHUB, TMB, ZIN, ZSM) and so on.

Notes. This species is easily distinguishable due to its deep black body with rather dark appendages (antennae and legs), acute or subacute pygidial apex in the both sexes and peculiar structure of genitalia. It is connected with decomposed plant substrates of different consistence.

Carpophilus (Carpophilus) quadrisignatus ERICHSON, 1843

WILLIAMS et al., 1983: Europe, Algeria; AUDISIO, 1993: also Macaronesia, Caucasus, Iran, Arabian peninsula.

Specimens examined: Namibia: 11 (MNHUB, ZIN) – “Bushmanland, Klein Dobe, Fruchtköder, 19°25'S/20°21'E, 20–21. II. 92, leg. M. Uhlig”; some dozens specimens from Equatorial Guinea (MNHUB, ZIN), Kenya (TMB, ZIN).

Notes. This species is closely related *C. (C.) hemipterus* and, perhaps, lives in the same localities and conditions (at least in the East Hemisphere). Nevertheless, the both species can be easily distinguished according to keys of S. HISAMATSU (1963), K. SPORNRAFT (1967) and P. AUDISIO (1993). The males of the species under consideration have a characteristic very wide hypopygidium with a pair of rather large and well raised depression occupying the most part of sclerite. Besides, both species have peculiar structure of aedeagus.

Carpophilus (Myothorax) dimidiatus (FABRICIUS, (1792)

(= *Carpophilus pilosellus* MOTSCHULSKY, 1858, non auctorum).

WILLIAMS & al., 1983; KIREJTSHUK, 1992 and others: cosmopolitous.

Specimens examined: Namibia: 8 (MNHUB, ZIN, SMWN) – “Bushmanland, Klein Dobe, 19°25'S/20°21'E, 20–21. II. 92, leg. M. Uhlig”; 6 (MNHUB) – “E. Caprivi: Katimo Mulilo, 17°29'S/24°17'E, 3–8. III. 92, lux, leg. U. Göllner”; Republic of South Africa: 1 (STD) – “Kapland”; and also some hundreds from Ivory Coast (MAT, ZIN), Cameroon (STD, ZIN), Kenya (ZIN), Ethiopie (ZIN, ZMM), Zaire (DEI), Madagascar (DEI, MNHUB, ZIN) and so on.

Notes. This species is rather small and dark with peculiar structure of genitalia in the both sexes (with blunt apex of generalized structure of ovipositor). It has hind femora in males partly similar to those in *C. (M.) truncatus*, but with gentle curvation of inner edge. The species under consideration as other members of the subgenus has a wide range of possible food from different plant substrates, frequently recorded from grains and corn and other plant stored products.

Carpophilus (Myothorax) fumatus BOHEMAN, 1851

(= *Carpophilus (Myothorax) ochropterus* KLUG, 1862).

AUDISIO, 1982: Sierra Leone; WILLIAMS & al., 1983: Africa, USA (Florida), West Indies, Malagasy Republic; DELOBEL, TRAN, 1993: Antilles, Africa.

Specimens examined: Namibia: 1 (MNHUB) – “E. Caprivi: Katimo Mulilo, 17°29'S/24°17'E, 3–8. III. 92, lux, leg. M. Uhlig”; 1 (MNHUB) – “East Caprivi: Mudumu NP, 18°10'S/23°26'E, 8–13. III. 92, lux, leg. M. Uhlig”; 2 (MAK) (light specimens) – “Bunja/Okavango, 24–27. 02. 1991, leg. Roer”; Republic of South Africa: LECTOTYPE *C. fumatus*, male (NRS), here designated after designation in collection by S. Endrödi-Younga in 1968 – “Cafrazi J. Wahlb.”; 1 Paralectotype, male (NRS) – “Cap. B. Spei. J. Wahlb.”; 2 (ZSM) – “Transvaal, Umg. Pretoria, VII. 1953, A. v. Peez”; 3 (ZSM) – “Natal, Umtentweni River, July 1955 (VI. 1952), A. L. Capener”; 1 (ZSM) – “Swaziland, Franchi, 15–31. 12. 1954, A. L. Capener”; and also some thousands specimens from Republic of South Africa (TMB, ZIN), Sudan (TMB, ZIN), Togo (SMS), Liberia (SMS), Ghana (CMC, MAT, SMS, ZIN), Gabon (MAT, NRS, SMD, ZIN), Cameroon (SMS, TMB, ZIN), Nigeria (SMS), Ivory Coast (SMS), Republic of Central Africa (MAT, ZIN), Congo (MAT, ZIN), Zaire (MAT, ZIN), Ethiopie (MAT, ZIN, ZMM), Tanzania (ZSM), Uganda (CMC, ZMK), Rwanda (MAT, ZIN), Mali (MAT, ZIN), Madagascar (ZIN, ZSM), Réunion (MAT).

Notes. This species is recorded from dry fruits, seeds, grains, corn and other plant substrates.

Dark specimens correspondent to the type *C. fumatus* are quite rare, but light ones correspondent to *C. ochropterus* are more abundant in all parts of the African area of the species under consideration which is also collected in India (KIREJTSHUK, in litteris). Nevertheless, *C. (M.) mutilatus* ERICHSON, 1843 is,

perhaps, more usual in subtropics and tropics eastwards from Africa (Asia, the island systems of Indian and Pacific oceans). In particular, *C. (M.) mutilatus* is collected together with *C. (M.) fumatus* and *C. (M.) truncatus* on Seychelles (ZIN).

***Carpophilus (Myothorax) nepos* MURRAY, 1864**

(= *freemani* DOBSON, 1956: KIREJTSHUK, in litteris).

KIREJTSHUK, 1992: Jordan, Morocco, Guinea, Canary and Mascarene Islands, South and North America; DELOBEL, TRAN, 1993: West Africa, Brazil, Australia; AUDISIO, 1993: also Mediterranean.

Specimens examined: 4 (MAT, ZIN) – “Dundo, 7.22 S 20.50 E, Forêt claire, detr., sol., 10. XII. 48, A. B. MACHADO!

Notes. This species is quite probable in the region under consideration being associated with dry fruits, seeds, grains, corn and other plant substrates. However, it is also expectable that some of the records on this species as *C. (M.) freemani* should be regarded as doubtful.

***Carpophilus (Myothorax) truncatus* MURRAY, 1864 (figs. 10–12)**

(= *Carpophilus pilosellus auctororum* (AUDISIO, 1982), non MOTSCHULSKY, 1858).

MURRAY, 1864: Madagascar.

Specimens examined: Republic of South Africa: 2 (ZSM) – “Natal, Durban, 27 August 1980, leg. Spornraft” (det. as *C. pilosellus*); Madagascar: 5 Syntypes (MNHUB) – “Madagascar, Goudot, 8371”; 2 ? SYNTYPES (ZIN) – “Madagascar”; and also some specimens from Seychelles (ZIN) and Morocco (ZSM).

Notes. Some specimens determined by coleopterists as *C. pilosellus* actually belong to some (at least two) species, though the african form should be in many cases regarded as this species. The true *C. pilosellus* should be regarded as a junior synonym of *C. (M.) mutilatus* ERICHSON, 1843 (KIREJTSHUK, 1992). It is well characterised by dark coloration and male hind tibiae sharply dilated along inner edge. The bionomy of this species, perhaps, is similar to that in other members of the subgenus with wide distribution.

***Carpophilus (Myothorax) zeaphilus* DOBSON, 1969**

WILLIAMS et al., 1983: Kenya, Uganda; JELINEK, 1988: Yemen, Tanzania, Republic of South Africa; AUDISIO, 1993: tropical and southern Africa, Arabian peninsula, Spain, Portugal, ?Italy.

Specimens examined: Namibia: 2 (MNHUB) – “Osona bei Okahandja, p. III–m. IV. 1989, leg. J. Irish (UG)”; 1 (MAK) – “Okahandja, 1240 m, 14.3.1979, H. Roer”; also about a hundred specimens from Nigeria (ZIN), Kenya (TMB, ZIN, ZSM), Tanzania (MNHUB), Republic of South Africa: (SMD, ZIN, ZSM), Madagascar (ZIN, ZSM).

Notes. This species has probably been confused with *C. (M.) fumatus* BOHEMAN, 1851, *C. (M.) mutilatus* ERICHSON, 1843 and *C. (M.) nepos* MURRAY, 1864, because of some resemblance in both appearance and similar mode of life, but each of the mentioned species is quite distinct from others in peculiar structure of aedeagus.

***Carpophilus (Ecnomorphus) apicipennis* FAIRMAIRE, 1869, comb. n.**

AUDISIO, 1982: Sierra Leone, tropical Africa and Madagascar; WILLIAMS & al., 1983: also Ivory Coast, Republic of Equatorial Guinea, Tanzania.

Specimens examined: Namibia: 2 (MNHUB, ZIN), – “Bushmanland, Klein Dobe, 19°25'S/20°21'E, 20–21. II. 92, leg. M. Uhlig”; 1 (MNHUB) – “Kavango, Popa Falls, 18°07'S/21°35'E, 26. II–3. III. 92, leg. M. Uhlig”; and also about a hundred specimens from Gambia (ZML, ZIN), Ivory Coast (SMS), Cameroon (MNHUB, ZIN), Equatorial Guinea (MNHUB), Ethiopie (ZIN, ZMM), Tanzania (MNHUB, TMB, ZIN, ZMK), Uganda (CNC, ZIN), Republic of South Africa (CMO, CNC, ZIN, ZSM).

Notes. This species having a very flattened body with characteristic shape of pronotum and elytra and proportion of these sclerites, aedeagal structure should without doubt be regarded in composition of the subgenus *Ecnomorphus* MOTSCHULSKY, 1858 being similar to the indo-malayan *C. (E.) plagiaticipennis* (MOTSCHULSKY, 1858). It is easily distinguished from other african members of the subgenus in a

comparatively small and very shiny body with lightened spots on elytra. As other species of the subgenus it can be associated with subcortical localization in arboreal communities.

***Carpophilus (Ecnomorphus) deplanatus* (BOHEMAN, 1851)**

WILLIAMS et al., 1983: Republic of South Africa.

Specimens examined: Namibia: 2 (MNHUB, ZIN) – „Distr. Grootfontein, leg J. Irish (UG)”, “Farm Hurisib, 19°23’S/17°55’E, Anfang IV. 1989”; 4 (ZIN, ZSM) – “Abachaus, Ojiwarongo Distr., 27. 3. 1955, leg. G. Hobohn” (15. 4. 54, 27 3. 55); 2 (ZSM) – “Njassasee, Mango, 600 m, 21–22. 11. 1958, leg. C. Lindemann”; Republic of South Africa: Lectotype, male (NRS) here designated and 1 Paralectotype, female (NRS) – “Caffraria, J. Wahlb.” (designated in collection by E. Endrödy-Younga in 1968); Angola: 7 (MAT, ZIN) – “Cazomo, Alto Zambese, 4897-7, II-1955, E. Luna de Carvalho”.

Notes. This species is known only from the south part of Africa, but another species vicariant to it remains still undescribed and is known to the writer from East Africa. It could be expected that the species under consideration is connected with some subcortical places and habits like them.

***Urophorus (Anophorus) humeralis* (FABRICIUS, 1798)**

Kirejtshuk, 1992 and others: cosmopolitan with initial natural areal in tropics and subtropics of the Eastern Hemisphere.

Specimens examined: Some hundreds of specimens from Republic of South Africa: 1 (STD) – “Kapland”; 1 (ZMK) – “Durban”; and also from Ghana (MAT, ZIN), Equatorial Guinea (MNHUB, ZIN), Zaire (MAT, ZIN), Uganda (MAT, ZIN, ZMK), Kenya (ZIN, ZMK), Tanzania (MNHUB, ZIN, ZMK), Burundi (MAT, ZIN), Madagascar (MNHUB), Seychelles (MAT, ZIN).

Notes. This species in contrast to other african congeners is characteristic by a comparatively slender and rather pubescent body with peculiar abdominal apex, especially in males (including aedeagal structures). This species can live in both natural and artificial (stored products) conditions, however, it is more common on decaying and dry fruits.

***Urophorus (Anophorus) picinus* (BOHEMAN, 1851), comb. n.**

(= *Urophorus rufangulus* GROUVELLE, 1899, **syn. n.**; *Carpophilus foveicollis*; AUDISIO, 1982)

BOHEMAN, 1851: South Africa; AUDISIO, 1982: Sierra Leone (*C. foveicollis*); WILLIAMS & al., 1983: Cameroon, West, Central and East Africa.

Specimens examined: Namibia: 1 (SMS) – “30. 9–4. 10. 1990, P. Schüle, Bagani”; Republic of South Africa: Holotype *Brachypterus picinus*, female (NRS) – “Caffraria, I. Wahlb.”; Cameroon: Lectotype *Carpophilus rufangulu*, male (DEI) here designated (in collection designated by S. Endrödy-Younga in 1966) – “Lolodorf”; and also about a hundred from Guinea (ZIN), Cameroon (DEI, NRS, ZIN), Zaire (MAT, ZIN), Tanzania (MNHUB, STD, ZIN), Uganda (CNC, ZIN), Rwanda (MAT, ZIN), Burundi (MAT, ZIN).

Notes. The testing of the both types specimens mentioned above give an evidence that they should be regarded as conspecific. This species is mostly similar to *U. (A.) foveicollis* MURRAY, 1864, widely distributed in the Indo-Malayan region and in Réunion (ENDRÖDY-YOUNGA, 1982) differing from the latter in a comparatively smaller and more haired body with peculiar abdominal apex in the both sexes and aedeagal structure.

Amphicrossinae

***Amphicrossus namibiensis* KIREJTSHUK, 1987**

KIREJTSHUK, 1987: 83.

Notes. This species as other representatives of this genus is connected with places of exuded tree sap.

Meligethinae

***Pria (Pria) angustula* COOPER, 1982 (figs. 13–19)**

(= *Pria fallax* GROUVELLE, 1808/1909 – (figs. 20–25).

COOPER, 1982: Republic of South Africa.

Specimens examined: Namibia: 7 (MNHUB, ZIN, SMWN) – “East Caprivi: Mudumu NP, Buffalo Trails Camp, lux, 18°10’S/23°26’E, 12. III. 92, leg. M. Uhlig”; 1 (MNHUB) – “East Caprivi: Mudumu NP, Nakatwa, 18°10’S/23°26’E, 8–13. III. 92, lux, leg. M. Uhlig”; Republic of South Africa: 1 Paratype *P. angustula*, male (NHL) – “Estcourt, Natal, G. A. K. Marshall”; “717”; 1 Paratype, female (NHL) – “Port St. John, Pondoland, April, 1–5, 1923”, “R. E. Turner”; Angola: 1 (NHL) – “5 mls NE Negola, 25. III. 1972”; 1 (ZIN) – “Changa, 21–24. III. 1972”. Ethiopia: Lectotype *P. fallax*, male (MNP) – “Abyss. Raffrai” (designated in collection by S. Endrödy-Younga in 1966).

Notes. This species is characterized by a slender and moderately flattened, straw reddish and slightly shiny body; dense, fine and long greyish hairs on dorsum, almost twice longer than the distance between their roots; more or less distinct punctures on dorsum, nearly as large as eye facets, and the interspaces between them with a dense and fine cellular microreticulation; weakly emarginate fore edge of frons; antennal club in males with 3 somewhat loosen segments; rather narrow legs; pygidial apex acute in males and widely rounded in females. Besides, this species has a peculiar structure of genitalia in the both sexes. Penis trunk of the types *P. fallax* and *P. angustula* has an alone difference in a short excision at apex, and their antennal club are different in width.

This species as other representatives of the Meligethinae is anthophagous.

Pria (Pria) cinerascens ERICHSON, 1843

(= *Pria argenteola* REITTER, 1872)

COOPER, 1982: Republic of South Africa.

Specimens examined: Republic of South Africa: 1 (MNHUB) – “Kapstadt, 29. X–2. XI. 1991, leg. U. Göllner”; 3 (NHL) – “False Bay, 2 mls E Mulzenburg, 3. I. 1972, general sweeping”; 4 (NHL, ZIN) – “Die Panne N.P., Cape Prince, 5–6. I. 1972, swept from dune vegetation”; 5 (MAT, ZIN) – “Cape Prov.: Clanwilliam distr., Sederberg, VII. 1958, J. Smith”; 8 (NRS, ZIN) – “De Vylder, Cap. b. sp.”; 7 (NRS, ZIN) – “Cap. b. sp.”, “Drege”.

Notes. This species has extremely conspicuous silver hairs on dorsum. In other aspects it is with medium size; blackish metasternum and abdomen, comparatively narrow tibiae; punctuation and sculpture of dorsum, antennae as those in *P. dulcamarae* (SCOPOLI, 1763) but darkened (except light scapus); simple tarsal claws and rather generalized structure of aedeagus (as that in *P. dulcamarae*, *P. vicina* and so on). See also notes to *P. (P.) notata*.

This species as other representatives of the Meligethinae is anthophagous.

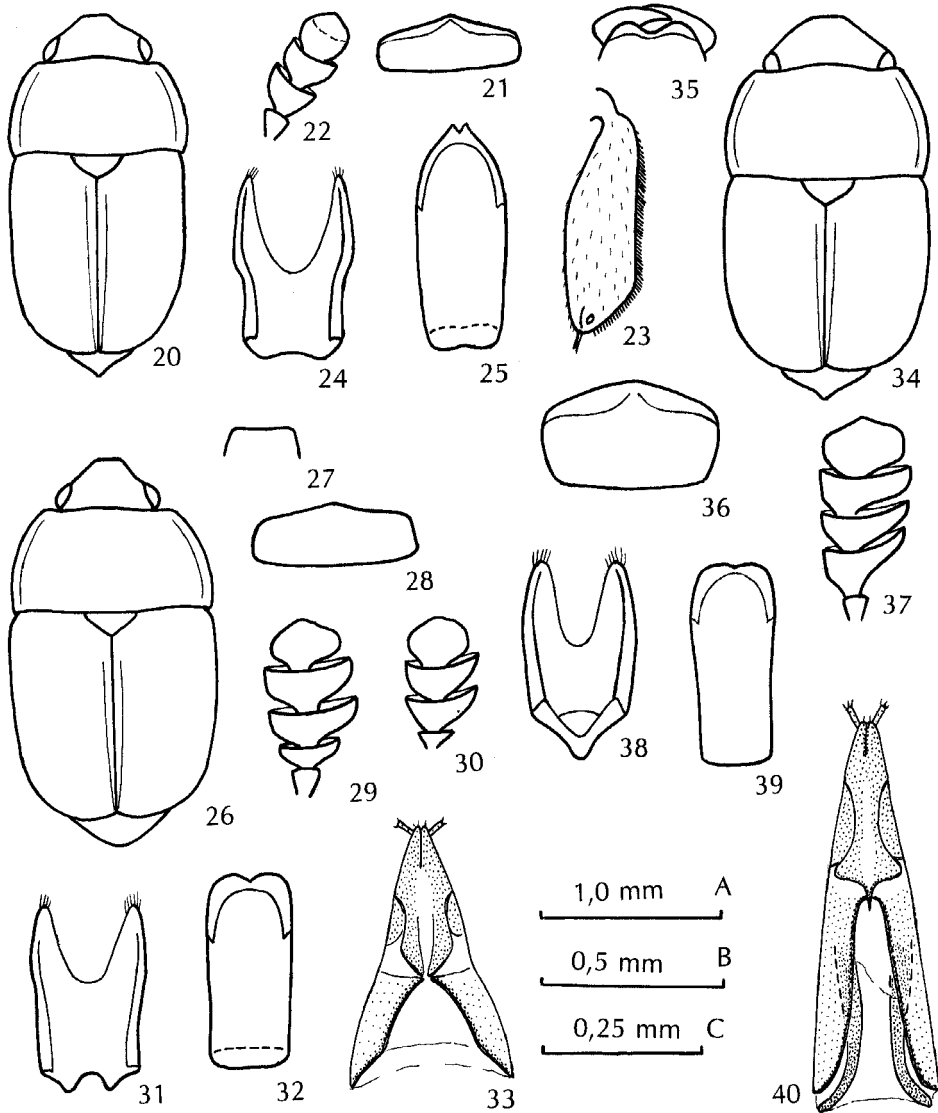
Pria (Pria) magna REITTER, 1872 (figs. 26–33)

(= *Pria concolor* GROUVELLE, 1899, syn. n.; *Pria obscura* COOPER, 1982, syn. n.; ? *Pria crassa* GROUVELLE, 1906).

REITTER, 1872; Grouvelle, 1899; COOPER, 1982, Republic of South Africa.

Specimens examined: Namibia: 1 (MNHUB) – “Darling, 8.83, F. Bahrmann”; Zimbabwe: 2 (MHL, ZIN) – “Rhodesia, Matopo Hills, E. A. C. Schelfe, 1954 (3920)”, “Ex flowers of *Protea lanceolata* E. Mey ex Meissn.”; Republic of South Africa: Lectotype, female *P. magna* (MNHUB), here designated – “Cap. Bon. sp., Dr. Fritsch”, “56968”, “Lectotypus 1967 Endrödy-Younga”; 2 (DEI, ZIN) – “Cape Town, Andreae ded”; 1 Paralectotype *P. concolor*, female (DEI) – “P. B. Spei., Zulus”, “*Pria concolor* GROUV. ty.”; 1 Paratype *P. obscura*, male (NHL) – “Orange Free State, Between Witzieshoek & Mont-aux-Sources, 25. II. 1929”, “Dr Hugh Scott”.

Notes. The body of this species is of medium size (2.5–3.1 mm), but comparatively convex dorsally and with coloration varying reddish to brownish with lighter disks of pronotum and elytra and appendages. It has dense and fine punctuation and microreticulation on interspaces of dorsal surface, 4-segmented antennal club in males. Conspecific position of the types series of *P. magna*, *P. concolor* syn. n. and *Pria obscura* syn. n. is quite evident after comparison of their (alone difference found is 4 segmented and wider antennal club in the first and 3-segmented and narrower one in the second and third type series, but this peculiarity is scarcely traceable in other studied specimens). Another thing is connected with probable attribution to the same species *P. crassa* (known to the writer after 1 Paralectotype, male (MNP), designated in collection as the lectotype by S. Endrödy-Younga in 1966 – “Sicard”, “Madagascar”; 1 female (MNHUB) – “Madagascar int. austr. Hildebrandt S.”; 2 (MAT, ZIN) – “Madagascar, Tamatave, ex coll. Breuning”) which differs from the species under consideration only in a little more oval and wider body with lighter coloration.



Figs. 20–40. Genus *Pria*, subgenus *Pria* s. str. (orig.). – 20–25 *P. (P.) fallax*; 20 body with contour of explanate pronotal sides and subsutural lines, dorsal; 21 mentum; 22 male antennal club; 23 male mid tibia; 24 tegmen, ventral; 25 penis trunk, dorsal; 26–33 *P. (P.) magna*; 26 body with contour of explanate pronotal sides and subsutural lines, dorsal; 27 fore edge of frons; 28 mentum; 29 male antennal club of the lectotype of *P. magna*; 30 male antennal club of the paralectotype of *P. concolor*; 31 tegmen, ventral; 32 penis, dorsal; 33 ovipositor, ventral; 34–40 *Pria (Pria) vicina*; 34 body with contour of explanate pronotal sides and subsutural lines; dorsal; 35 fore edge of frons with mandibles; 36 mentum; 37 male antennal club; 38 tegmen, ventral; 39 penis trunk, dorsal; 40 ovipositor, ventral; Scales: A – to figs. 20, 26, 34; B – to figs. 23, 27, 35; C – to figs. 21–22, 24–25, 28–33, 36–40.

***Pria (Pria) notata* COOPER, 1982**

COOPER, 1982: Republic of South Africa.

Specimens examined: Zimbabwe: 1 (NHL) – “Rhodesia, Matopo Hills, E. A. C. Schelfe, 1954 [3920]”, “Ex flowers of *Protea lanceolata* E. Mey ex Meissn.”; Republic of South Africa: 2 (NHL, ZIN) – “Natal, Drakensberg, Cathedral Peak, 18. XII. 1951”, “From *Protea abyssinica*, Flower N 3, Stage 4”.

Notes. The specimen from Zimbabwe is more robust, with slightly shorter pronotum, narrower tibiae and less darkened apices of elytra. This species is well characterized by its coloration and genital structures, having general appearance as in *P. (P.) cinerascens* and *P. (P.) robiginosa* but differing from the both in truncate fore edge of frons, and also from the former in coloration and pubescence. Moreover, all three forms have quite distinguishable genitalia in the both sexes.

***Pria (Pria) palpata* KIREJTSHUK, 1989**

KIREJTSHUK, 1989: Mozambique, Republic of South Africa.

Specimens examined: Zimbabwe: 5 (NHL, ZIN) – “Salisbury, Mashanaland, G. A. K. Marshall, VIII. 1900”.

Notes. This species is rather similar to *P. (P.) magna* differing from it in slightly darker body, larger mentum, longer antennae and palpi. Size and shape of mentum of *P. (P.) palpata* have somewhat an intermediate position between those of *P. (P.) magna* and *P. (P.) vicina*. Moreover, this species has abnormally long palpi quite distinct from those in other member of the genus.

***Pria (Pria) robiginosa* COOPER, 1982**

COOPER, 1982: Republic of South Africa.

Specimens examined: Zimbabwe: 8 (NHL, ZIN) – “Rhodesia, Matopo Hills, E. A. C. Schelfe, 1954 (3920) “Ex flowers of *Protea lanceolata* E. Mey ex Meissn.”; Republic of South Africa: 2 paratypes *P. robiginosa* (NHL) – “Zululand, Eshowe, 23–30. IV. 1926”, “R. E. Turner” (6–31. V. 1926).

Notes. Externally this species is strongly similar to *P. (P.) notata* (see above), but well diagnosed from it due to widely emarginate fore edge of frons and peculiar genital structures (especially in ovipositor).

***Pria (Pria) testacea* GROUVELLE, 1908/1909**

COOPER, 1982: Zimbabwe.

Specimen examined: Angola: 1 (NHL) – “A.11), Bruco, 26.II–2. III. 1972, at light”; Zimbabwe: 5 (NHL, ZIN) – “Salisbury, Mashanaland, G. A. K. Marshall, VIII. 1990”.

Notes: This species is very similar to *P. (P.) pauli* GROUVELLE, 1908/1909 (= *P. (P.) fragilis* COOPER, 1982, **syn. n.**; ? *P. (P.) ochracea* COOPER, 1982; ? *P. (P.) rufipes* COOPER, 1982)², differing from it in more oval body with longer and more conspicuous pubescence, truncate fore edge of frons, wider antennal club in males and shorter ovipositor. At the same time, this species has some resemblance to *P. (P.) magna*, but it is somewhat smaller and comparatively wider and less convex.

² *P. (P.) pauli* GROUVELLE, 1908/1909 is known to the writer after study 1 ? paralectotype *P. pauli* (MNHUB), designated in collection as the lectotype by S. Endrödy-Younga – “Usambara, Kwai, P. Weise S.” and 3 paralectotypes (MNHUB, ZIN) – “Afr. or. all. Kwai, Weise”; 2 paratypes *P. fragilis* (NHL) – “R. A. Turner, Kimosi, Mch. Apl. 1932”; 1 paratype *P. adusta* COOPER, 1982 (NHL) – “on blossom”, “Mt. Cameroon, Onyanga, 8100 ft, 22. 1. 1932, M. Steele”; 2 paratypes *P. rufipes* COOPER, 1982 (NHL) – “Beaten from trees near lake shore”, “Abyssinia, Mt. Zuqua’la circa, 9000 ft, 22. X. 1926; Dr. H. Scott”. The proposed synonymy has been made evident due to comparison of the specimens and descriptions of names mentioned above, though the names *P. adusta* and *P. rufipes* can be reliably synonymized only after a study of the holotypes of these type series because the studied paratypes of them include some unsimilar species.

***Pria (Pria) vicina* GROUVELLE, 1908/1909 (figs. 34–40)**

(= *Pria villosa* COOPER, 1982, syn. n.)

GROUVELLE, 1908/1909: Tanzania; COOPER, 1982 (as synonym of *P. dulcamarae* (SCOPOLI, 1763)): ? Madeira, ? Southern Yemen, ? Ethiopia; ? Kenya, ? Tanzania, ? Rwanda, ? Zaire, ? Zimbabwe.

Specimens examined: Zimbabwe: 2 paratypes *P. villosa* COOPER (NHL) – “S. Rhodesia, Umtali, III. 1957, N. L. H. Krass”; Tanzania: 1 paralectotype *P. vicina* (MNP) designated by S. Endrödy-Younga – “Nuelo, Usambara, H. Rolle”; and also more than a hundred specimens from Zaire (MAT, ZIN), Ethiopia (ZIN), Uganda (MAT, ZIN), Kenya (MNHUB, NHL, ZIN), Tanzania (MNHUB, ZIN), Mozambique (NHL, ZIN).

Notes. This species has light brown and not strongly convex body of moderate size (1.8–2.3 mm); dense and short rather conspicuous hairs on dorsum, slightly longer than the distance between their roots; fine and dense punctation, densely and finely microreticulated interspaces on dorsum; very large mentum of its comparatively large head with a rather long antennae bearing a large 4-segmented club. Externally it is similar to *P. (P.) magna*, *P. (P.) palpata* and other species like them, but differs from them in a comparatively deeply emarginate fore edge of frens, large mentum and strongly developed antennae.

Indication of the palaeartic *P. P. dulcamarae* (SCOPOLI, 1763) for the composition of South Africa (see COOPER, 1982) should be regarded as rather questionable, and most specimens from localities southern from Sahara identified by M. C. COOPER as this species probably belong to *P. P. vicina*.

***Meligethes (Lariopsis) arcuatus* REITTER, 1872**

(= *Meligethes arcuatus* REITTER, 1872; *Meligethes limbatus* REITTER, 1872, syn. n.; *Meligethes (Lariopsis) arcuatus*: KIREJTSHUK, 1989).

REITTER, 1872: Republic of South Africa.

Specimens examined: Namibia: 1 (MNHUB) – “Brit. S. W. Afrika, Kl., Namaland, Steinkopf, L. Schultze”; Republic of South Africa: Lectotype *M. arcuatus*, male (MNHUB), here designated and 2 paralectotypes, females – “Cap, Edmüller”, “*arcuatus* Reitt., Cap”, “8600”; 1 paralectotype *M. rufiventris*, female (MNHUB) – “*rufiventris* Reitt., Cap, Reitter”, “56961”; Lectotype *M. limbatus*, female (MNHUB), here designated – “*limbatus* Reitt., Cap, Reitter”, “56963”; 1 paralectotype *M. arcuatus*, male (ISW) – “Cap, Drege, *arcuatus*”; 1 paralectotype *M. rufiventris*, female (IWS) – “Cap, Drege, *rufiventris* Reitt.”; 1 (MNP) – “C. B. Esp. Raffray” (det. Grouvelle as *M. limbatus*); 26 exx (ZIN, ZMU) – “C. b. sp.”, “*flavoventris*”, “*luridipennis*” (coll. V. Motschulsky); 1 (ZIN) – “W. Capland, Bushmann, Darling, VIII. 83”; 34 (SMD, ZIN) – “Capland, M. H. Muche”; 5 ex (MAT, ZIN) – “Le Cap, ex Staudinger”; 1 (MAT) – “Richmond, Natal. XII. 54, ex coll. Breuning”; 28 (NRS, ZIN) – “De Vylder, Cap. b. sp.”; 1 (NHL) – “Cap b. Esp., *chrysocoma*”; 1 (NHL) – “Cape G. H., 18.45”; 1 (CAS) – “Paarl, Cape Prov., X.16.49, Malkin”; 37 (NHL, ZIN) – “Lion’s Head, Cape Town, Aug. 1920, R. E. Turner”; 3 (NHL, ZIN) – “Seapoint, Aug. 1905, G. A. K. Marschal”.

Notes. Synonymy of *M. (L.) arcuatus* and *M. (L.) limbatus* is quite evident because these names were proposed for colour varieties of the same species, characterizing by a comparatively slender body with medium size (2.0–2.5 mm), rather narrow all tibiae, moderately sparse and large teeth along outer edge of fore tibiae, strongly curved male hind tibiae and peculiar aedeagal structure. The differences between *M. (L.) arcuatus* and *M. (L.) rufiventris* can be traced mostly in character of dorsal punctation and sculpture, shape of tibiae and will be considered in detail in one paper by the writer and P. AUDISIO.

***Meligethes (Lariopsis) haagii* REITTER, 1872**

(= *Meligethes haagii* REITTER, 1872; *Meligethes (Lariopsis) haagii*: KIREJTSHUK, 1989).

REITTER, 1872: Republic of South Africa.

Specimen examined: Namibia: 1 male (NHL) – “S. W. Africa, R. E. Turner”, “Aus, 8=30. XI. 1929”; Republic of South Africa: 1? syntype (MNP) – “Cap C. Hagen”, “Cap, puperula Chev.”; 3 (ZIN, ZMU) – “Cape Bon. Sp.”, “coll. Motschulsky”; 6 (TMP, ZIN) – “Willowmote, XII. 1919, Dr. Brawns”.

Notes. This species is similar to *M. (L.) nebulosus* REITTER, 1872, and *M. (L.) sphaeroides* KIREJTSHUK et EASTON, 1988, but distinct from it in larger (2.1–2.4 mm) and more subquadrangular body, with coarser

punctuation and more conspicuous pubescence on dorsum, longer head with distinctly emarginate fore edge of it viewed as a shiny stripe, widely truncate pygidial apex in males and peculiar aedeagal structure.

Meligethes (Lariopsis) pulchellus REITTER, 1872

(= *Meligethes pulchellus* REITTER, 1872; ? *Meligethes (Lariopsis) pulchellus*: KIREJTSHUK, 1989).

REITTER, 1872: Republic of South Africa.

Specimens examined: Angola: 45 (NHL, ZIN) – “Tundavala, 8–10 mls Sa da Bandeira, 27–29. III. 1972, general sweeping” (and “at light”); 4 (NHL, ZIN) – “(A 37) 5 mls NE Negola, 25. III. 1972, at light”; Republic of South Africa: 1 (MNP) – “C. B. Esp. Peringuey” (det. Grouvelle as *M. pulchellus*) (26. 01. 1992); 1 (ZMK) – “Caffraria, *N. viridiaenea* Dej.”; 7 (NHL, ZIN) – “Orange F. State Harrismith, Feb. 1927”, “R. E. Turner”; 1 (NHL) – “Transvaal”; 1 (NHL) – “Natal, Van Reenen, Drakensberg, Nov. 1926”, “R. E. Turner”; 1 (NHL) – “Port St. John, Pondoland, 16–28. IV. 1924”, “R. E. Turner”; 1 (TMP) – “S. Afr. Namagualand, Koekenaap, 31.32S-18.14E, 31. 8. 1979, white Mesem, bush, Endrödy-Younga”; 1 (ZIN) – “S. Afr. Namagualand, Katzesrus, 30.57S-17.50E, white dunes day, Endrödy-Younga”.

Notes. The preliminary synonymy of *M. pulchellus* and *M. cercoides* till study the type series is connected that the light specimens of the latter are almost identical externally with smallest specimens of *M. (L.) rufiventris* REITTER, 1872, reliably differing from it only in aedeagal structure. At the same time, *M. (L.) pulchellus* is strongly variable in coloration (straw reddish to dark brownish) punctuation (size and density) and microreticulation on dorsal interpaces between punctures (beginning from completely smooth to fine, dense and extremely conspicuous microreticulation). However, all specimens from Angola are dark and most of them with comparatively sparse dorsal punctuation and dull interspaces.

Meligethes (Lariopsis) variabilis REITTER, 1872

(= *Meligethes variabilis* REITTER, 1872; *Meligethes (Lariopsis) variabilis*: KIREJTSHUK, 1989).

REITTER, 1872: Republic of South Africa.

Specimens examined: Namibia: 3 (MNHUB, ZIN) – “Darling, 8.83, S. W. Africa, M. Calba F. Bartmann”; 1 (TMP) – “S. W. Afr., S. Namib, Rosh-Pinah, 27.53D-18.50E”, “22. 9. 1973; E-Y: 137, from flowers, leg. Endrödy-Younga”; 1 (MNHUB) – “Brit S. W. Afrika Kl. Namaland Steinkoff VIII.04 L. Schultze S.”; Republic of South Africa: Lectotype *M. variabilis*, male (MNHUB), here designated and 2 paralectotypes (MNHUB) – “757”, “Par. G.”, “*variabilis* REITT., Pr. b. sp.”; 1 paralectotype var. *suturalis* (MNHUB) – “Reitt. 876”; 2 paralectotypes ? var. *suturalis* (MNHUB) – “16”; 3 paralectotypes var. *suturalis* (IZW) – “Cap”, “*variabilis* REITT. var. *suturalis* (IZW) – “Cap”, “*variabilis* REITT. var. *suturalis* REITT., Cap, Drege”, “ex Coll. Mus. Stettin”; 1 paralectotype var. *luridipennis* (IZW) – “Cap”, “*variabilis* REITT., var. *luridipennis* REITT., Cap, Drege”; 3 paralectotypes var. *confluens* (IZW) – “Cap”, “*variabilis* REITT. var. *confluens* REITT., Cap, Drege”, “ex Coll. Mus. Stettin”; 1 plt (*variabilis*) des. Kir. (Warszawa Inst.) – “Cap”, “*variabilis* REITT., Cap, Drege”; 2 paralectotypes var. *bimaculatus* (IZW) – “Cap”, “*variabilis* REITT. var. *bimaculatus* REITT., Cap., Drege”, “ex Coll. Mus. Stettin”; 5 ? paralectotypes (MNHUB, ZIN) – “*fenestrata* N.”, “8604”, “*plagiatus* v. WINTH”, “Pr. b. sp. Luchtenst”, (coll. Klug); 2 paralectotypes (MNHUB) – “*suturalis* v. WINTH”, “8603”, “Afr. austr., Ec. Kl.”, (coll. Klug); 1 ? paralectotype (MNHUB) – “*N. fusca* v. WINTH”, “Afr. austr., Ec. Kl.”, (coll. Klug); 2 ? paralectotype (MNHUB) – “*N. fusca* v. WINTH”, “Afr. austr., Ec. Kl.”, “8606”, (coll. Klug) (26. 01. 1992); 5 ? paralectotypes (MNHUB, ZIN) – “*Chrisoromae* v. WINTH”, “Vitenh, Krebs”, “8602”, (coll. Klug); 2 ? paralectotypes (MNHUB, ZIN) – “*cinctus* v. WINTH”, “Afr. austr. Ec. Kl.”, “8605”, (coll. Klug); 1 ? paralectotype (MNHUB) – “*fuscipes* v. WINTH”, “Afr. austr. Ec. Kl.”, “8607”, (coll. Klug); 1 (MNHUB) – “S. Afrika VIII. 04, L. Schultze S.”; 48 (ZIN, ZMU) – “C. b. sp.”, “*fuscus*”, “*palliat*”, “*flavolimbatus*”, “*biplagiatus*”, “*suturalis*”, “*conferta*” (“*Aethina*”), “*angusta*”, “*aciculata*” (“*Aethina*”) (Coll. V. Motschulsky); 3 ex (MAT, ZIN) – “Cape Prov., Clauwilliam distr., Sederberg, VII. 1958, J. Smith”; 3 (NHL) – “Cap, Drege”, “*cinctus*” (“*fuscipes*”, “*extensus*”); 6 (NHL, ZIN) – “Lion’s Head, Cape Town, Aug. 1920, R. E. Turner”; 1 (NHL) – “Camps Bay, Cape Peninsula, Sept/1920, R. E. Turner”; 3 (NHL) – “Cape Province, Somerset East, November 1930, R. E. Turner” (October 1930 and Sept. 1930); 1 (NHL) – “Cape Province, Montag, 1–21 Oct. 1924, R. E. Turner”; 1 (NHL) – “Ceres, cape Province, 1500 ft. 27. X–1. XI. 1920, R. E. Turner”; 1 (NHL) – “Worchester, Cape Province, Sept. 1928, R. E. Turner”; 1 (ZIN) – “Cape Province, Mont Bay, 3. XI. 51, A. Duke, On Compositae”; 3 (NHL) – “Town Lion’s Hd Mt, 15. XI. 51, A. Duke, On Compositae”; 1 (NHL) – “Cape Province, Cape District, J.

Balfour-Browne", "on flowers of *Euryops abrotanifolius* (Compositae)"; 2 (NHL, ZIN) – "Stellenbosch C. C., Nov. 1904, G. A. K. Marschall"; 2 (NHL, ZIN) – "Cape Town, Sea Point, X. 1950, G. A. K. Marschall"; 1 (CAS) – "Cape Province, Oudtshoorn, 25. X. 49, Boris Malkin"; 7 (CAS, ZIN) – "Elgin, Cape Prov., XI. 5. 49, B. Malkin"; 1 (NHL) – "Natal, Park Rynie, 12. X. 51, A. Duke"; and a hundred other specimens from RSA (MAT, TMP and many other collections).

Notes. This species differs from the member of the subgenus *Lariopsis* KIREJTSHUK, 1989 in comparatively large (3.2–4.1 mm) and rather shiny body with great variation in coloration, reduced dorsal pubescence, excised fore edge of frons, slightly curved hind tibiae and peculiar aedeagal structure.

Meligethes (Chromogethes) illustris GROUVELLE, 1899

(= *Meligethes illustris* GROUVELLE, 1899; *Meligethes (Chromogethes) illustris*: KIREJTSHUK, 1989).

GROUVELLE, 1899; SPORNRAFT & KIREJTSHUK, 1993: Republic of South Africa.

Specimens examined: Zimbabwe: 1 (NHL) – "S. Rhodesia: Mtarazi Falls, III. 1957, N. L. H. Krauss"; 1 (ZIN) – "S. Rhodesia: Chapangu Falls, III. 1957, N. L. H. Krauss"; Republic of South Africa: 2 (NHL, ZIN) – "Natal: Nqutu, 11. XII. 54, A. H. Newton, Spec. AB, on *Haplocarpus*"; 26 (NHL, ZIN) – "Natal: Nqutu, 8. II. 54, A. H. Newton, Spec. AB, "Yarrow"; 3 (NHL, ZIN) – "Natal: Nqutu, 11–13. III. 54, On *Sedum*, A. H. Newton"; 2 (NHL, ZIN) – "Natal: Nqutu, On *Helichrisum nudiflorum* Nees, 6. II. 61, A. H. Newton"; 2 (CMO, ZIN) – "Natal, 75 km WSW Estcourt Cathedral, Peak For. Sta., S. & J. Peck", "Philipps Folly Trail, Sweeping grassland, 8. XII. 1979" and "Riverside shrubd, night beating, 14. XII. 1979"; 10 (TMP, ZIN) – "2 m. W of Iron Crown, Wolkberge, Haenertsburg distr., 7000 ft, III. 1970, O. & L. Prazesky"; 1 (NHL) – "Sterkfontein, Transvaal, H. P. Thomasset"; 1 (NHL) – "Natal, Van Reenen, Drakensberg, Nov. 1926", "R. E. Turner"; and also about a hundred of specimens from RSA (MNHUB, NHL, ZIN and other collections).

Notes. This species is characterized by rather dense and deep punctures on dorsum (especially on elytra, where they elongate and contiguous), rather elongate head (much longer than the distance between eyes), truncate or even weakly convex fore edge of frons, widely rounded and unprojecting fore and hind pronotal corners, moderately wide brownish legs, even crenellation along fore edge of fore tibiae, flattened to slightly concave metasternum in males, peculiar aedeagal structure.

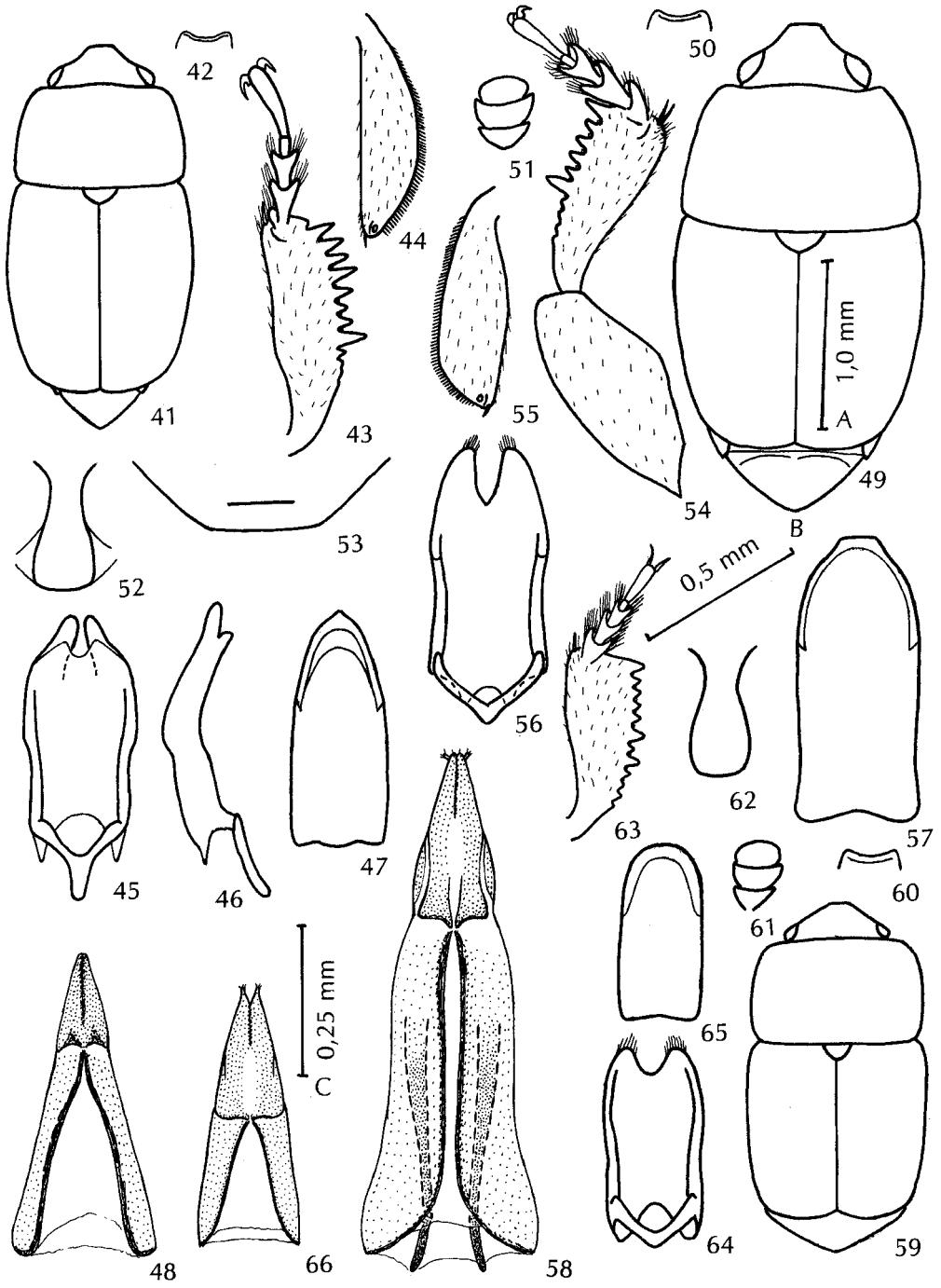
Meligethes (Clypeogethes) annae KIREJTSHUK, sp. n. (figs. 41–48)

Specimens examined: Republic of South Africa: Holotype, male (TMP) and 15 paratypes (CMO, TMP, ZIN) – "Natal, 75 km WSW Westcourt, Cathedral Peaks For. Sta., 7–31. II. 79, S. & J. Peck", "Phillips Folly, 2070 m, trail head, 8. XII. 79, beating *Protea* woodland shrubs".

Description. Male, holotype. Length 2.2, breadth 1.1, height 0.7 mm. Moderately convex dorsally and ventrally; black with reddish dark brown mouth parts, fore part of frons, antennae and fore tibiae, and with brownish antennal club and tarsi; body with a moderate shine; dorsum with moderately fine and long, yellowish grey hairs, 1.5 times as long as the distance between their roots on elytra.

Head and pronotal surface with oval and distinct punctures, somewhat larger than eye facets, interspaces between them on pronotum about one puncture diameter, with a smoothed and cellular microreticulation. Elytral surface with smaller and less regular punctures, disposed behind fine transverse striae, interspaces between punctures broader than one puncture diameter. Pygidial surface almost finely granulose and coarsely punctured. Ventral surface more sparsely punctured; prosternal process and metasternum with deep punctures, larger than eye facets, interspaces between them more than one puncture diameter, smooth; ventrites with punctures, less than eye facets, interspaces 2–4 puncture diameters, alutaceous to smoothly microreticulated.

Figs. 41–66. Genus *Meligethes*, subgenus *Clypeogethes* (orig.). – 41–48 *M. (C.) annae* sp. n.; 41 body shape, dorsal; 42 fore edge of head, dorsal; 43 male fore tibia and tarsus, dorsal; 44 male mid tibia, dorsal; 45 tegmen, ventral; 46 idem, lateral; 47 penis trunk, dorsal; 48 ovipositor, ventral; 49–58 *M. (C.) antlia* sp. n.; 49 body shape, dorsal; 50 fore edge of head, dorsal; 51 antennal club; 52 prosternal process, ventral; 53 male hypopygidium, ventral; 54 male fore legs, dorsal; 55 male mid tibia, dorsal; 56 tegmen, ventral; 57 pensil trunk, dorsal; 58 ovipositor, ventral; 59–66 *M. (C.) arcopenis* sp. n.; 59 body shape, dorsal; 60 fore edge of head, dorsal; 61 antennal club; 62 prosternal process, ventral; 63 male fore tibia and tarsus, dorsal; 64 tegmen, ventral; 65 penis trunk, dorsal; 66 ovipositor, ventral; Scales: A – to figs. 41, 49, 59; B – to figs. 42, 50, 52–53, 60, 62; C – to figs. 43–48, 51, 54–58, 61, 63–66.



Head slightly convex from above, somewhat shorter than the distance between eyes, with an arched shiny line at inner edge of each eye. Antennae $4/5$ as long as head broad, their club nearly $2/3$ as wide as long, with rounded apex and composing about $1/3$ of total antennal length. Pronotum with gently sloping and unexplanate sides, its corners widely rounded. Elytra with steeply sloping sides and expressed subsutural lines in distal half, but without a trace of humeral stria. Pygidium with widely rounded and slightly explanate apex. Prosternal process rather broad, $1/4$ as wide as antennal club, and with hemicircular and unbordered apex. The distance between mid coxae 2 times and that between hind ones almost 4 times more than that between fore coxae. Metasternum flattened. Caudal marginal line behind hind coxal cavities follows closely hind edge of coxal cavities. Hypopygidium simple and with a gently curved hind edge.

Tibiae subparallel sided and with subequal width; mid and hind tibiae with dense, thin and comparatively long setae. Femora nearly 2 times as wide as correspondent tibia. Fore tarsi $1/2$ as wide as fore tibiae, but mid and hind ones much narrower, claws with a weak tooth at base.

Aedeagus well sclerotized.

Female. Differs only in narrower fore tarsi ($1/3$ as wide as fore tibiae).

Ovipositor moderately sclerotized and without styli.

Variations. Length 1.9–2.4 mm. Smaller specimens are more slender and convex. Some paratypes strongly shiny and with more sparse punctation. Pubescence varies from greyish to golden reddish.

Notes. This new species is similar to small representatives of the *amplicollis* species group sensu lato (SPORNRAFT & KIREJTSHUK, 1993), but quite distinct from them in comparatively narrow tibiae and peculiarities of genitalia of the both sexes.

The name of this species is created from the surname of the author's mother ((Anna A. KIREJTSHUK (Zelenina)).

Meligethes (Clypeogethes) antlia KIREJTSHUK, sp. n. (figs. 49–58)

Specimens examined: Republic of South Africa: Holotype (NHL) and 28 paratypes (NHL, ZIN) – “Natal, Nqutu, 1. III. 62, A. H. Newton”, “On Germanica geniculata” (21. II. 1962).

Description. Male, holotype. Length 3.0, breadth 1.5, height 0.9 mm. Oval, rather convex dorsally and moderately ventrally; dark chestnut brown (head and pronotum almost blackish, with reddish palpi of mouth parts, antennae and legs; dorsum rather shiny, with dense, well conspicuous, greyish hairs, about twice as long as the distance between punctures; ventral surface slightly shiny because of very dense pubescence, as conspicuous and long as that on dorsum).

Head surface with oval punctures not larger than eye facets, interspaces between them 0.2–0.5 puncture diameter, completely smooth. Pronotal surface about as that on head, but punctures larger than eye facets and interspaces between them 0.5 puncture diameter, smooth. Elytral surface with feebly expressed transrugosity in combination with transverse rows of punctures nearly as large as those on pronotum disposed behind striae, interspaces completely smooth. Scutellum with few sparse punctures and with dotted microreticulation. Pygidial surface almost finely granulose with a partly reduced obscure punctation. Surface on prosternal process and metasternum with punctures considerably larger than those on dorsum with smooth interspaces nearly as broad as a puncture diameter, the middle of 1st ventrite about as that on head, and the remainder of ventrites similar to that on pygidium though with more expressed deep and dense punctures.

Head feebly convex, $5/7$ as long as the distance between eyes. Antennae $4/5$ as long as head broad, their club composing $1/4$ of total antennal length, with suboblique apex. Pronotum with gently sloping and unexplanate sides, its hind corners very widely rounded. Scutellum subhemispherical. Elytra with steeply sloping sides, expressed parallel subsutural lines and traceable humeral striae. Pygidium with a widely rounded, nearly transverse apex. Prosternal process 1 and $1/4$ as wide as antennal club, and slightly arcuately widened before almost hemicircular and bordered apex. The distance between mid coxae 2 times and that between hind ones almost 4 times more than that between fore coxae. Metasternum flattened with a short medial depression in the middle, its fore edge straight. Caudal marginal line behind hind coxal cavities follows closely hind edge of coxal cavities. Hypopygidium with a transverse, smooth and shiny protuberance before the middle of transverse hind edge.

Fore tibiae slightly narrower, mid and hind ones slightly wider than prosternal process; mid and hind tibiae with dense, stout and short setae. Fore femur twice, mid one -1.5 times and hind one $-1\ 3/4$ as wide as correspondent tibiae. Fore tarsi half, but mid and hind ones more than 3 times narrower than correspondent tibiae, claws simple.

Aedeagus well sclerotized.

Female. Differs from the male in a convex metasternum, rounded and simple apices of pygidium and hypopygidium, narrower fore tarsi (nearly a third as wide as correspondent tibia).

Ovipositor moderately sclerotized.

Variations. Length 2.1–3.0 mm. A considerable variability is manifested in general coloration (chestnut brown to black), shape of pronotum comparatively wider than elytra in smaller specimens and scarcely wider in larger ones. Body appendages of some specimens are rather dark, i.e. hardly lighter than the general coloration. Configuration of crenellation along outer edge of fore tibiae, punctuation and pubescence are rather variable within the paratypes.

Notes. This species is rather similar to the *pubescens* species-group of the subgenus *Clypeogethes* SCHOLTZ, 1932, which should be defined in one of further papers by P. AUDISIO and K. SPORNRAFT, but it is easily diagnosed due to widened fore femur, peculiar outline of fore tibiae with rather narrow tarsi, character of punctuation and genital structure of the both sexes.

***Meligothes (Clypeogethes) arcopenis* KIREJTSCHUK, sp. n.** (figs. 59–66)

Specimens examined: Namibia: Holotype, male (MNHUB) & 3 paratypes (MNHUB, ZIN, SMWN) – “Distr. Grootfontein, leg. J. Irish (U. G.)”, “Farm Hurisib, 19°23’S/17°55’E, Anfang IV. 1989”; 1 paratype (MNHUB) – “Grootfontein: Askavolt-Farm, 20 km E Otavi, 19°40’S/17°33’E, 18. II. 92, leg. U. Göllner”; 3 paratypes (TMP, ZIN) – “Abachaus, Damaraland”, “VI. 1951, C. Koch” and “XII. 1951, G. Hobohm”.

Description. Male, holotype. Length 1.8, breadth 1.0, height 0.5 mm. Oval, moderately convex dorsally and ventrally; unicoloured black, with reddish palpi of mouth parts and dark brown antennae and legs; dorsum shiny, with not dense and moderately conspicuous greyish hairs, nearly 1.5 times as long as the distance between their roots; ventral surface slightly shiny, with more conspicuous and longer pubescence in comparison with that on dorsum.

Head surface with oval punctures nearly as large as eye facets or slightly larger, interspaces between them 0.5–1.0 puncture diameter, with scarcely expressed alutination. Pronotal surface similar to that on head, but with a little larger punctures becoming larger to base (up to 1.5 times as large as eye facets). Elytral surface as that on pronotum at base, but punctures yet larger (to 2 times as large as eye facets). Scutellum with sparse punctures as large as those on pronotum and with a little smoothed cellular microreticulation. Pygidial surface almost finely granulose with punctures as those on pronotum and scutellum. Surface on prosternal process and 1st ventrite with punctures as those on pronotum and scutellum, with interspaces between them more than one puncture diameter, smooth or smoothed, but punctures on metasternum as large as elytra or somewhat larger, with interspaces less than one puncture diameter; remainder of ventrites as that on pygidium though with less expressed granulation between punctures.

Head slightly convex, nearly as long as the distance between eyes, its fore edge viewed as a wide shiny brownish stripe without punctuation. Antennae 3/4 as long as head broad, their club nearly 1/3 total antennal length, with suboblique apex. Pronotum with gently and steeply sloping unexplanate sides, its hind corners blunt with subangular apex. Scutellum subhemicircular. Elytra with steeply sloping sides, without expressed subsutural lines, but with a trace of humeral striae. Pygidium with a subacute apex. Prosternal process almost twice as wide as antennal club, and arcuately widened before almost hemicircular apex with a transverse unbordered hind edge. The distance between mid coxae 2 times and that between hind ones 3.5 times more than that between fore coxae. Metasternum flattened, its fore edge almost straight. Caudal marginal line behind hind coxal cavities follows closely hind edge of coxal cavities. Hypopygidium simple and with a transverse hind edge.

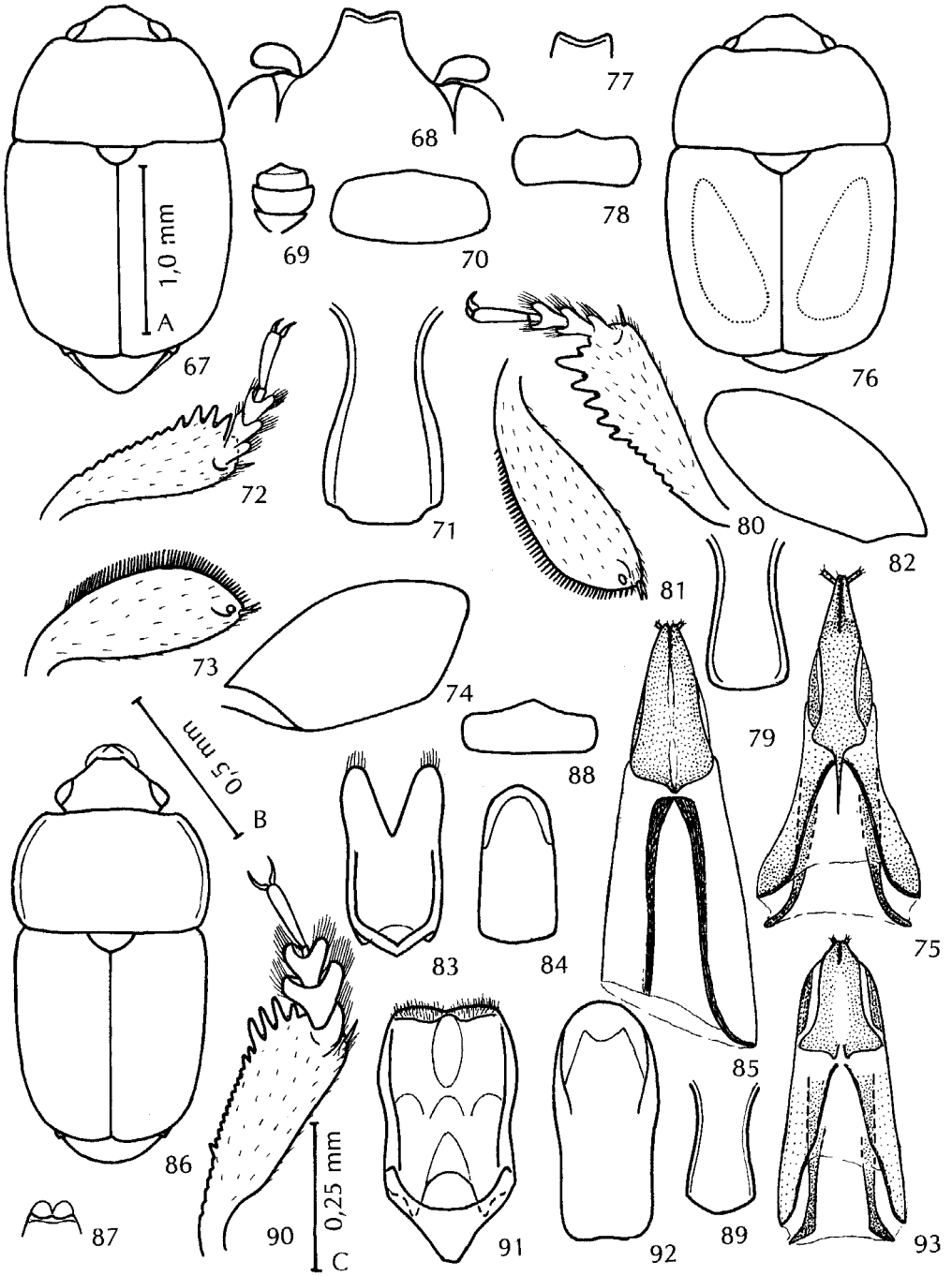
Fore tibia 1 1/3, mid and hind ones as wide as antennal club or slightly wider; mid and hind tibiae with dense, stout and short setae. Femora nearly 1.5 times as wide as correspondent tibia. Fore tarsus 2/3 as wide as corresponding tibia, but mid and hind ones much narrower, claws slightly bulged ad base.

Aedeagus moderately sclerotized.

Female. Differs from the male in narrower fore tarsi and gently rounded apices of pygidium and hypopygidium.

Ovipositor slightly sclerotized.

Variations. Length 1.7–2.3 mm. A little variability is manifested in configuration of outer edge of fore tibiae, punctuation and microreticulation. The male from Askavolt-Farm has a well sclerotized aedeagus.



- ◀ Figs. 67–93. Genus *Meligethes*, subgenus *Clypeogethes* (orig.). 67–75 *M. (C.) asignifer* sp. n.; 67 body shape, dorsal; 68 fore part of head with shiny liniae at inner edge of eye, dorsal; 69 antennal club; 70 mentum; 71 prosternal process, ventral; 72 female fore tibia and tarsus, dorsal; 73 female mid tibia, dorsal; 74 female hind femur, ventral; 75 ovipositor, ventral; 76–85 *M. (C.) bisignifer* sp. n.; 76 body shape with a dotted contour of reddish spot on elytra, dorsal; 77 fore edge of head, dorsal; 78 mentum; 79 prosternal process, ventral; 80 female fore tibia and tarsus, dorsal; 81 female mid tibia, dorsal; 82 female hind femur, ventral; 83 tegmen, ventral; 84 penis trunk, dorsal; 85 ovipositor, ventral; 86–93 *M. (C.) rugifer*; 86 body shape with a dotted contour of reddish spot on elytra, dorsal; 87 fore edge of head, dorsal; 88 mentum; 89 prosternal process, ventral; 90 male fore tibia, dorsal; 91 tegmen, ventral; 92 penis trunk, dorsal; 93 ovipositor, ventral; Scales: A – to figs. 67, 76, 86; B – to figs. 68, 77, 87; C – to figs. 69–75, 78–85, 88–93.

Notes. This new species can be considered as a member of the species group, including *M. (C.) reticulatus* REITTER, 1872, and *M. (C.) serrator* REITTER, 1872, (see 2d part of this paper) differing from them in fore edge of frons, configuration of fore tibiae, comparatively wide prosternal process and peculiar genitalia of the both sexes.

***Meligethes (Clypeogethes) asignifer* KIREJTSHUK, sp. n. (figs. 67–75)**

Specimen examined: Namibia: Holotype, female (SMWN) – “Bushmanland, Klein Dobe, 19°25'S/20°21'E, 20–21. II. 92, leg. M. Uhlig”.

Description. Female, holotype. Length 2.2, breadth 1.7, height 0.9 mm. Oval, rather convex dorsally and ventrally; unicoloured bright reddish, comparatively light, but head and last tarsomeres somewhat darkened; body with a faint fat shine; dorsum with sparse, hardly conspicuous, very short hairs, shorter than the distance between their roots; ventral surface with more conspicuous and longer hairs (longer than the distances between their roots).

Head surface with oval and comparatively shallow punctures considerably less than eye facets, interspaces between them 1.0–2.0 puncture diameters, with a smoothed, cellular microreticulation. Pronotal and scutellar surface as that on head, but punctures in the middle a little smaller and interspaces between them 2.0–3.5 puncture diameters. Elytral surface with a not quite distinct transrugosity and asymmetric, very shallow punctures somewhat larger than those head and pronotum, but more sparse and arranged into transverse rows behind each furrow, interspaces between punctures in a row about 2–3 cross-section of a puncture. Pygidial surface almost finely granulose with a scarcely expressed punctation and dense, very relief microreticulation. Surface on prosternal process, metasternum and 1st ventrite with large and deep distinct punctures much larger than eye facets, interspaces between them 1/3–2/3 puncture diameter and smooth. The last ventrite with punctures nearly as large as eye facets, interspaces between them about 2 puncture diameters, alutaceous to relieffy microreticulated at hind edge; rest ventrites with fine sparse and indistinct punctation and cellular, more or less conspicuous microreticulation.

Head slightly convex from above, 3/4 as long as the distance between eyes, with an arched shiny linia at inner edge of each eye. Antennae 3/4 as long as head broad, their flagelli much shorter than the distance between eyes, their club nearly as wide as long, with subtruncate apex and composing about 2/7 of total antennal length. Pronotum with gently sloping and unexplanate sides, its hind corners blunt with a widely rounded apex. Scutellum subhemicircular. Elytra with steeply sloping sides and expressed subsutural lines, but without a trace of humeral stria. Pygidium with widely rounded apex. Prosternal process rather broad, more than twice wider than antennal club, and arcuately widened before unbordered apex. The distance between mid coxae 2 times and that between hind ones almost 3 times more than that between fore coxae. Metasternum flattened or slightly convex, without a trace of medial suture, its fore edge moderately emarginate. Caudal marginal line behind hind coxal cavities follows closely hind edge of coxal cavities. Hypopygidium simple and with a gently curved hind edge.

Fore tibia slightly narrower, mid and hind ones slightly wider than prosternal process; mid and hind tibiae with dense, thin and comparatively long setae. Femora nearly 1.5 times as wide as correspondent tibia. Fore tarsi 2/5 as wide as fore tibiae, but mid and hind ones much narrower, claws with a raised blunt tooth at base.

Ovipositor well sclerotized.

Notes. This new species seems to be closely related to *M. (C.) bisignifer* sp. n. differing from it and other members of *gloriosus* (or *ruficollis*) and *spissus* species-group in the characters given in the below key.

Key to the species of the *gloriosus* (or *ruficollis*) and *spissus* species groups
of the subgenus *Clypeogethes* SCHOLTZ, 1932, from Namibia and adjacent territories

- 1 Elytral punctation without a trace of transrugosity, punctures more or less distinctly oval and diffuse 2
 – Elytral punctation a more or less expressed transrugosity, punctures frequently asymmetric 5
 2 (1) Body larger (2.2–2.6 mm), in general, more convex and more robust; fore tibia almost twice wider than antennal club 3
 – Body smaller than 2.4 mm, less convex and more slender; fore tibia slightly wider than antennal club 4
 3 (2) Entirely reddish, with scarcely lighter appendages; dorsum more or less alutaceous; fore edge of head narrowly and deeply excised, with widely rounded side corners. Male: hypopygidium with an indistinctly isolated, larger and concave plate (with punctures and microreticulation) before the middle of hind edge; tegmen with rather narrowed lateral lobes; penis with a distinctly acute apex. Female: ovipositor with narrower apex and distal projection of gonocoxites behind “central point”. Figs. 94–100. Namibia. *M. (C.) mitis* **sp. n.**
 – Reddish with not infrequently darkened elytra and abdomen: dorsum shiny; fore edge of head angularly and more shallowly, with very narrowly rounded side corners. Male: hypopygidium with a distinctly isolated, larger and concave plate (without punctures and shiny) before the middle of hind edge; tegmen with scarcely narrowed lateral lobes; penis with a projecting but blunt apex. Female: ovipositor with narrower apex and distal projection of gonocoxites behind “central point”. Figs. in EASTON, 1960: 56, 57, 105. Republic of South Africa, Zaire, Uganda, Kenya, Rwanda, Tanzania *M. (C.) ruficollis* REITTER, 1872
 4 (2) Body more oval, more convex and larger (2.3–2.4 mm); reddish; fore edge of head with narrowly rounded side corners; pygidium coarsely microreticulated between clear punctures. Male: hypopygidium with a distinctly isolated, larger and concave plate (without punctures and shiny) before the middle of hind edge; tegmen with a simple angular excision between lateral lobes; penis with a blunt apex. Female: ovipositor with a narrow apex of shorter gonocoxites. Figs. in KIREJTSHUK, 1988: 58–63. Namibia
 *M. (C.) imitans* KIREJTSHUK, 1988, **comb. n.**
 – Body more elongate, slightly convex and smaller (1.8–1.9 mm); dark brown; fore edge of head with widely rounded side corners; pygidium coarsely granulose and without clear punctation. Male: hypopygidium with a slightly expressed plate (with punctures and microreticulation) before the middle of hind edge; tegmen with a deep and narrow excision between lateral lobes; penis with a acute apex. Female: ovipositor with a wide apex of moderately long gonocoxites. Figs. 101–108. Namibia *M. (C.) opacidorsum* **sp. n.**
 5 (1) Dark brown with a large reddish spot on each elytral disk; pronotum subparallelsided at basal half; elytral transrugosity very fine but quite distinct with small punctures behind each furrow (much smaller than on head and pronotum); fore edge of metasternum slightly convex between coxae; prosternal process 1.5 times as wide as antennal club; crenellation of fore tibia more regular along distal half of outer edge; hind femur moderately curved along hind edge, somewhat more than twice wider than antennal club. Male: metasternum and hypopygidium simple. Female: ovipositor with comparatively short gonocoxites. 1.8–2.3 mm. Figs. 76–85. Namibia *M. (C.) bisignifer* **sp. n.**
 – Body more or less unicoloured and lighter (though in *M. (C.) verniceus* comb. n. elytra can be entirely lighter); pronotum narrowed beginning from base; crenellation of fore tibiae irregular along distal half of outer edge; hind femur with almost straight or slightly curved along outer edge 6
 6 (5) Dorsal punctation shallower and finer; pronotum with punctures much less than eye facets and with interspaces 2–3 puncture diameters; elytral transrugosity somewhat reduced with shallow punctures behind each indistinct furrow; prosternal process almost 2.5 times as wide as antennal club; fore edge of metasternum distinctly emarginate between coxae; hind femur with almost straight hind edge, nearly 3 times as wide as antennal club; unicolored reddish. Male: unknown. Female: ovipositor with longer gonocoxites, rather narrow and acute apex bearing long styli, and also with a distinct spicula proximally from “central point”. 2.2 mm. Figs. 67–75. Namibia *M. (C.) asignifer* **sp. n.**

- Dorsal punctation deeper and coarser; pronotum with punctures a little less than eye facets and with interspaces 1–2 puncture diameters; elytral transrugosity well expressed with distinct and deep punctures behind each indistinct furrow; prosternal process only a little wider than antennal club; fore edge of metasternum slightly convex between coxae; hind femur slightly curved along outer edge, a little more than twice wider than antennal club; dark reddish brown, sometimes with slighter elytra. Male: secondary sexual characters in metasternum and hypopygidium unexpressed. Female: ovipositor with shorter gonocoxites, moderately narrow and acute apex bearing short styli, and also without a distinct spicula proximally from “central point”. 2.1–2.5 mm. Figs. 109–110 and in KIREJTSHUK, 1990: 15–20. Namibia. *M. (C.) verniceus* KIREJTSHUK, 1990

***Meligethes (Clypeogethes) bisignifer* KIREJTSHUK, sp. n.** (figs. 76–85)

Specimens examined: Namibia: Holotype, male (TMP) and 5 paratypes (TMP, ZIN) – “Khomas hoch 1, Farm Wissenfels, 23.20 S–16.25 E” (“Khomas H1, Farm Hohenheim, 23.20 S–16.20 E”), “8. 3. 1975, groundtrap, day, Endrödy-Younga” (singled); 1 paratype, female (TMP) – “Abachaus, S. W. A., C. Koch”; 1 paratype, female (MAK) – “Okahandja, 1240 m, 31. 1–3. 2. 1979, leg. H. Roer”.

Description: Male, holotype. Length 2.3, breadth 1.2, height 0.8 mm. Oval, rather convex as from above as from below; bright reddish brown with slight spots on elytra, palpi of mouth parts, hypomera, epipleura, antennae and legs somewhat lighter; dorsum moderately shiny, with sparse, moderately conspicuous, short hairs, nearly as long as the distance between striae on elytra; ventral surface slightly shiny, with more conspicuous and longer pubescence.

Head surface with distinct punctures nearly as large as eye facets, interspaces between them 0.5–1.0 puncture diameter, with a smoothed, cellular, fine microreticulation. Pronotal surface similar to that on head, but punctures somewhat larger and more sparse, and interspaces smooth. Elytral surface with fine and not dense transrugosity, with punctures, about smaller than on head and pronotum and interspaces much more than a puncture diameter, alutaceous or somewhat smoothed. Scutellum with few sparse punctures as large as those on head and with contrast cellular microreticulation. Pygidial surface coarsely granulose and with a scarcely expressed punctation. Surface on prosternal process, metasternum and 1st ventrite with sparser punctures than those on head and pronotum and with smoothed interspaces; remainder of ventrites with small and moderately dense punctures, interspaces between them more or less densely and finely microreticulated.

Head flattened from above, nearly as long as the distance between eyes. Antennae 4/5 as long as head broad, their club nearly 3/4 as wide as long, with suboblique apex and composing about 2/7 of total antennal length. Pronotum with gently sloping and extremely narrowly explanate sides, its hind corners blunt with rounded apex. Scutellum subhemicircular. Elytra with steeply sloping sides and expressed subsutural lines, but without a trace of humeral stria. Pygidium with widely rounded, nearly transverse apex. Prosternal process 1 and 3/4 as wide as antennal club, and arcuately widened before very widely rounded apex with a unbordered hind edge. The distance between mid coxae 2 times and that between hind ones 4 times more than that between fore coxae. Metasternum flattened, its fore edge rather deeply emarginate. Caudal marginal line behind hind coxal cavities follows closely hind edge of coxal cavities. Hypopygidium.

Fore tibia slightly wider, mid and hind ones 1.5 times wider than antennal club; mid and hind tibiae with dense, stout and short setae. Femora nearly 1.5 times almost to twice as wide as correspondent tibia. Tarsi much more than 3 times narrower than correspondent tibiae, claws with a weakly raised tooth at base.

Aedeagus moderately sclerotized.

Female. Secondary sexual characters in structure of legs and metasternum not expressed. Pygidial apex very widely rounded. Hypopygidial apex slightly and simple.

Ovipositor moderately sclerotized.

Variations. Length 1.8–2.3 mm. The paratype collected by H. ROER is with a more contrast pubescence, black dorsum with a bright reddish spot on each elytron, dark brown ventral surface, light antennae and legs.

Notes. This new species is closely related to previous species, but easily diagnosed from all members of *gloriosus* (or *ruficollis*) and *spissus* species-group from the territory under consideration in the characters given in the above key.

***Meligethes (Clypeogethes) imitans* KIREJTSHUK, 1988, comb. n.**

KIREJTSHUK, 1988: Namibia.

Specimens examined: Namibia: 1 (MAK) – “Grootfontein, Farm Mariabroun, 28. I. 1975; leg. H. Roer”; 1 (TMP) – “Abachaus, Otjivarongo, N South-west Africa, XII. 1949, G. Hobohn”.

***Meligethes (Clypeogethes) rugifer* SPORNRAFT & KIREJTSHUK, 1993 (figs. 86–93)**

SPORNRAFT & KIREJTSHUK, 1993: Republic of South Africa.

Specimens examined: Republic of South Africa: 32 specimens (NHL, ZIN) – “Natal, Nqutu, 11. I. 51, Newton, *Haplocarpa scabiosa*” (12. I. 51, 26. I. 51, 7. XII. 51, 25. XII. 51, 31. X. 54, 11. XII. 54, 8. I. 55, 23. XII. 61, “Yellow Compositae Spec. G.”, “Yarrow”, “Vetch”).

***Meligethes (Clypeogethes) mitis* KIREJTSHUK, sp. n. (figs. 94–100)**

Specimens examined: Namibia: Holotype, male (ZSM) and 2 paratypes (ZIN, ZSM) – “Umg. v. Windhuk, I. 1936–XII. 1937, leg. K. Hartmann, Mus. Frey Tutzing”.

Description. Male, holotype. Length 2.6, breadth 1.6, height 1.0 mm. Oval, rather convex as from below; unicoloured bright reddish, palpi of mouth parts, hypomera, epipleura, antennae and legs somewhat lighter; dorsum nearly dull, with sparse, hardly conspicuous, very short hairs; ventral surface slightly shiny, with more conspicuous and longer pubescence.

Head surface with oval punctures nearly as large as eye facets, interspaces between them 0.5–1.0 puncture diameter, with distinct, cellular, dense and fine microreticulation. Pronotal and elytral surface as that on head, but punctures at base of pronotum and at sides coarser and interspaces between them more than a puncture diameter, densely microreticulated and somewhat smoothed on pronotal disk. Scutellum with sparse punctures as large as those on head and with contrast cellular microreticulation. Pygidial surface almost finely granulose with scarcely expressed punctation. Surface on prosternal process, metasternum and 1st ventrite with shallower and sparser punctures than those on dorsum, remainder of ventral surface as that on pygidium though with more expressed shallow punctures.

Head flattened from above, nearly as long as the distance between eyes. Antennae 4/5 as long as head broad, their flagelli shorter than the distance between eyes, their club nearly 3/4 as wide as long, with suboblique apex and composing about 2/7 of total antennal length. Pronotum with gently sloping and unexplanate sides, its hind corners blunt with rounded apex. Scutellum subhemicircular. Elytra with steeply sloping sides and expressed subsutural lines, but without a trace of humeral stria. Pygidium with widely rounded, nearly transverse apex. Prosternal process 1 and 3/4 as wide as antennal club, and arcuately widened before almost hemicircular apex with a transverse unbordered hind edge. The distance between mid coxae 2 times and that between hind ones 3 times more than that between fore coxae. Metasternum flattened with a medial suture traced at distal half, its fore edge rather deeply emarginate. Caudal marginal line behind hind coxal cavities follows closely hind edge of coxal cavities. Hypopygidium with a large depression before the middle of transverse hind edge, which has unbordered margin.

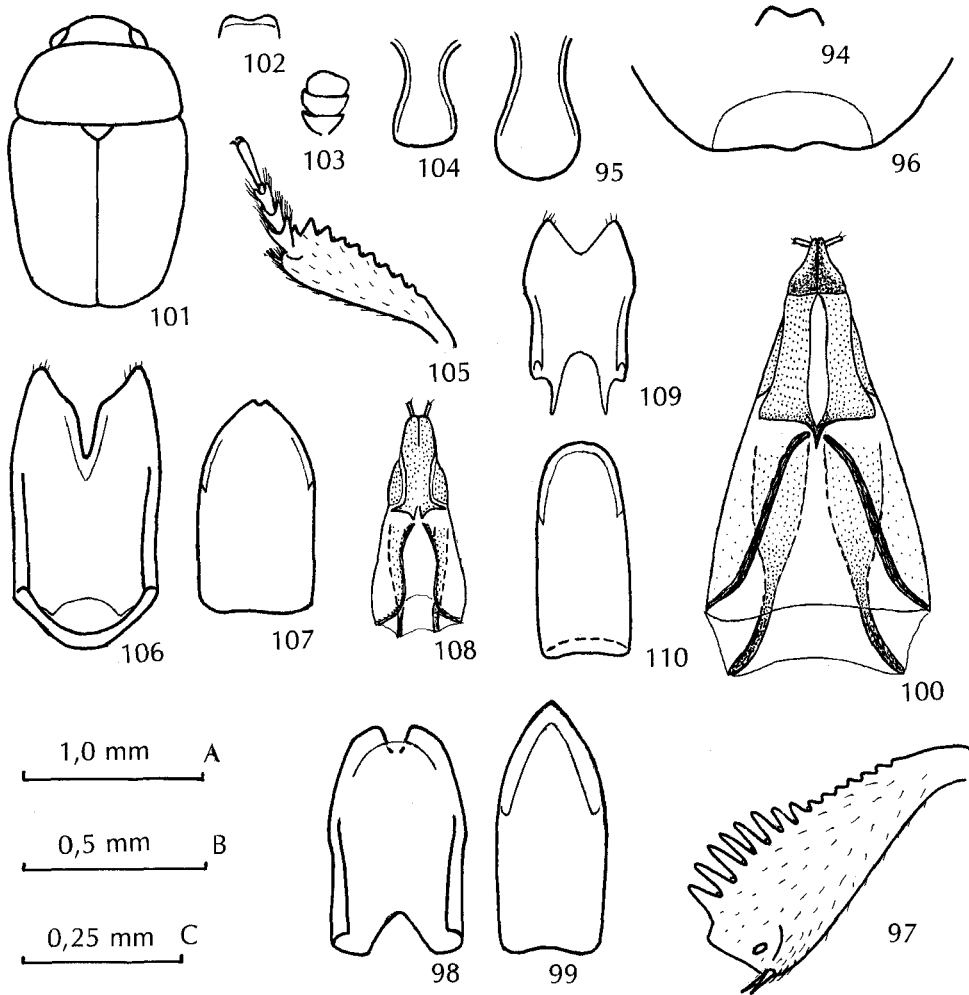
Fore tibia slightly narrower, mid and hind ones slightly wider than prosternal process; mid and hind tibiae with dense, stout and short setae. Femora nearly 1.5 times as wide as correspondent tibia. Fore tarsi missing, but mid and hind ones more than 3 times narrower than correspondent tibiae, claws with a raised tooth at base.

Aedeagus well sclerotized.

Female. Differs from the male in convex metasternum and widely rounded apex of hypopygidium. Ovipositor moderately sclerotized.

Variations. Length 2.3–2.6 mm. A little variability is manifested in configuration of outer edge of fore tibiae, punctation and microreticulation.

Notes. This new species together with some other species considered below in these notes are characterized by a deeply emarginate fore edge of metasternum (feature unusual for the genus in general). It is similar and related to *M. (C.) ruficollis* REITTER, 1872, although it can be easily distinguished from the latter by more unicoloured and nearly dull body, fore edge of head narrowly excised and with widely rounded side corners, indistinctly isolated, larger and concave plate (with punctures and microreticulation) on male hypopygidium, genital structures (in particular shape of tegmenal lateral lobes in males and sclerites of gonocoxites in females). Besides it, *M. (C.) mitis* sp. n., perhaps, is closely related to *M. (C.) imitans* KIREJTSHUK, 1988, but distinct from it in larger body with dull surface, indistinctly isolated, larger



Figs. 94–110. Genus *Meligethes*, subgenus *Clypeogethes* (orig.). – 94–100 *M. (C.) mitis* sp. n.; 94 fore edge of head, dorsal; 95 prosternal process, ventral; 96 male hypopygidium, ventral; 97 male fore tibia, dorsal; 98 tegmen, ventral; 99 penis trunk, dorsal; 100 ovipositor, ventral; 101–108 *M. (C.) opacitorsum* sp. n.; 101 body shape, dorsal; 102 fore edge of head, dorsal; 103 antennal club; 104 prosternal process, ventral; 105 male fore tibia and tarsus, dorsal; 106 tegmen, ventral; 107 penis trunk, dorsal; 108 ovipositor, ventral; 109–110 *M. (C.) verniceus* comb. n.; 109 tegmen, ventral; 110 penis trunk, dorsal; Scales: A – to fig. 101; B – to figs. 94, 95–96, 102, 104; C – to figs. 97–100, 105–110.

and concave plate (with punctures and microreticulation) on male hypopygidium, narrower excision between lateral lobes of tegmen and strongly acute apex of ovipositor.

***Meligethes (Clypeogethes) opacitorsum* KIREJTSHUK, sp. n.** (figs. 101–108)

Specimens examined: Namibia: Holotype, male (MAK) and 4 paratypes (MAK, ZIN) – “Okahandja, 1240 m, 31. 1–3. 2. 1979, leg. H. Roer”; 1 paratype, female (MAK) – ibid . . . “14. 3. 1979, H. Roer”.

Description. Male, holotype. Length 1.8, breadth 1.1, height 0.6 mm. Moderately convex as from above as from below; dorsum unicoloured dark reddish brown, ventral surface and appendages somewhat

lighter – chestnut brown; dorsum nearly dull, with sparse, hardly conspicuous, very short hairs; ventral surface slightly shiny, with more conspicuous and longer pubescence.

Head surface with oval punctures nearly as large as eye facets, interspaces between them about one puncture diameter, with distinct, cellular, dense and fine microreticulation. Pronotal and elytral surface similar to that on head, but punctures on pronotum as those on head and ones on elytra almost twice larger, and interspaces more than 2–3 puncture diameters, with dense and conspicuous cellular microreticulation, and partly smoothed on elytra. Scutellum with sparse punctures as large as those on head and with contrast cellular microreticulation. Pygidial surface coarsely granulose and without expressed punctation. Ventral surface on prosternal process, metasternum and 1st ventrite with shallower and sparser punctures than those on dorsum, interspaces smoothed on prosternal process and metasternum, but with more or less expressed microreticulation of ventrites.

Head slightly convex, nearly as long as the distance between eyes. Antennae 4/5 as long as head broad, their club nearly 3/4 as wide as long, with suboblique apex and composing about 2/7 of total antennal length. Pronotum with gently sloping and unexplanate sides, its hind corners blunt with rounded apex. Scutellum subhemicircular. Elytra with steeply sloping sides, expressed subsutural lines, but without a trace of humeral stria. Pygidium with widely rounded, nearly transverse apex. Prosternal process more than twice as wide as antennal club, and arcuately widened before almost hemicircular apex with a transverse unbordered hind edge. The distance between mid coxae 2 times and that between hind ones 3 times more than that between fore coxae. Metasternum flattened, its fore edge rather deeply emarginate. Caudal marginal line behind hind coxal cavities follows closely hind edge of coxal cavities. Hypopygidium with a transverse hind edge.

Fore tibia slightly wider, mid and hind ones nearly twice wider than antennal club; mid and hind tibiae with extremely dense, stout and long setae. Femora nearly 1.3–1.5 times as wide as correspondent tibia. For tarsi twice, but mid and hind ones more than 3 times narrower than correspondent tibiae, claws with a weak tooth at base.

Aedeagus moderately sclerotized.

Female. Differs in rounded apices of pygidium and hypopygidium.

Ovipositor slightly sclerotized.

Variations. Length 1.8–1.9 mm. A little variability is manifested in configuration of outer edge of fore tibiae, punctation and microreticulation. The female collected on 14. 3. 1979 has somewhat smoothed dorsal surface, but microreticulation of this specimen is quite dense, though punctures are indistinct.

Notes. This new species seems to be closely related to *M. (C.) bisignifer* sp. n. differing from it and other members of *gloriosus* (or *ruficollis*) and *spissus* species-groups in the characters given in the above key.

Meligethes (Clypeogethes) verniceus KIREJTSHUK, 1990, **comb. n.** (figs. 109–110)

(= *Meligethes verniceus* KIREJTSHUK, 1990).

KIREJTSHUK, 1990: Namibia.

Specimens examined: Namibia: 2 (MNHUB, ZIN) – “Windhuk, Swakopmund, Lindt S. G.”; 3 (NHL, ZIN) – “S. W. Africa (13) Barby Farm, 25 mls, W. Helmeringhausen, 17–18. I. 1972, at light”; 1 (MAK) – “Naos/Rehoboth, 9–15. 3. 1985, H. Roer”; 1 (TMP) – “S. W. Afr., Windhoek, Regenstein, 22.36S – 16.59E”, “7. 3. 1975; E-Y: grassnetting, leg. Endrödy-Younga”.

Notes. This species was described after the study only females. The males has somewhat wider fore tarsi, unexpressed secondary sexual characters in metasternum and hypopygidium, and moderately sclerotized aedeagus.

It seems to be closely related to *M. (C.) bisignifer* sp. n. differing from it and other members of *gloriosus* (or *ruficollis*) and *spissus* species-groups in the characters given in the above key.

Nitidulinae

Anachramus distinctus (GROUVELLE, 1899)

(= *Soronia distincta* GROUVELLE, 1899; *Anachramus distinctus*: KIREJTSHUK, 1995).

KIREJTSHUK, 1995: Republic of South Africa.

Specimens examined: Namibia: 1 (MNHUB) – “Lüderitz, 22. X. 1991, leg. U. Göllner”.

Notes. This species is described from Bushmenland (NW Namibia) and therefore the occurrence of it on the territory under consideration is quite expectable. Bionomy of this species remains unknown.

***Taracta varia* (GROUVELLE, 1898)**

(= *Platychora varia* GROUVELLE, 1898; *Taracta varia*: KIREITSHUK, 1988).

GROUVELLE, 1898: Zaire (“Congo”).

Specimens examined: Namibia: 5 (MAK, ZIN) – “Tordoro/Okahango, 14–19. I. 1975, leg. H. Roer” (23. 3. 1975); Angola: 22 (MAT, ZIN) – “Dungo Ang., 2074.9, VII-1953, à la lumière, E. Luna de Carvalho” (3330.13, XI. 1953); 4 (MAT) – “3115-3, Dundo, 7.22S. 20.50E, à la lumière, 10/13. VIII. 1953, Luna de Carvalho” (3176-14, 8. IX. 1953); some dozens specimens from Ivory Coast (MAT, ZIN), Angola (MAT, ZIN), Tchad (MAT, ZIN), Sudan (TMB), Republic of Central Africa (MAT), Zaire (MAT, ZIN), Tanzania (MAT, ZIN).

Notes. This species is closely related to *Taracta fryil* MURRAY, 1867 which is more widely distributed than the typical locality (“Old Calabar”), at least in Ivory Coast (MAT), Zaire (MAT), Ethiopie (ZIN). These relatives are reliably distinguishable only due to structure of their male genitalia. Bionomy of this form is unknown, but any association of it with subcortical life is quite probable.

***Stelidota didyma* REITTER, 1875**

(= *Stelidota remillei* ENDRÖDI-YOUNGA, 1982, syn. n.).

REITTER, 1875; ENDRÖDI-YOUNGA, 1982: Madagascar.

Specimens examined: Angola: 1 (ZIN) – “(A 11) Bruco, 26. ii–2. iii. 1972”, “Damp leaf litter by stream”; 1 (NHL) – “(A 21) Chianga, 6. iii. 1972”, “Cut grass heaps”; Madagascar: Lectotype, female (MNHUB), here designated (studied and designated in collection by S. Endrödy-Younga) – “Madagascar, Goudot”, “didyma Kl., Madagasc., Goud.”; 1 male (ZIN) – “N. suffusa Kl. Madagascar”.

Notes. The lectotype has length – 3.0, breadth 1.7 mm, brownish with slightly visible 3 yellowish spots on each elytra. The studied male from Madagascar (ZIN) is originated from the same series than the lectotype, strongly similar to it and should be recognised as conspecific. The latter has genital structure and other characters (including secondary sexual ones) are correspondent to features indicated in the description of *S. remillei*, and therefore the both names are obvious synonyms.

2 specimens (males) from Angola available for study look like immature, but have secondary sexual character and aedeagal structure as those in the Madagascarean representatives of *S. didyma*, though they slightly smaller (2.5 mm) and unicoloured bright reddish body without spots on elytra.

Bionomy of this species, perhaps, as that in other species of the genus, scoping many types of habits in litter and near soil with decomposed plant substrates.

***Lordites (Lordites) costipennis* (BOHEMAN, 1851), comb. n.**

(= *Soronia costipennis* BOHEMAN, 1851; *Soronia exaratus* BOHEMAN, 1851; syn. n.; *Lordites brevisculus* FAIRMAIRE, 1868, syn. n.; *Lasiodyctylus subproductus* REITTER, syn. n.).

BOHEMAN, 1851; Republic of South Africa; GROUVELLE, 1913: also Guinea, Madagascar; DELOBEL, TRAN, 1993 – Central and East Africa.

Specimens examined: Namibia: 1 (MAK) – “Windhoek, 8. 2. 75, leg. H. Roer, Lichtfang”; 1 (ZIN) – “Nyangana, Okavanga, 14.–22. 01. 1985”; 1 (MAK) – “Gobabeb/Kuiseb 23.3 S 150 E, 406 m, 20. 2.–6. 3. 1979 H. Roer”; 2 (MNHUB) – “Osona, bei Okahandja, p. III–m. IV. J. Irish”; 3 (MNHUB, ZIN) – “Grootfontein: Askavolt-Farm, 20 km E Otavi, 19°40’S/17°33’E, 18. II. 92, leg. M. Uhlig”; 3 (MNHUB, ZIN, SMWN) – “Kavango: Kaudom-Camp, 18°31’S/20°43’E, lux, 22–25. II. 92, leg. M. Uhlig”; 2 (MNHUB) – “Kavango: Popa Falls, 18°07’S/21°35’E, 26. II–3. III. 92, leg. M. Uhlig”; 1 (MNHUB) – “E. Caprivi: 30 km SE Katimo Mulilo, 17°31’S/24°25’E, Zambezi-Altwasserarm, lux, 6. III. 92, lux, leg. M. Uhlig”; 3 (MNHUB) – “East Caprivi: Mudumu NP, Nakatwa, 18°10’S/23°26’E, 8–13. III. 92, lux, leg. M. Uhlig”; 1 (ZSM) – “Okagandya, 29. 1. 1956, F. Gaerdes”; Republic of South Africa: Lectotype *S. costipennis*, male (NRS) here designated (but designated in collection by S. Endrödy-Younga) – “Caffraria J. Wahlb.”, “*Soronia costipennis* Boh.”; Lectotype *S. exarata*, female (NRS) here designated (but designated in collection by S. Endrödy-Younga) – “Caffraria J. Wahlb.”, “*Soronia exarata* Boh.”; 2 (RNL) – “Johannesburg, Z. A., 20. 10. 1960, van Doesburg”; Angola: 10 (MAT, ZIN)

– “Dundo Ang., 16-IX-1953, E. Luno de Carvalho” (8. IX. 1953); 1 (MAT) – “Dundo Ang., 4-IX-1948, A. de Barros Machado”; 3 (MAT, ZIN) – “Dundo, 7.22S, 20.50E, Sources Dundundo detr. sol. 16. III. 1954, A. B. Machado”; Rhodesia: 1 (CMO) – “28°E 18S, Hostes Nicolle Inst. Wildlife Res., Jan 1974, M. B. Fenton”; 2 (CMO, ZIN) – “Atlantica, 16 mi W Salisbury, Jan/74, M. B. Fenton”; 2 (CMO, ZIN) – “Zimbabwe, Sengwa Wildlife Res. Area, 12. X–12. XI. 79, Gary P. Bell, lighth”; about a hundred specimens from Namibia: (MAK, MNHUB, ZIN), R.S.A. (MNHUB, ZIN) and some thousand specimens from Somalia (MAT, MCG); Mauritanie (MAT, ZIN), Senegal (MAT, ZIN), Guinea (MNHUB, TMB, ZIN), Liberia (RNL, SMS, ZIN), Ivory Coast (MAT, MNHUB, SMS, ZIN), Ghana (MAT), Togo (MNHUB, SMS, ZIN), Benin (SMS), Nigeria (SMS, ZSM), Cameroon (DEI-subproductus, MAT, MNHUB, TMB, ZIN), Equatorial Guinea (MNHUB), Congo (MAT), Zaire (MAT, MNHUB, TMB, ZIN), Upper Volta (MAT, ZIN), Tschad (MAT, ZIN), Sudan (MNHUB, TMB, ZIN), Ethiopie (MAT, SMS, TMB, ZIN, ZMM), Kenia (MNHUB), Uganda (MAT, ZIN), Tanzania (MAT, MNHUB, NMW, TMB, ZIN), Rwanda (MAT), Burundi (MAT, ZIN), Mali (MAT), Madagascar (MAT, ZIN, ZSM), Réunion (DEI-breviusculus).

Notes. A. DELOBEL and M. TRAN, 1933 used the synonymy *L. costipennis* = *subproductus*. Some specimens from the collection of Eberswalde (DEI) determined by A. GROUVELLE and S. ENDRÖDY-YOUNGA as *Lasiodactylus brevisculus* and *L. subproductus* as well as description of these names are completely correspondent to characters of the species named here as *Lordites (Lordites) costipennis* because this specific name should be recognised as the oldest synonym.

Bionomy of this species, perhaps, as that in other species of the genus, and live in both natural and artificial conditions, however, it is more common on decaying and dry fruits, wet decomposed plant substrates, including leaf litter.

Lordites (Lordites) tibialis (BOHEMAN, 1851), **comb. n.**

(= *Soronia tibialis* BOHEMAN, 1851; *Soronia caffer* BOHEMAN, 1851, syn. n.; *Lordites maculipennis* KRAATZ, 1895, syn. n.; *Lordites curvitiibus* KRAATZ, 1895, syn. n.; ? *Nitidula maculata* PALISOT, 1811; ? *Lordites inquinatus* ERICHSON, 1843; ? *Lasiodactylus elongatus* REITTER, 1873; ? *Lasiodactylus substriatus* REITTER, 1873; BOHEMAN, 1851: Republic of South Africa; GROUVELLE, 1913: also Togo; BORGES, 1990: Azores, Madeira, Afrotropical region; DELOBEL & TRAN, 1993: Congo.

Specimens examined: Namibia: 10 (MAK) – “Okahandja, 13–15. 12. 1974, leg. H. Roer” (31. 1–3. 2. 1979, 8–12. 3. 1979); 11 (MAK, ZIN) – “Tondoro/Okavango, 14–19. I. 1975, leg. H. Roer, Lichtfang” (23. I. 1975); 2 (MAK) – “Farm Mariabroun, 28. I. 1975, leg. H. Roer”; 2 (MAK, ZIN) – “Andara/Okavango, 24–26. I. 1975, leg. H. Roer, Lichtfang”; 16 (MAK, ZIN) – “Windhoek, 8. 2. 75, leg. H. Roer, Lichtfang” (2. 3. 75, 7. 2. 75); 1 (MAK) – “Gobabeb/Kuiseb, 23.3S-150E, 406 m, 20. 02–6. 03. 1979, H. Roer”; 3 (MAK, ZIN) – “Nyangana/Okavango, 14–22. I. 1985, leg. H. Roer”; 4 (MAK, ZIN) – “Nyangana/Raum Grootfontein, 23–31. I. 1982, leg. H. Roer”; 9 (MAK, ZIN) – “Okahandja, 1240 m, 17–20. 03. 1982, leg. H. Roer” (31. 1–3. 2. 1979, 18–21. 3. 1985); 5 (MAK, ZIN) – “Okahandja, Swakoprivier, 18–21. 3. 1985, leg. H. Roer”; 2 (MAK) – “Naos Bez, Rehoboth, Khomas-Hochland, 5–10. 2. 1982, H. Roer”; 1 (MNHUB) – “E. Caprivi: Katimo Mulilo, 17°29'S/24°17'E., 3–8. III. 92, lux, leg. M. Uhlig”; 1 (ZSM) – “Okagandya, 29. 1. 1956, F. Gaerdes”; Angola: 1 (MAT) – “Lunda: 8.00S, 20.55E, Route Dundo-Sombo, detr. sol., 21. IX. 53, Luno Carvalho”; 4 (MAT, ZIN) – “Dundo Ang., 3-XI-1953, à la lumiere, E. Luna de Carvalho”; 1 (MAT) – “Dundo Ang., 4. III. 1948, A. de Barros Machado”; Zimbabwe: 4 (CMO, ZIN) – “Atlantica, 16 mi W Salisbury, Jan/74, M. B. Fenton”; Republic of South Africa: Lectotype *S. tibialis*, male (NRS), here designated (but designated in collection by S. Endrödy-Younga) – “Caffraria, J. Wahlb.”, “*Soronia tibialis*”; Lectotype *S. caffer*, male (NRS), here designated (but designated in collection by S. Endrödy-Younga) – “Caffraria, J. Wahlb.”, *caffra*; Lectotype *L. maculipennis*, male (DEI), here designated (but designated in collection by S. Endrödy-Younga) – “Togo”, “*Lordites maculipennis* KRTZ 95”; Lectotype *L. curvitiibus*, male (DEI), here designated (but designated in collection by S. Endrödy-Younga) – “Togo”, “*Lordites curoitibus* KRTZ 95” and 1 paralectotype (DEI) – “Togo, Conradt”; 1 (MNHUB) – “*Lordites substriatus*, Coll. L. W. Schaufuss” (with a green and a black quadrangles); 1 (TMB) – “N Transvaal, Louis Trichardt, UV-Licht/falle, XII. 1961, F. Neubecker”; 28 (TMB, ZIN) – “Transvaal, Pretoria, on light, 28-30. XII. 1977, Leg. Dr. S. Endrödy” (30. I. 1978, 10–25. XI. 1980, 6, 9, 11, 14, 22, 24. XII. 1980); 4 (TMB, ZIN) – “Transvaal, Nelspruit, singled, 8. I. 1980, Leg. Dr. S. Endrödy”; 2 (RNL) – “Sunnyside, Pretoria, 2. I. 1959, A. C. v. Bruggen”; 1 (RNL) – “Johanesburg, Z. A., 21. 10. 1960, van Doesburg”; 2 (CMO) – “Natal, 75 WSW Estcourt Cathedral Peaks For. Sta, S. & J. Peck, below rotten fruits, 17. XII. 1979”; and some

thousands specimens including named by A. GROUVELLE as *L. curvitiubius*, *L. maculipennis*, *L. substriatus* from Ethiopie (MAT, SMS, ZIN, ZMM), Guinea (MAT, MNHUB, TMB, ZIN), Togo (DEI, MAT, MNHUB, SMS, ZIN), Nigeria (SMS, ZIN), Cameroon (MAT, MNHUB, TMB, ZIN, ZSM), Ivory Coast (CMO, MAT, ZIN), Uganda (ZMH), Ghana (MAT, ZIN), Tanzania (MAT, MNHUB, NMW, TMB, ZIN), Kenya (MCG, MNHUB, ZIN), Equatorial Guinea (MNHUB, ZIN), including Sao Tomé – TMB), Congo (MAT, TMB), Zaire (MAT – including with label “Las inquitatus var. ou sp. n. det. Grouvelle”, ZIN, ZSM), Zambia (MNHUB), RSA (MNHUB, ZSM), Liberia (RNL), Burundi (MAT, ZIN), Rwanda (MAT, ZIN), Malawi (MAT), Mali (MAT), Uganda (MAT), Madagascar (MAT, ZIN).

Notes. P. A. V. BORGES (1990), and also A. DELOBEL and M. TRAN (1993) have published the synonymy of *L. tibialis* = *maculipennis* = *curvitiubius* with reference to paper of this publication. The synonymy proposed above are based on comparison of type series with taking into consideration of scope of variability of this species which is one of most common in Afrotropical and Capean regions. Moreover, some probable synonyms are listed with sign “?” till retesting of the type series used for proposition of these names.

Bionomy of this species, perhaps, as that in other species of the genus, and live in both natural and artificial conditions, however, it is more common on decaying and dry fruits, wet decomposed plant substrates, including leaf litter. This species is very abundant in fruit remains (banana and so on).

Aethina (Circopes) peringueyi GROUVELLE, 1908/1909, **comb. n.**

(= *Circopes peringueyi* GROUVELLE, 1908/1909).

GROUVELLE, 1908/1909: Republic of South Africa; AUDISIO, 1982: Sierra Leone (tropical and austral Africa).

Specimens examined: Namibia: 7 (MNHUB) – “Distr. Grootfontain, I. Irish”, “Farm Hurisib, 18°23’S/17°55’E, Anfang IV. 1989”; 3 (MNHUB) – “Farm b. Omaruru, 5–7. X. 1991, leg. U. Göllner”; 2 (MNHUB, ZIN) – “b. Grootfontain, Farm Hurisib, 8–9. X. 1991, leg. U. Göllner”; 1 (MNHUB) – “Osona b. Okahanja, 19–20. X. 1991, leg. U. Göllner”; 5 (MNHUB, ZIN, SMWN) – “Namibia-Exp. ZMB 1992, Kavango: Kaudom-Camp, 18°31’S/20°43’E, 22–25. II. 92, leg. U. Göllner”; 7 (MNHUB, ZIN) – “Kavango: Popa Falls, 18°07’S/21°35’E, 26. II–3. III. 92, leg. M. Uhlig” (U. Göllner); 2 (MNHUB) – *ibid.* “Buffalo-Camp, 18°09’S/21°42’E, 28. II–3. III. 92, leg. M. Uhlig” (U. Göllner); 1 (MNHUB) – *ibid.* “Gelukkies, Kavango-Ufer, 18°03’S/21°08’E, 1. III. 92, leg. U. Göllner”; 1 (MNHUB) – “E. Caprivi: Katimo Mulilo, 17°29’S/24°17’E, 3–8. III. 92, lux, leg. M. Uhlig” (U. Göllner); 5 (MNHUB, ZIN) – *ibid.* “Mudumu NP, Nakatwa, 18°10’S/23°26’E, 8–13. III. 92, U. Göllner”R; 1 (MNHUB) – “Grootfontein”; 1 (MNHUB) – “Omaruru: 30 km NON Omaruru Farm Otjua, 21°7’S/16°804E, 17. III. 92, leg. U. Göllner”; 3 (MAK, ZIN) – “Tondororo/Okavango, 14. I. 1975, H. Roer” (20–23. I. 1975); 1 (MAK) – Grootfontein, Farm Mariabroun, 1. 2. 1975, H. Roer”; 8 (MAK, ZIN) – “Nyangana/Okavango, 14–22. I. 1985, H. Roer” (1–9. 4. 1988); 1 (SMS) – “Gobabis S. W. Afrika, 1933, Dr. Maag”; 1 (SMS) – “30. 9–4. 10. 1990, P. Schüle, Bagane”; Republic of South Africa: Lectotype *Circopes peringueyi*, male (MNP), here designated (designated in collection by S. Endrödy-Younga) and 1 paralectotype, female (MNP) – “Transvaal, Hartmann”; 2 (MNHUB, ZIN) – “Bushmanland, Klein Dobe, 19°25’S/20°21’E, 19–21. II. 92, leg. M. Uhlig” (U. Göllner); 1 (TMB) – “Transvaal, Hylsvley, cow dung trap, 3. II. 1978, Dr. S. Endrödi”; 1 (ZIN) – “Transvaal, Bergvliet Gorge, 15 km E from Sabia, netted, 4. XI. 1980, Dr. S. Endrödi”.

Notes. This species has an intermediate position between *Circopes* REITTER, 1873 and *Ithyra* REITTER, 1873, but well distinguished by tarsal claws strongly toothed at base and very dark to blackish coloration with frequently dark legs. This species seems to be anthophagous.

Aethina (Ithyra) hirsutala REITTER, 1873, **comb. n.**

(= *Ithyra hirsutala* REITTER, 1873)

AUDISIO & KIREJTSCHUK, 1983: Sicily, Jemen, Sudan, Ethiopia, Somalia, Sierra Leone, Ivory Coast, Zaire, Kenia, Uganda, Ruanda, Tanzania, RSA.

Specimens examined: Namibia: 1 (MNHUB) – “Namibia-Exp. ZMB 1992, East Caprivi: Mudumu NP, Nakatwa, 18°10’S/23°26’E, 8–13. III. 92, U. Göllner”; 1 (MNHUB) – “Namibia-Exp. ZMB 1992, Omaruru: 30 km NON Omaruru Farm Otjua, 21°7’S/16°4’E, 17. III. 92, leg. U. Göllner”; 8 (MAK –

“Okahandja, 1240 m, 8–12. 3. 1979, H. Roer”; 48 (MAK, ZIN) – *ibid.* . . “Okahandja, 1240 m, 31. 1–3. 2. 1979, H. Roer”; 1 (MNHUB) – “Distr. Grootfontein, leg. J. Irish (O. G.)”, “Farm Hurisib. 19°23’S/17°55’E, Anfang IV. 1989”; Botswana: 1 (NHL) – “(B22), R. Semowane, 20°25’S 26°23’E, 23–24. IV. 1972, general sweeping”; and also some hundreds specimens from Yemen (NHL, ZIN), Cameroon (TMB, ZIN), Zaire (MAT, ZIN), Ethiopie (MCG, ZIN), Uganda (NHL, ZIN, ZMK), Kenya (MAT, MCG, ZIN, ZMK), Rwanda (MAT, ZIN), Tanzania (MAT, MNHUB, TMB, ZIN, ZMK).

Notes. The taxon *Ithyra* REITTER, 1873 should be changed because the only difference of it from *Circopes* REITTER, 1873 is splitted tarsal claws. The „peculiar” structure of ovipositor of it has been found in some species of *Circopes* remaining still undescribed. Comments on bionomy of this species – see in P. AUDISIO & A. G. KIREJTSHUK, 1983.

Anister hintoni JELINEK, 1981b

JELINEK, 1981b: Zambia, Zimbabwe.

Specimen examined: Namibia: 1 (MNHUB) – “Kavango: Buffalo-Camp, 18°09’S/21°42’E, 28. II. 92, leg. U. Göllner”; Angola: 1 (NHL) – “Luanda, G. R. Gradwell & D. Snow”; and also a dozen from Zaire (MAT, ZIN), Rwanda (MAT).

Notes. This specimens is smaller (2.5 mm in length) than the range mentioned by J. JELINEK (1981 in original description 2.6–3.0 mm) and its fore tibiae only with two distinct subapical teeth (as in *Anister raffrai* GROUVELLE, 1901). As another congeners, this species seems to be associated with the Brassicaceae.

Monafricus major (GROUVELLE, 1899)

(= *Tricanus major* GROUVELLE, 1899; *Monafricus major*: KIREJTSHUK, 1995).

Kirejtshuk, 1995: Cameroon, Zaire, Namibia.

Specimen examined. Namibia: 2 (MNHUB) – “Windhuk, S. V. Krause”. 1 (ZMS) – “S. Afr., N. Transvaal, Nylsvley hill base, 24.40S-28.42E”, “24. 4. 1979, E-Y., 1143, groundtraps day, leg. Breytenbach”, “groundtrap, banana bait, replication 2”.

Notes. Systematic placement and distribution are discussed in detail in KIREJTSHUK, 1995.

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Author's address: Dr. ALEXANDER G. KIREJTSHUK, Zoological Institute of the Russian Academy of Sciences, Sanct-Petersburg, Universitetskaya emb. 1, 199034, Russia.